

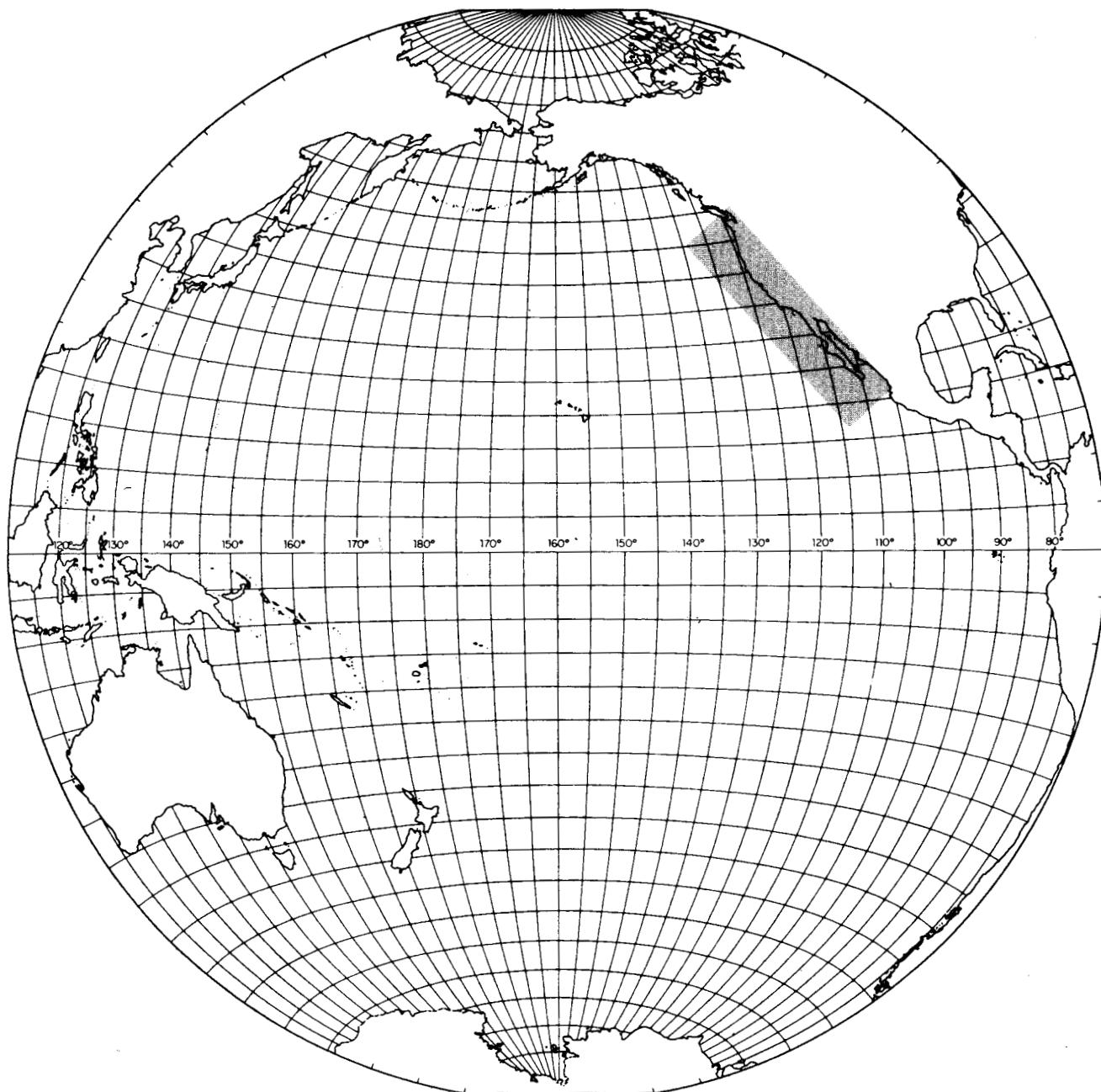


Atlas 35: Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, W. Watson, S. R. Charter, and E. M. Sandknop. Distributional atlas of fish larvae and eggs from Manta (surface) samples collected on CalCOFI surveys from 1977 to 2000. Published May 2002.

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References to the data, published in annual ichthyoplankton data reports are given in the introduction to the Atlas. In addition, these data are available in PDF format on the SWFSC web site at <http://swfsc.noaa.gov/publications/swcpub/qryPublications.asp>, enter "ichthyoplankton" in the Subject line and "California Cooperative Oceanic Fisheries Investigations" in the Title line. Checking the ALL YEARS button will produce the entire list of available data.

The report for each year usually is published about 7-9 months after the fall cruise, and includes notes about nomenclature changes, etc. The ultimate goal is to update the old ichthyoplankton identifications to current standards; the database is updated as re-identifications for each cruise are completed.



CALIFORNIA COOPERATIVE OCEANIC FISHERIES INVESTIGATIONS

ATLAS No. 35

CALIFORNIA
COOPERATIVE
OCEANIC
FISHERIES
INVESTIGATIONS

Atlas No. 35

Cooperating Agencies:

CALIFORNIA DEPARTMENT OF FISH AND GAME
NATIONAL OCEANOGRAPHIC AND ATMOSPHERIC ADMINISTRATION, NATIONAL MARINE FISHERIES SERVICE
UNIVERSITY OF CALIFORNIA, SCRIPPS INSTITUTION OF OCEANOGRAPHY

MAY, 2002

THE CALCOFI ATLAS SERIES

This is the thirty-fifth in a series of atlases containing data on the hydrography and plankton from the region of the California Current. The field work was carried out by the California Cooperative Oceanic Fisheries Investigations¹, a program sponsored by the following agencies:

California Department of Fish and Game

National Oceanic and Atmospheric Administration, National Marine Fisheries Service

University of California, Scripps Institution of Oceanography

CalCOFI atlases² are issued as individual units as they become available. They provide processed physical, chemical, and biological measurements of the California Current region. Each number may contain one or more contributions. A general description of the CalCOFI program with its objectives appears in the preface of Atlas No. 2.

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CalCOFI atlases in this series, through May 2000 are:

- No. 1. Anonymous. 1963. CalCOFI atlas of 10-meter temperatures and salinities 1949–1959.
- No. 2. Fleminger, A. 1964. Distributional atlas of calanoid copepods in the California Current region, Part I.
- No. 3. Alvariño, A. 1965. Distributional atlas of Chaetognatha in the California Current region.
- No. 4. Wyllie, J. G. 1966. Geostrophic flow of the California Current at the surface and at 200 meters.
- No. 5. Brinton, E. 1967. Distributional atlas of Euphausiacea (Crustacea) in the California Current region, Part I.
- No. 6. McGowan, J. A. 1967. Distributional atlas of pelagic molluscs in the California Current region.
- No. 7. Fleminger, A. 1967. Distributional atlas of calanoid copepods in the California Current region, Part II.
- No. 8. Berner, L. D. 1967. Distributional atlas of Thaliacea in the California Current region.
- No. 9. Kramer, D. and E. H. Ahlstrom. 1968. Distributional atlas of fish larvae in the California Current region: northern anchovy, *Engraulis mordax* (Girard), 1951–1965.
- No. 10. Isaacs, J. D., A. Fleminger, and J. K. Miller. 1969. Distributional atlas of zooplankton biomass in the California Current region: Spring and Fall, 1955–1959.
- No. 11. Ahlstrom, E. H. 1969. Distributional atlas of fish larvae in the California Current region: jack mackerel, *Trachurus symmetricus*, and Pacific hake, *Merluccius productus*, 1951–1959.
- No. 12. Kramer, D. 1970. Distributional atlas of fish eggs and larvae in the California Current region: Pacific sardine, *Sardinops caerulea* (Girard), 1951–1966.
- No. 13. Smith, P. E. 1971. Distributional atlas of zooplankton volume in the California Current region, 1951–1966.
- No. 14. Isaacs, J. D., A. Fleminger, and J. K. Miller. 1971. Distributional atlas of zooplankton biomass in the California Current region: Winter, 1955–1959.

¹Usually abbreviated CalCOFI, sometimes CALCOFI

²For citation this issue in the series should be referred to as CalCOFI Atlas No. 35. Library of Congress Catalog Card Number 67-4236.

- No. 15. Wyllie, J. G. and R. J. Lynn. 1971. Distribution of temperature and salinity at 10 meters, 1960–1969 and mean temperature, salinity and oxygen at 150 meters, 1950–1968 in the California Current.
- No. 16. Crowe, F. J. and R. A. Schwartzlose. 1972. Release and recovery records of drift bottles in the California Current region, 1955–1971.
- No. 17. Ahlstrom, E. H. 1972. Distributional atlas of fish larvae in the California Current region: six common mesopelagic species –*Vinciguerria lucetia*, *Triphoturus mexicanus*, *Stenobrachius leucopsarus*, *Leuroglossus stibius*, *Bathylagus wesethi*, and *Bathylagus ochotensis*, 1955–1960.
- No. 18. Brinton, E. 1973. Distributional atlas of Euphausiacea (Crustacea) in the California Current region, Part II.
- No. 19. Bowman, T. E. and M. W. Johnson. 1973. Distributional atlas of calanoid copepods in the California Current region, 1949 and 1950.
- No. 20. Thomas, W. H. and D. L. R. Seibert. 1974. Distribution of nitrate, nitrite, phosphate, and silicate in the California Current region, 1969.
Owen, R. W., Jr. 1974. Distribution of primary production, plant pigments and Secchi depth in the California Current region, 1969.
Smith, P. E. 1974. Distribution of zooplankton volumes in the California Current region, 1969.
- No. 21. Fleminger, A., J. D. Isaacs, and J. G. Wyllie. 1974. Zooplankton biomass measurements from CalCOFI cruises of July 1955 to 1959 and remarks on comparison with results from October, January, and April cruises of 1955 to 1959.
- No. 22. Namias, J. 1975. Northern hemisphere seasonal sea level pressure and anomaly charts, 1947–1974.
- No. 23. Ahlstrom, E. H. and H. G. Moser. 1975. Distributional atlas of fish larvae in the California current region: flatfishes, 1955–1960.
- No. 24. Brinton, E. and J. G. Wyllie. 1976. Distributional atlas of euphausiid growth stages off southern California, 1953–1956.
- No. 25. Eber, L. E. 1977. Contoured depth-time charts (0 to 200 m, 1950–1966) of temperature, salinity, oxygen and sigma-t at 23 CalCOFI stations in the California Current.
- No. 26. Ahlstrom, E. H., H. G. Moser, and E. M. Sandknop. 1978. Distributional atlas of fish larvae in the California Current region: rockfishes, *Sebastodes* spp., 1950–1975.
- No. 27. Namias, J. 1979. Northern hemisphere seasonal 700 mb height and anomaly charts, 1947–1978, and associated North Pacific sea surface temperature anomalies.
- No. 28. Hewitt, R. 1980. Distributional atlas of fish larvae in the California Current region: northern anchovy, *Engraulis mordax* (Girard), 1966–1979.
- No. 29. Namias, J. 1981. Teleconnections of 700 mb height anomalies for the northern hemisphere.
- No. 30. Lynn, R. J., K. A. Bliss, and L. E. Eber. 1982. Vertical and horizontal distributions of seasonal mean temperature, salinity, sigma-t, stability, dynamic height, oxygen, and oxygen saturation in the California Current, 1950–1978.
- No. 31. Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, S. R. Charter, C. A. Meyer, E. M. Sandknop, and W. Watson. 1993. Distributional atlas of fish larvae and eggs in the California Current region: taxa with 1000 or more total larvae, 1951–1984.
- No. 32. Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, S. R. Charter, C. A. Meyer, E. M. Sandknop, and W. Watson. 1994. Distributional atlas of fish larvae in the California Current region: taxa with less than 1000 total larvae, 1951–1984.
- No. 33. Moser, H. G. (ed.). 1996. The early stages of fishes in the California Current region.
- No. 34. Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, W. Watson, S. R. Charter, and E. M. Sandknop. 2001. Distributional atlas of fish larvae and eggs in the Southern California Bight region: 1951–1998.
- No. 35. Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, W. Watson, S. R. Charter, and E. M. Sandknop. 2002. Distributional atlas of fish larvae and eggs from Manta (surface) samples collected on CalCOFI surveys from 1977 to 2000.

**DISTRIBUTIONAL ATLAS OF FISH LARVAE AND EGGS FROM MANTA (SURFACE)
SAMPLES COLLECTED ON CALCOFI SURVEYS FROM 1977 TO 2000**

**Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, W. Watson,
S. R. Charter, and E. M. Sandknop**

CalCOFI ATLAS NO. 35

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May, 2002

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Introduction

Surface waters are inhabited by the early life stages and adults of a large array of fish species. Eggs and larvae of some species occur throughout the upper water column whereas those of others are found exclusively in surface waters and have evolved special adaptations for life in that zone (Zaitsev 1970; Hempel and Weichert 1972; Moser 1981). An

early survey (Ahlstrom and Stevens 1976) in the northeast Pacific (20° – 48° N, seaward to 148° W), employing a neuston⁴ net, provided initial information on the distribution and abundance of surface-living ichthyoplankton in this region. Gruber et al. (1982) used a neuston net of the same design to sample ichthyoplankton in a two-year study in the Southern California Bight (SCB). These early studies provided useful information; however, the volume of water filtered by the nets could not be measured, thus the catches could not be standardized. Doyle (1992a, b), used a quantitative surface sampler (Sameoto and Jaroszynski 1969) in a study of the distribution and abundance of ichthyoplankton off northern California, Oregon, and Washington. The Manta net (Brown and Cheng 1981) was used to sample surface ichthyoplankton in a number of site-intensive studies off the southern California coast (Brewer et al. 1981; Schlotterbeck and Connally 1982; Barnett et al. 1984; Moser and Pommeranz 1999).

This atlas summarizes the spatial and temporal distribution and abundance of 93 ichthyoplankton taxa or categories collected in Manta net tows on CalCOFI biological-oceanographic survey cruises from December, 1977 to October, 2000. CalCOFI Atlases 31 and 32 presented distributional

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⁴Usage of the term “neuston” for surface-living marine organisms is controversial because it was applied originally to organisms associated with the surface film in freshwater habitats (Naumann 1917). Banse (1975) reviewed in detail the evolution of the usage of this term, a related term, “pleuston”, and the various subdivisions of each. Neuston is now used by most workers in referring to the uppermost (upper ~10–20 cm) layer of the sea and to the assemblage of organisms that lives in that zone, either permanently or facultatively (Zaitsev 1970; Hempel and Weikert 1972; Peres 1982; Doyle 1992b). We accept this definition and use it interchangeably with the more general term “surface” (e.g., surface waters, surface zone, surface tow, surface assemblage). The net used by Ahlstrom and Stevens (1976) consisted of a rectangular frame, modified from 1-m ring net frame, with styrofoam floats attached inside the ends. The netting was identical to that used in a standard CalCOFI 1-m ring net with 0.505 mm mesh. There was no flow meter attached to the net frame, which was partially out of the water during the tow.

summaries for all taxa taken in oblique tows on surveys that covered the greater CalCOFI sampling area (Figure 1) extending from northern California to Cabo San Lucas, Mexico during 1951–84. Since 1985, CalCOFI surveys have been conducted on a quarterly basis over a sampling grid consisting of 66 standard stations in the SCB region (Figure 2). CalCOFI Atlas 34 summarized distributions and abundances of fish eggs and larvae taken in oblique tows from 1951 to 1998 in the area defined by the present survey pattern. Surface sampling with the Manta net was initiated during the 1977–78 survey. Manta tows were taken on some (~40%) of the total net tow stations during 1977–78 because surface tows were not yet part of the standard station protocol. On subsequent surveys Manta tows were taken on most net tow stations (86–88% of the stations during surveys in 1980–85, 92% during 1986–87, and >97% during 1988–2000).

A data base for ichthyoplankton and station data for all Manta tows taken on 19 CalCOFI surveys from 1977 to 2000 has been established. A data report for the 1977–78 survey is published (Moser et al. 2001b) and the remaining reports are in press (Table 1).

The California Current region is one of the most oceanographically and zoogeographically complex regions in the world ocean. The CalCOFI sampling grid overlays the California Current, three coastal zoogeographic provinces (Oregonian, San Diegan, and Panamic), a coastal upwelling zone, and three oceanic water masses (Subarctic, Pacific Central, and equatorial water of the eastern tropical Pacific) (Moser 1996; Moser et al. 2001a). These water masses converge in the SCB region where complexity is heightened by a field of mesoscale eddies that extends from the inner margin of the California Current into the Bight, an undercurrent flowing poleward along the slope, a poleward flowing inshore countercurrent, and a persistent frontal zone (Ensenada Front) at the southwest corner of the Bight (Lynn and Simpson 1987; Haury et al. 1993). The inner third of the present CalCOFI survey pattern overlies the continental borderland off southern California, a region of complex topography that includes the mainland continental shelf and slope, ~13 deep-water basins, and numerous islands and banks and their shelves. The islands and banks

extend coastal habitats >100 n.m. offshore and the basins provide deep-water habitats and zoogeographic refugia for a wealth of midwater species (Moser et al. 2001a).

The cyclic warming and cooling of equatorial waters, known as the El Niño/Southern Oscillation (ENSO), has produced a series of El Niños (1957–58, 1963, 1982–83, 1992–93, and 1997) and La Niñas (1954–56, 1988–89, 1998–99) in the California Current region. ENSO episodes are embedded in a low frequency oscillation in the North Pacific, the Pacific Decadal Oscillation (PDO), that is related to basin scale changes in atmospheric pressure, particularly the intensification and position of the Aleutian Low Pressure System (Mantua et al. 1997; Hollowed et al. 1998; Schwing et al., in press.). The resulting 20–30 year cooling and warming cycle produced two cool ocean regimes (1900–24 and 1947–76) and two warm regimes (1925–46 and 1977–present?) during the past century. Some investigators (e.g., Minobe 2000; Schwing and Moore 2000) have suggested that the 1998–99 La Niña may have marked a shift from a warm to a cool regime in the northeast Pacific. Recently, Hare and Mantua (2000) hypothesized another regime shift in 1989. They found that the biological signals associated with this shift were much clearer than the physical climate indices and that the shift did not result in a return to pre-1977 ocean conditions. Among the apparent biological consequences of these regime shifts are basin-wide changes in primary and secondary production and in the abundance of eastern Pacific fish stocks. Production in the California Current region decreased dramatically after the regime shift of 1977 (Brodeur and Ware 1992; Roemmich and McGowan 1995a, b; Ware 1995; Hayward 1997; McGowan et al. 1998) as did groundfish and salmon stocks (Francis and Hare 1994; Mantua et al. 1997; Francis et al. 1998; Hollowed et al. 1998). Major shifts occurred in the distributions of larval fishes in response to ENSO events and the PDO, as the boundaries between Subarctic and Equatorial water masses shifted latitudinally and intrusion of the Central Water Mass into the SCB region waxed and waned (Moser et al. 1987, 2001a; Smith and Moser 1988). The time series for surface-caught fish larvae represents primarily the warm ocean regime

initiated in 1977; however, it spans the regime shifts hypothesized for 1988–89 and 1998–99 and includes El Niño events in 1982–83, 1992–93, and 1997 and La Niña events in 1988 and 1998–99. In the SCB region the long-term trend of increasing surface temperatures during the warm regime was punctuated by episodic increases and decreases during El Niño and La Niña events, respectively (Figure 3A). The trend for zooplankton volumes during this period was opposite to temperature, with short-term increases during La Niña episodes (Figure 3A). During this period the average seasonal temperature range was ~4.5°C, with a March minimum and a September maximum (Figure 3B). Maximum zooplankton volume preceded the temperature peak by 3–4 months with highest values in May–June (Figure 3B). Average temperatures in the borderland region of the CalCOFI survey pattern generally were slightly lower than in the offshore region, except during 1981 and 1984 when they were ~0.5–1.0 °C higher (Figure 3C). Average borderland surface temperatures were lower than offshore during the first five months of the year, became slightly higher during June and August, and were lower during October and November (Figure 3D). Zooplankton volumes in the borderland region were consistently slightly higher than in offshore waters during most years (Figure 3E) and during most months (Figure 3F). The seasonal peaks differed by one month between the two regions, with the maximum in May in the borderland and in June in offshore waters (Figure 3F). The information in this atlas is presented in a format that permits the reader to interpret, in general terms, the affects that high and low frequency changes in ocean conditions may have had on larval fish populations inhabiting surface waters of the SCB region from December 1977 to October 2000.

The primary time series for this atlas is composed of 4,704 net tows taken during 1977–2000 within the boundaries of the present CalCOFI survey (Tables 2 and 3). These include all standard CalCOFI survey Manta tows taken since 1985 when occupancy of the present pattern was initiated. On surveys prior to 1985 in the Southern California Bight nearshore stations did not always correspond to the exact positions of the 66 nominal stations used on surveys since 1985. In order to expedite the construction of the distribution maps, data for stations other than the

nominal stations of the present pattern were assigned to the closest of the 66 nominal stations (Table 3). Maps and graphs summarizing seasonal and annual occurrence and abundance in the SCB region from 1977 to 2000 are based on the part of the time series defined by the present survey pattern. Tows prior to 1985 are a subset from the wider-ranging CalCOFI surveys (Hewitt 1988; Moser et al. 1993, 1994). An additional 1,415 tows taken north and south of the present survey area during 1977–78, 1980–81, and 1984 were used in the preparation of broad-scale maps that show the overall distribution and abundance of taxa taken in Manta tows on CalCOFI surveys.

Surface tows were made with a modified version of the Manta net originally designed by Brown and Cheng (1981). It consists of a rectangular mouth 15.5 cm deep and 86 cm wide attached to a frame that supports square lateral extensions covered with plywood and urethane foam (Figure 4). These extensions stabilize the net when it is towed and keep the top of the net at the sea surface. The net is constructed of 0.505 mm nylon mesh. The towing bridle is asymmetrical with one side longer than the other; when the net is towed this bridle arrangement forces the mouth away from the ship at a slight angle. A General Oceanics flowmeter is suspended across the center of the net mouth to measure the amount of water filtered during each tow. At each Manta tow station the tow line from the bridle is attached to the hydrographic wire and then lowered to slightly below the surface of the water before the net is deployed. The net is towed at a ship speed of 1.0–2.0 knots. In 1977–78 tow duration was 3 minutes; in subsequent survey years the towing time for each tow was 15 min. Procedures for sample handling and preservation were the same as those used for samples from oblique net tows (Kramer et al. 1972).

All taxa from the part of the CalCOFI time series used in this atlas are listed in rank order of abundance (raw larval count) summed for all occurrences (Table 4) and in rank order of total occurrences (Table 5). These tables include 188 total larval taxa (including the unidentified and disintegrated categories). Distributional summaries are presented for the larval fish taxa that had ~10 or

more total larvae and also for five other categories: total fish larvae, total fish eggs, and the eggs of northern anchovy *Engraulis mordax*, Pacific sardine *Sardinops sagax*, and Pacific saury *Cololabis saira*. These summaries were not produced for some higher taxonomic categories (genera, families, orders) that contained >10 total larvae (Table 4) because their distributions were better represented in analyses of component taxa. In one case, *Sebastolobus altivelis* and *Sebastolobus* spp. were combined in order to show the distribution and abundance of the genus.

Since 1951, fish larvae in CalCOFI samples have been identified to species or to the lowest taxon that knowledge and specimen condition permitted. The number of identifiable taxa increased with the improvement of taxonomic knowledge and competency. Historical identifications were evaluated thoroughly during the development of the CalCOFI ichthyoplankton computer data base in 1984–88 and historical advances in the identification process were standardized at approximately decadal intervals. Our ability to identify larvae in the California Current region improved greatly during 1988–95 as a result of taxonomic research that culminated in a taxonomic monograph on the ontogenetic stages of fishes of this region (Moser 1996). This resulted in a ~ 60% increase in the number of identifiable taxa beginning in 1985. These changes are documented in previous CalCOFI ichthyoplankton data reports and atlases; the following interpretive notes and caveats are pertinent to this atlas:

Ceratoscopelus townsendi—*Ceratoscopelus* larvae in the CalCOFI sampling region are *C. townsendi*, a species associated with the California Current; at the western margin of the California Current (approx. westward of station 200), *C. townsendi* is replaced by the tropical-subtropical cosmopolite, *C. warmingii* (Wisner 1976; Butler et al. 1997); larvae of the two species are not distinguishable.

Cheilopogon spp.—small damaged specimens that could not be identified to species; most, perhaps all, are *C. heterurus* or *C. pinnatibarbatus*.

Diaphus spp.—*Diaphus theta* is the dominant *Diaphus* species in the survey area and most, if not all, of the larvae from the SCB region are this

species; the generic category is used because a small proportion of the *Diaphus* larvae captured at the outer margin of the survey pattern may represent other species whose larvae are identical to those of *D. theta*.

Disintegrated fish larvae—larvae that could not be identified because of their poor condition; separated from the "unidentified" category to monitor the general condition of the ichthyoplankton samples through the time series.

Nannobrachium spp.—most of the larvae in this category are small (< 5 mm), often poorly preserved specimens of this genus, recently separated from *Lampanyctus* by Zahuranec (2000) on the basis of small or absent pectoral fins in adults; two *Nannobrachium* species, *N. ritteri* (formerly *L. ritteri*) and *N. regale* (formerly *L. regalis*), occur commonly in the present CalCOFI survey pattern; larvae of these species > ~ 5 mm have been identified since 1954 and were referred to the genus *Lampanyctus* in CalCOFI Atlases 31 and 32; adults of *Nannobrachium* species are much more common than those of *Lampanyctus* in the present CalCOFI survey pattern, however, in samples from the subtropical-tropical eastern Pacific the opposite may be true.

Parophrys vetulus—Sakamoto (1984) changed pleuronectid generic designations for some of the species in the CalCOFI area, including *Parophrys vetulus*, which was transferred into *Pleuronectes*; although these changes were incorporated in the lists of Robins et al. (1991) and Eschmeyer (1998) we follow Nelson (1994) in retaining the older nomenclature because Sakamoto's (1984) changes were based on a phenetic study; also, the older names are used in the major identification guides to fishes of our region (Miller and Lea 1972, Eschmeyer et al. 1983, Matarese et al. 1989, and Moser 1996).

Unidentified fish larvae—larvae that were generally in good condition but could not be identified because of their small size or early stage of development.

Vinciguerria lucetia—*V. lucetia*, an eastern tropical Pacific species, is common in the present CalCOFI region whereas the central water mass species *V. poweriae* is rarely encountered; a small percentage of *V. poweriae* larvae may have been included in the *V. lucetia* category because of the difficulty in separating early larvae of the two species.

Relative Abundance: Manta vs. Oblique

Northern anchovy was the most abundant taxon in both the Manta and the oblique time series during 1977–2000 in the present survey area (Table 6), accounting for 64.6% of the total larvae in Manta samples and for 43.2% of the total in oblique samples. Other species that were represented in relatively large numbers in Manta samples were poorly sampled by oblique net tows. For example, Pacific saury ranked third in total count in Manta samples but only 87th in oblique samples. Mussel blenny *Hypsoblennius jenkinsi* ranked fourth in total count in Manta samples and 71st in oblique samples; its congener, *H. gilberti*, ranked 18th in Manta samples and 193rd in oblique samples. Jacksmelt *Atherinopsis californiensis* and grunion *Leuresthes tenuis* ranked 10th and 13th in Manta catches but only 162nd and 215th, respectively, in oblique catches. This also was true for some commercially important species such as kelp greenling *Hexagrammos decagrammus* and cabezon *Scorpaenichthys marmoratus*, which ranked 8th and 11th in Manta samples but only 197th and 105th, respectively in oblique samples. Three flyingfish categories (Family Exocoetidae) were captured in moderate numbers by Manta nets but were not taken at all in oblique tows (Table 6). Some taxa that were relatively abundant in oblique samples were rare in Manta catches. For example, the blacksmelt (Bathylagidae) species, *Leuroglossus stilbius*, *Bathylagus ochotensis*, and *B. wesethi*, have relatively deep-living larvae (Ahlstrom 1959; Moser and Smith 1993), and ranked 5th, 8th, and 11th, respectively, in oblique samples but ranked 81st, 70th, and 112th, in Manta samples. Other examples are the myctophids *Protomyctophum crockeri* and *Diogenichthys atlanticus*, which ranked 12th and 17th, respectively, in oblique samples but ranked 84th and 81st in Manta net catches during 1977–2000. Comparison of catches from the two types of tows emphasizes the importance of taking surface tows as

well as integrated tows, particularly for sampling obligate neustonic larvae of heavily impacted nearshore fishes such as cabezon, greenlings, and lingcod (*Ophiodon elongatus*).

Larval Fish Assemblages

A robust method of revealing assemblages of organisms is to examine their degree of co-occurrence. Recurrent group analysis (Fager 1957, 1963) defines groups of taxa that occur together frequently and thus share a common environment. Two procedures are used: first an index of affinity (program name AFFINITY) is calculated for each pair of taxa that ever occur together and then taxa are formed into groups (program name REGROUP) using a chosen minimum affinity index (0.2 in this study). The category “group member” is supplemented by the term “associate” for taxa that have significant affinity indices with one or more but not all members in one or more of the groups. Previous recurrent group analyses of the CalCOFI ichthyoplankton time series (Moser et al. 1987; Moser and Smith 1993) have defined larval fish assemblages from oblique tows taken over the extended CalCOFI survey pattern from 1951–84. A recurrent group analysis of neuston and oblique ichthyoplankton samples from 17 stations within the continental borderland of the SCB during 1974–76 revealed two neustonic recurrent groups (Gruber et al. 1982). Group 1 consisted of northern anchovy, the rockfish genus *Sebastodes*, and white croaker *Genyonemus lineatus*, with the blenny genus *Hypsoblennius*, the silversides family Atherinidae, and kelp greenling as associates, and Group 2 contained Pacific saury and the flyingfish family Exocoetidae (Gruber et al. 1982). Our analysis of 4,704 net tows taken during 1977–2000 within the boundaries of the present CalCOFI survey pattern, extending seaward to a maximum of ~376 n. mi. revealed four neustonic recurrent groups (Figure 5A; Table 7). One group, consisting of northern anchovy, *Sebastodes* spp., and cabezon, had Pacific sardine as an associate and was weakly connected to another three-member group consisting of Pacific barracuda *Sphyraena argentea*, mussel blenny, and blacksmith *Chromis punctipinnis*. The latter group had the kelp and sandbass genus *Paralabrax* and chub mackerel *Scomber japonicus* as associates

(Figure 5A; Table 7). The other two groups each contained two members and lacked connections with each other or with the three-member groups. Panama lightfish *Vinciguerria lucetia* and dogtooth lampfish *Ceratoscopelus townsendi* comprised one group and the other consisted of two haemulids, sargo *Anisotremus davidsoni* and salema *Xenistius californiensis*, with garibaldi *Hypsypops rubicundus* as an associate. Only two taxa, cabezon and mussel blenny, in the neustonic recurrent groups are neuston obligates; the others have relatively shallow distributions or, in the case of multiple-species taxa (e.g., *Sebastodes*), include species with shallow distributions.

The recurrent group assemblage structure for the oblique samples from the same part of the time series was considerably more complex, consisting of two large groups representing the “northern” and “southern” complexes of Moser et al. (1987) and Moser and Smith (1993) (Figure 5B; Table 8). The eight-member southern complex group (Oblique Group 2) is a mix of species from several of the southern complex groups described by Moser et al. (1987) and has 11 southern-complex associate taxa. The six-member northern complex group (Oblique Group 1) contains two species (*E. mordax* and *Sebastodes* spp.) in common with one of the neustonic groups and is strongly interconnected to a group (Oblique Group 3) composed of three northern complex midwater species (Figure 5B). Oblique Group 3 also is strongly connected to the 11-member southern complex group. Associate taxa to the three-member group are two rockfish species, Pacific sardine, and jack mackerel. The remaining two groups, each with two members, contain shorefish of the San Diegan province and lack critical affinities with the other recurrent groups and with each other (Figure 5B; Table 8).

Length Frequencies

Neuston nets tend to capture relatively larger larvae (both average and maximum lengths) within a given taxon, compared to oblique nets (Moser 1981; Doyle 1992b). Obligate neustonic species generally hatch from large eggs that produce relatively large larvae. This is true for demersal spawners (e.g., cabezon and other cottids, hexagrammids), pelagic spawners (e.g.,

sablefish), and species that attach their eggs to flotsam (e.g., sauries, flyingfishes). For facultative taxa the larger sizes may indicate movement of larger larvae to surface waters during ontogeny or may reflect diel movements. Hunter and Sanchez (1976) suggested that northern anchovy larvae swim to the surface at night, fill their gas bladders with air, and remain suspended near the surface during non-feeding hours as an energy conservation mechanism. Another possibility for the presence of larger larvae in neuston samples may be related to the fact that the upper escape plane is eliminated in a surface tow, thus limiting the avoidance options for a larva trying to escape capture.

In the CalCOFI time series, body lengths are measured for five species during the sorting and identification processes. Of these species, northern anchovy shows the largest disparity in length frequencies between Manta and oblique catches (Figure 6A). In oblique samples, the peak is at ~3–4 mm whereas it is at ~10 mm in neuston samples, the length which Hunter and Sanchez (1976) proposed for migration to the surface. In Pacific sardine, peak lengths are at ~5–6 mm for both surface and oblique nets; however, the length frequency curve declines abruptly in oblique samples whereas relatively large numbers of larvae are present in the neuston in length classes up to 20 mm (Figure 6B). Sardine larvae, like anchovy, are physostomous and may undergo a diel migration similar to that suggested for northern anchovy. Length distributions of the other measured species are less disparate for the two samplers. In chub mackerel (Figure 6C), the peak length in Manta catches is ~1 mm larger than in oblique net catches (3–4 mm vs. 4–5 mm); otherwise the length frequency curves are similar. In jack mackerel the curves are similar with slightly larger larvae in Manta catches (Figure 6D). Pacific hake samples are dominated by small size classes and average larval lengths of oblique net catches are slightly larger than those from Manta nets (Figure 6E) (Moser et al. 1997).

Explanation of Charts and Graphs

The distributions are summarized in a series of maps and graphs, one page per taxon. Abundance of

larvae or eggs is expressed as the number per 100 m³ of water filtered or as the raw count when appropriate. Occurrence is expressed as the proportion of positive tows for larvae or eggs of a particular taxon. The upper left panel is a map of average abundance at individual CalCOFI stations on the wide-ranging surveys of 1977-78, 1980-81, and 1984. The upper right panel is a map of average abundance at stations within the boundaries of the present survey pattern from 1977 to 2000. If a taxon did not occur during 1977–84 there is no map in the upper left panel. The total raw count of larvae represented in each map is listed on the map below the range of years. The middle left panel is a graph of seasonal occurrence for the stations where the taxon occurred consistently within the survey area (see Table 9 for the range of stations used in these calculations). The middle right panel is a graph of average seasonal abundance for the stations where the taxon occurred consistently within the survey area (see Table 9). The lower left panel is a graph of annual occurrence during the principal spawning period at the stations where the taxon occurred consistently within the survey area (see Table 9 for the range of months used in these calculations). The lower right panel is a graph of average annual abundance during the principal spawning period of the taxon for the stations where its larvae were consistently found within the survey area (see Table 9). ENSO events are indicated by vertical bars (El Niño, shading; La Niña, hatching). Dashed lines indicate the period of triennial CalCOFI surveys.

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Literature Cited

- Ahlstrom, E. H. 1959. Vertical distribution of pelagic fish eggs and larvae off California and Baja California. *Fish. Bull.* 60:107–146.
- Ahlstrom, E. H. and E. G. Stevens. 1976. Report of neuston (surface) collections made on an extended CalCOFI cruise during May 1972. *Calif. Coop. Oceanic Fish. Invest. Rep.* 18:167–180.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002a. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1980-81. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-319. 100 pp.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002b. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1985. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-321. 36 pp.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002c. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1989. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-325. 45 pp.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002d. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative

- Oceanic Fisheries Investigations survey cruises in 1993. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-329. 41 pp.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002e. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1997. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-333. 41 pp.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002f. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1998. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-334. 43 pp.
- Ambrose, D. A., R. L. Charter, and H. G. Moser. 2002g. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1999. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-335. 39 pp.
- Banse, K. 1975. Pleuston and neuston: on the categories of organisms in the uppermost pelagic. Int. Rev. ges. Hydrobiol. 60(4):439–447.
- Barnett, M. A., A. E. Jahn, P. D. Sertic, and W. Watson. 1984. Distribution of ichthyoplankton off San Onofre, California, and methods for sampling very shallow coastal waters. Fish. Bull. 82:97–111.
- Brewer, G. D., R. J. Lavenberg, and G. E. McGowen. 1981. Abundance and vertical distribution of fish eggs and larvae in the Southern California Bight: June and October 1978. Rapp. P.-v. Réun. Cons. int. Explor. Mer 178:165–167.
- Brodeur, R. D. and D. M. Ware. 1992. Long-term variability in zooplankton biomass in the subarctic Pacific Ocean. Fish. Oceanogr. 1(1):32–38.
- Brown, D. M. and L. Cheng. 1981. New net for sampling the ocean surface. Mar. Ecol. Prog. Ser. 5:224–227.
- Butler, J. L., H. G. Moser, W. Watson, D. A. Ambrose, S. R. Charter, and E. M. Sandknop. 1997. Fishes collected by midwater trawl during two cruises of the *David Starr Jordan* in the northeastern Pacific Ocean, April–June and September–October, 1972. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-TM-NMFS-SWFSC-244. 83 pp.
- Charter, S. R., R. L. Charter, and H. G. Moser. 2002a. Ichthyoplankton and the station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations suvey cruises in 1984. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-320. 84 pp.
- Charter, S. R., R. L. Charter, and H. G. Moser. 2002b. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1986. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-322. 40 pp.
- Charter, S. R., R. L. Charter, and H. G. Moser. 2002c. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1990. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-326. 41 pp.
- Charter, S. R., R. L. Charter, and H. G. Moser. 2002d. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1994. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-330. 40 pp.
- Doyle, M. J. 1992a. Patterns in distribution and abundance of ichthyoplankton off Washington, Oregon, and northern California (1980–1987). U.S. Dep. Commer., Nat. Mar. Fish. Serv., Alaska Fish. Sci. Ctr. Proc. Rep. 92-14. 344 pp.
- Doyle, M. J. 1992b. Neustonic ichthyoplankton in the northern region of the California Current ecosystem. Calif. Coop. Oceanic Fish. Invest. Rep. 33:141–161.

- Eschmeyer, W. N. (ed.). 1998. Catalog of fishes. Center for Biodiversity Research and Information. California Academy of Sciences. Spec. Publ. 1. Vols. I-III. 2905 pp.
- Eschmeyer, W. N., E. S. Herald, and H. Hammann. 1983. A field guide to Pacific coast fishes of North America. Houghton Mifflin Co. Boston. 336 pp.
- Fager, E. W. 1957. Determination and analysis of recurrent groups. *Ecology* 38:586–595.
- Fager, E. W. 1963. Communities of organisms. Pages 415–437. In M. N. Hill (ed.). *The sea*. Vol. II. Interscience. New York, N.Y.
- Francis, R. C. and S. R. Hare. 1994. Decadal-scale regime shifts in the large marine ecosystems of the North-east Pacific: a case for historical science. *Fish. Oceanogr.* 3(4):279–291.
- Francis, R. C., S. R. Hare, A. B. Hollowed, and W. S. Wooster. 1998. Effects of interdecadal climate variability on the oceanic ecosystems of the NE Pacific. *Fish. Oceanogr.* 7(1):1–21.
- Gruber, D., E. H. Ahlstrom, and M. M. Mullin. 1982. Distribution of ichthyoplankton in the Southern California Bight. *Calif. Coop. Oceanic Fish. Invest. Rep.* 23:172– 179.
- Hare, S. R. and N. J. Mantua. 2000. Empirical evidence for North Pacific regime shifts in 1977 and 1989. *Prog. Oceanogr.* 47:103-145.
- Haury, L. R., E. L. Venrick, C. L. Fey, J. A. McGowan, and P. P. Niiler. 1993. The Ensenada Front: July 1985. *Calif. Coop. Oceanic Fish. Invest. Rep.* 34:69–88.
- Hayward, T. L. 1997. Pacific ocean climate change: atmospheric forcing, ocean circulation and ecosystem response. *TREE*. 12(4):150–154.
- Hempel, G. and H. Weikert. 1972. The neuston of the subtropical and boreal northeastern Atlantic Ocean. A review. *Mar. Biol.* 13:70–88.
- Hewitt, R. P. 1988. Historical review of the oceanographic approach to fishery research. *Calif. Coop. Oceanic Fish. Invest. Rep.* 29:27– 41.
- Hollowed, A. B., S. R. Hare, and W. S. Wooster. 1998. Pacific Basin climate variability and patterns of Northeast Pacific marine fish production. Pages 89–104 in G. Holloway, P. Muller, and D. Henderson (eds.) *Biotic impacts of extratropical climate variability in the Pacific*. Proc. ‘Aha Huliko’a Hawaiian Winter Workshop, Univ. Hawaii at Manoa, January 25–29, 1998. SOEST Spec. Publ. 156 pp.
- Hunter, J. R. and C. Sanchez. 1976. Diel changes in swim bladder inflation of the larvae of the northern anchovy, *Engraulis mordax*. *Fish. Bull.* 74: 847–855.
- Kramer, D., M. J. Kalin, E. G. Stevens, J. R. Thrailkill, and J. R. Zwiefel. 1972. Collecting and processing data on fish eggs and larvae in the California Current region. NOAA Tech. Rep. NMFS Circ-370, 38 pp.
- Lynn, R. J. and J. J. Simpson. 1987. The California Current System: The seasonal variability of its physical characteristics. *J. Geophys. Res.* 92 (C12):12,947–12,966.
- Mantua, N. J., S. R. Hare, Y. Zhang, J. M. Wallace, and R. C. Francis. 1997. A Pacific interdecadal climate oscillation with impacts on salmon production. *Bull. Amer. Meteor. Soc.* 78:1069–1079.
- Matarese, A. C., A. W. Kendall, Jr., D. M. Blood, and B. M. Vinter. 1989. Laboratory guide to early life history stages of northeast Pacific fishes. U.S. Dep. Commer., NOAA Tech. Rep. NMFS 80. 652 pp.
- McGowan, J. S., D. R. Cayan, and L. M. Dorman. 1998. Climate-ocean variability and ecosystem response in the northeast Pacific. *Science*. 281:210–217.

- Miller, D. J. and R. N. Lea. 1972. Guide to the coastal marine fishes of California. Calif. Dep. Fish Game Fish Bull. 157. 235 pp.
- Moser, H. G. 1981. Morphological and functional aspects of marine fish larvae. Pages 89–131 in R. Lasker, ed. Marine fish larvae. Morphology, ecology, and relation to fisheries. Univ. Wash. Press, Seattle.
- Moser, H. G. (ed.). 1996. The early stages of fishes in the California Current region. CalCOFI Atlas 33. 1505 pp.
- Moser, H. G. and T. Pommeranz. 1999. Vertical distribution of eggs and larvae of northern anchovy, *Engraulis mordax*, and of the larvae of associated fishes at two sites in the Southern California Bight. Fish. Bull. 97:920–943.
- Moser, H. G. and P. E. Smith. 1993. Larval fish assemblages of the California Current region and their horizontal and vertical distributions across a front. Bull. Mar. Sci. 53: 645–691.
- Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, S. R. Charter, C. A. Meyer, E. M. Sandknop, and W. Watson. 1993. Distributional atlas of fish larvae and eggs in the California Current region: taxa with 1000 or more total larvae, 1951 through 1984. CalCOFI Atlas 31. 233 pp.
- Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, S. R. Charter, C. A. Meyer, E. M. Sandknop, and W. Watson. 1994. Distributional atlas of fish larvae in the California Current region: taxa with less than 1000 total larvae, 1951 through 1984. CalCOFI Atlas 32. 181 pp.
- Moser, H. G., R. L. Charter, P. E. Smith, D. A. Ambrose, W. Watson, S. R. Charter, and E. M. Sandknop. 2001a. Distributional atlas of fish larvae and eggs in the Southern California Bight region: 1951–1998. CalCOFI Atlas 34. 166 pp.
- Moser, H. G., R. L. Charter, D. A. Ambrose, and E. M. Sandknop. 2001b. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1977–78. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-313. 58 pp.
- Moser, H. G., N. C. H. Lo, and P. E. Smith. 1997. Vertical distribution of Pacific hake eggs in relation to stage of development and temperature. Calif. Coop. Oceanic Fish. Invest. Rep. 38:120–126.
- Moser, H. G., P. E. Smith, and L. E. Eber. 1987. Larval fish assemblages in the California Current region, 1954–1960, a period of dynamic environmental change. Calif. Coop. Oceanic Fish. Invest. Rep. 28:97–127.
- Naumann, E. 1917. Beiträge zur Kenntnis des Teichnannoplanktons. II. Über das Neuston das Süßwassers. Biol. Zentralbl. 37:98–106.
- Nelson, J. S. 1994. Fishes of the world. Third edition. John Wiley and Sons, N.Y. 600 pp.
- Peres, J. M. 1982. Specific pelagic assemblages: 1. Assemblages at the air-ocean interface In Marine Ecology. O. Kinne (ed.). 5 (1):313–372.
- Robins, C. R., R. M. Bailey, C. E. Bond, J. R. Brooker, E. A. Lachner, R. N. Lea, and W. B. Scott. 1991. Common and scientific names of fishes from the United States and Canada, fifth edition. Am. Fish. Soc. Spec. Publ. 20. 183 pp.
- Roemmich, D. and J. McGowan. 1995a. Climatic warming and decline of zooplankton in the California Current. Science. 267:1324–1326.
- Roemmich, D. and J. McGowan. 1995b. Sampling zooplankton: correction. Science. 268:352–353.
- Sakamoto, K. 1984. Interrelationships of the family Pleuronectidae (Pisces: Pleuronectiformes). Mem. Fac. Fish. Hokkaido Univ. 31:95–215.

- Sameoto, D. D. and L. O. Jaroszynski. 1969. Otter surface trawl: a new neuston net. J. Fish. Res. Bd. Can. 26:2240–2244.
- Sandknop, E. M., R. L. Charter, and H. G. Moser. 2002a. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1987. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-323. 40 pp.
- Sandknop, E. M., R. L. Charter, and H. G. Moser. 2002b. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1991. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-327. 41 pp.
- Sandknop, E. M., R. L. Charter, and H. G. Moser. 2002c. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1995. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-331. 42 pp.
- Schlotterbeck, R. E. and D. W. Connally. 1982. Vertical stratification of three nearshore southern California larval fishes (*Engraulis mordax*, *Genyonemus lineatus*, and *Seriphis politus*). Fish. Bull. 80:895–902.
- Schwing, F. B. and C. S. Moore. 2000. A year without summer for California or a harbinger of a climate shift. Trans. Am. Geophys. Union. 81:301.
- Schwing, F. B., T. Murphree, and P. M. Green. In press. A climate index for the northeast Pacific. Prog. Oceanogr.
- Smith, P. E. and H. G. Moser. 1988. CalCOFI time series: an overview of fishes. Calif. Coop. Oceanic Fish. Invest. Rep. 29:66–78.
- Ware, D. M. 1995. A century and a half of change in the climate of the NE Pacific. Fish. Oceanogr. 4 (4):267-277.
- Minobe, S. 2000. Spatio-temporal structure of the pentadecadal variability over the North Pacific. Prog. Oceanogr. 47:381–408.
- Watson, W., R. L. Charter, and H. G. Moser. 2002a. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1988. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-TM-NMFS-SWFSC-324. 44 pp.
- Watson, W., R. L. Charter, and H. G. Moser. 2002b. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1992. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-328. 40 pp.
- Watson, W., R. L. Charter, and H. G. Moser. 2002c. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 1996. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-332. 45 pp.
- Watson, W., R. L. Charter, and H. G. Moser. 2002d. Ichthyoplankton and station data for Manta (surface) tows taken on California Cooperative Oceanic Fisheries Investigations survey cruises in 2000. U.S. Dep. Commer., NOAA Tech. Memo. NOAA-TM-NMFS-SWFSC-336. 40 pp.
- Wisner, R. L. 1976. The taxonomy and distribution of lanternfishes (family Myctophidae) of the eastern Pacific Ocean. U.S. Government Printing Office, Washington, D.C. 229 pp.
- Zahuranec, B. J. 2000. Zoogeography and systematics of the lanternfishes of the genus *Nannobrachium* (Myctophidae: Lampanyctini). Smithson. Contrib. Zool. 607. 69 pp.
- Zaitsev, Y. P. 1970. Marine neustonology. Naukova Dumka. Kiev. 264 pp.[In Russian]. [English transl.: 1971. Israel Progr. Sci. Transl. No. 5976. 207 pp.]

Table 1. List of ichthyoplankton data reports for Manta (surface) net samples from CalCOFI surveys for 1977 to 2000. Citations for each report are included in the Literature Cited section of this atlas.

Survey Year	Senior Author	Publication Year
1977-78	Moser et al.	2001
1980-81	Ambrose et. al.	2002a
1984	Charter et al.	2002a
1985	Ambrose et al.	2002b
1986	Charter et al.	2002b
1987	Sandknop et al.	2002a
1988	Watson et al.	2002a
1989	Ambrose et al.	2002c
1990	Charter et al.	2002c
1991	Sandknop et al.	2002b
1992	Watson et al.	2002b
1993	Ambrose et al.	2002d
1994	Charter et al.	2002d
1995	Sandknop et al.	2002c
1996	Watson et al.	2002c
1997	Ambrose et al.	2002e
1998	Ambrose et al.	2002f
1999	Ambrose et al.	2002g
2000	Watson et al.	2002d

Table 2. Total stations within the current CalCOFI survey pattern used in this atlas, listed by month and year.

Year	Month												Total
	1	2	3	4	5	6	7	8	9	10	11	12	
1977													2
1978	4	4	11	31	33	15	19	50					167
1980											29	24	53
1981	48	32	7	29	48		47						211
1984	61	42	19	50	27	29	63			47			338
1985		42	4	1	39			55			47		188
1986		54			50				51	2	50		207
1987			60	3	63				62		45		233
1988	58	4		10	42			65		66			245
1989	51	10		63	12		53	5			66		260
1990			56	47			31	34			65		233
1991	60	12	42				36	29	13	50			242
1992	17	43		65			66		22	42			255
1993	65		9	57				65			66		262
1994	44	22	38	27				66	4	60			261
1995	65			51			66			66			248
1996	10	56		60	2			66		61	4		259
1997	13	53		61			66		46	12		9	260
1998	38	26		66			66		66				262
1999	65			61				65		64			255
2000	66			66		10	56			65			263
Total	665	400	246	748	316	54	569	500	264	601	306	35	4704

Table 3. Total stations within the current CalCOFI survey pattern used in this atlas, listed by line and station.

Station	76.7	80.0	82.0	Line 83.3	86.7	90.0	93.3	Total
27							76	76
28						81	81	162
30						81	81	162
33					75			75
35					80	71	80	231
37						80		80
40					80		79	159
40.6				79				79
42				80				80
45					77	78	77	232
46.9			74					74
49	65							65
50		74	79			72		78
51	74			72				225
53							76	76
55	72	76		72	71		77	368
60	77	80		72	73	76	75	453
70	65	75		68	69	72	75	424
80	64	72		62	65	77	73	413
90	63	71		63	66	74	73	410
100	55	58		63	59	72	65	372
110				50	49	61	60	220
120						59	59	118
Total	535	511	74	681	836	958	1109	4704

Table 4. Ranked abundance of larval fish taxa taken in Manta net tows on CalCOFI surveys from 1977 to 2000. The total for each taxon is the summed raw count for all recorded occurrences.

Rank	Taxon	Total	Rank	Taxon	Total
1	<i>Engraulis mordax</i>	149592	44	<i>Citharichthys sordidus</i>	75
2	<i>Sardinops sagax</i>	27247	46	<i>Pleuronichthys verticalis</i>	71
3	<i>Cololabis saira</i>	7459	47	<i>Icichthys lockingtoni</i>	66
4	<i>Hypsoblennius jenkinsi</i>	6155	48	<i>Neoclinus blanchardi</i>	64
5	<i>Scomber japonicus</i>	5633	49	<i>Oxylebius pictus</i>	62
6	<i>Sebastes</i> spp.	5348	50	<i>Anisotremus davidsoni</i>	60
7	Scorpaenidae	3791	51	<i>Nannobrachium</i> spp.	52
8	<i>Merluccius productus</i>	3501	52	Unidentified fish larvae	49
9	<i>Hexagrammos decagrammus</i>	2190	52	<i>Stomias atriventer</i>	49
10	<i>Chromis punctipinnis</i>	2155	52	Cottidae	49
11	<i>Atherinopsis californiensis</i>	1996	55	<i>Aristostomias scintillans</i>	42
12	<i>Scorpaenichthys marmoratus</i>	1868	55	<i>Nannobrachium ritteri</i>	42
13	<i>Trachurus symmetricus</i>	1526	57	<i>Sebastes aurora</i>	39
14	<i>Leuresthes tenuis</i>	1384	58	<i>Halichoeres semicinctus</i>	38
15	<i>Vinciguerria lucetia</i>	1086	58	<i>Semicossyphus pulcher</i>	38
16	<i>Sebastes jordani</i>	683	60	<i>Peprilus simillimus</i>	37
17	<i>Oxyjulis californica</i>	642	61	<i>Cyclothona</i> spp.	32
18	<i>Sphyraena argentea</i>	620	61	<i>Anoplopoma fimbria</i>	32
19	<i>Hypsoblennius gilberti</i>	592	63	<i>Atherinops affinis</i>	30
20	<i>Sebastes diploproa</i>	560	63	<i>Neoclinus stephensae</i>	30
21	<i>Ceratoscopelus townsendi</i>	506	63	<i>Coryphopterus nicholsii</i>	30
22	<i>Scorpaena guttata</i>	463	66	<i>Sebastes paucispinis</i>	26
23	<i>Xenistius californiensis</i>	398	67	<i>Macroramphosus gracilis</i>	25
24	<i>Paralabrax</i> spp.	389	68	<i>Fodiator acutus</i>	24
25	<i>Girella nigricans</i>	360	69	<i>Gigantactis</i> spp.	20
26	<i>Hypsoblennius</i> spp.	307	70	<i>Bathylagus ochotensis</i>	19
27	<i>Genyonemus lineatus</i>	291	70	<i>Pleuronichthys decurrens</i>	19
28	<i>Ophiodon elongatus</i>	255	72	<i>Hermosilla azurea</i>	18
29	<i>Stenobrachius leucopsarus</i>	229	72	<i>Neoclinus</i> spp.	18
30	<i>Medialuna californiensis</i>	223	74	<i>Diaphus</i> spp.	17
31	<i>Tetragonurus cuvieri</i>	187	74	<i>Raihubunella allenii</i>	17
32	<i>Pleuronichthys coenosus</i>	185	74	<i>Ophidion scrippsae</i>	17
33	<i>Citharichthys stigmaeus</i>	158	77	<i>Symbolophorus californiensis</i>	16
34	<i>Triphoturus mexicanus</i>	151	77	Atherinidae	16
35	<i>Seriphus politus</i>	142	79	<i>Bolinichthys longipes</i>	15
36	Disintegrated fish larvae	123	80	<i>Hemilepidotus spinosus</i>	14
37	<i>Cheilopogon heterurus</i>	119	81	<i>Synodus lucioceps</i>	13
38	<i>Hypsoblennius gentilis</i>	116	81	<i>Diogenichthys atlanticus</i>	13
39	<i>Hypsypops rubicundus</i>	102	81	<i>Leuroglossus stilbius</i>	13
40	<i>Cheilopogon pinnatibarbatus</i>	99	84	<i>Protomyctophum crockeri</i>	12
41	<i>Cheilopogon</i> spp.	88	84	<i>Citharichthys</i> spp.	12
42	<i>Lampadenia urophaos</i>	81	84	<i>Bathophilus flemingi</i>	12
43	<i>Paralichthys californicus</i>	76	87	<i>Nannobrachium regale</i>	11
44	<i>Cyclothona signata</i>	75	87	<i>Icosteus aenigmaticus</i>	11

Rank	Taxon	Total	Rank	Taxon	Total
87	<i>Rathbunella</i> spp.	11	126	<i>Sarda chiliensis</i>	4
87	<i>Cryptotrema corallinum</i>	11	137	<i>Myctophum nitidulum</i>	3
91	<i>Tactostoma macropus</i>	10	137	<i>Clinocottus analis</i>	3
91	<i>Parophrys vetulus</i>	10	137	<i>Lythrypnus dalli</i>	3
93	<i>Pleuronichthys ritteri</i>	9	140	<i>Taaningichthys minimus</i>	2
93	<i>Oneirodes</i> spp.	9	140	<i>Argyropelecus sladeni</i>	2
93	<i>Hippoglossina stomata</i>	9	140	<i>Artedius harringtoni</i>	2
93	<i>Brosmophycis marginata</i>	9	140	<i>Icelinus</i> spp.	2
97	Clupeidae	8	140	Labrisomidae	2
97	<i>Syphurus atricaudus</i>	8	140	<i>Oligocottus</i> spp.	2
97	<i>Typhlogobius californiensis</i>	8	140	<i>Cyclothona pseudopallida</i>	2
97	<i>Odontopyxis trispinosa</i>	8	140	<i>Notoscopelus resplendens</i>	2
97	<i>Hexagrammos lagocephalus</i>	8	140	<i>Diplophos taenia</i>	2
97	<i>Sebastolobus</i> spp.	8	149	<i>Stemonosudis macrura</i>	1
97	Kyphosidae	8	149	<i>Umbrina roncador</i>	1
97	<i>Atractoscion nobilis</i>	8	149	<i>Roncador stearnsii</i>	1
97	<i>Seriola lalandi</i>	8	149	<i>Diplospinus multistriatus</i>	1
97	<i>Hirundichthys marginatus</i>	8	149	<i>Coryphaena hippurus</i>	1
107	<i>Etrumeus teres</i>	7	149	<i>Pteraclis aesticola</i>	1
107	<i>Cyclothona acclinidens</i>	7	149	<i>Psenes pellucidus</i>	1
107	<i>Howella</i> spp.	7	149	Carangidae	1
107	<i>Hygophum reinhardtii</i>	7	149	<i>Scopelogadus bispinosus</i>	1
107	<i>Liparis mucosus</i>	7	149	Trachipteridae	1
112	<i>Bathylagus wesethi</i>	6	149	<i>Zaniolepis frenata</i>	1
112	<i>Microstomus pacificus</i>	6	149	<i>Lestidiops ringens</i>	1
112	<i>Sebastes levius</i>	6	149	<i>Parvilux ingens</i>	1
112	<i>Sternoptyx</i> spp.	6	149	<i>Nannobrachium hawaiiensis</i>	1
116	<i>Cheilotrema saturnum</i>	5	149	<i>Diogenichthys laternatus</i>	1
116	<i>Chiasmodon niger</i>	5	149	<i>Bathypterois</i> spp.	1
116	<i>Hirundichthys</i> spp.	5	149	<i>Scopelosaurus</i> spp.	1
116	<i>Trachipterus altivelis</i>	5	149	<i>Idiacanthus antrostomus</i>	1
116	<i>Desmodema lorum</i>	5	149	<i>Argentina sialis</i>	1
116	<i>Hypsopsetta guttulata</i>	5	149	<i>Melamphaes lugubris</i>	1
116	<i>Vinciguerria poweriae</i>	5	149	<i>Xystreurus liolepis</i>	1
116	<i>Leptocottus armatus</i>	5	149	Stomiiformes	1
116	<i>Tarletonbeania crenularis</i>	5	149	<i>Dolichopteryx longipes</i>	1
116	<i>Ruscarius creaseri</i>	5	149	<i>Lampanyctus bristori</i>	1
126	<i>Menticirrhus undulatus</i>	4	149	<i>Caulophryne pelagica</i>	1
126	Myctophidae	4	149	<i>Caulophryne</i> spp.	1
126	<i>Brama japonica</i>	4	149	<i>Antennarius avalonis</i>	1
126	Sciaenidae	4	149	<i>Pleuronichthys</i> spp.	1
126	<i>Sebastes goodei</i>	4	149	<i>Mugil cephalus</i>	1
126	<i>Artedius lateralis</i>	4	149	<i>Glyptocephalus zachirus</i>	1
126	<i>Lepidogobius lepidus</i>	4	149	<i>Sebastolobus altivelis</i>	1
126	<i>Chilara taylori</i>	4	149	<i>Cataetyx rubrirostris</i>	1
126	<i>Lyopsetta exilis</i>	4	149	Stichaeidae	1
126	Exocoetidae	4	149	Clinidae	1

Rank	Taxon	Total	Rank	Taxon	Total
149	Gobiidae	1	149	Cyclopteridae	1
149	<i>Liparis fucensis</i>	1	149	<i>Ruscarius meanyi</i>	1
149	<i>Liparis florae</i>	1	149	<i>Lepidopsetta bilineata</i>	1

Table 5. Ranked occurrence of larval fish taxa taken in Manta net tows on CalCOFI surveys from 1977 to 2000. The total for each taxon is the sum of all recorded occurrences.

Rank	Taxon	Total	Rank	Taxon	Total
1	<i>Cololabis saira</i>	1421	44	<i>Sebastes aurora</i>	34
2	<i>Engraulis mordax</i>	1365	45	Disintegrated fish larvae	32
3	<i>Sebastes</i> spp.	720	46	<i>Ophiodon elongatus</i>	30
4	<i>Sardinops sagax</i>	647	46	<i>Neoclinus blanchardi</i>	30
5	<i>Hypsoblennius jenkinsi</i>	349	46	<i>Seriphis politus</i>	30
6	<i>Scomber japonicus</i>	321	49	<i>Cheilopogon pinnatibarbatus</i>	28
6	<i>Scorpaenichthys marmoratus</i>	321	50	<i>Stomias atriventer</i>	26
8	<i>Trachurus symmetricus</i>	299	51	Unidentified fish larvae	25
9	<i>Vinciguerria lucetia</i>	262	52	<i>Cyclothona</i> spp.	22
10	<i>Sebastes diploproa</i>	188	52	<i>Coryphopterus nicholsii</i>	22
11	<i>Chromis punctipinnis</i>	183	54	<i>Semicossyphus pulcher</i>	21
12	<i>Ceratoscopelus townsendi</i>	172	54	<i>Neoclinus stephensae</i>	21
13	<i>Oxyjulis californica</i>	168	56	<i>Peprilus simillimus</i>	20
14	<i>Atherinopsis californiensis</i>	161	57	<i>Halichoeres semicinctus</i>	19
15	<i>Merluccius productus</i>	127	58	<i>Gigantactis</i> spp.	18
16	<i>Tetragonurus cuvieri</i>	122	59	<i>Pleuronichthys decurrens</i>	17
17	<i>Medialuna californiensis</i>	111	59	<i>Xenistius californiensis</i>	17
18	<i>Hypsoblennius gilberti</i>	106	59	<i>Anoplopoma fimbria</i>	17
19	<i>Sphyraena argentea</i>	97	62	<i>Diaphus</i> spp.	16
20	<i>Stenobrachius leucopsarus</i>	95	63	<i>Atherinops affinis</i>	15
21	<i>Leuresthes tenuis</i>	90	63	<i>Fodiator acutus</i>	15
22	<i>Pleuronichthys coenosus</i>	89	63	<i>Sebastes paucispinis</i>	15
23	<i>Triphoturus mexicanus</i>	84	63	<i>Macroramphosus gracilis</i>	15
24	<i>Sebastes jordani</i>	80	67	<i>Diogenichthys atlanticus</i>	13
24	<i>Girella nigricans</i>	80	67	<i>Symbolophorus californiensis</i>	13
26	<i>Paralabrax</i> spp.	60	69	<i>Protomyctophum crockeri</i>	12
27	<i>Citharichthys stigmaeus</i>	58	70	<i>Bathophilus Flemingi</i>	11
28	<i>Icichthys lockingtoni</i>	56	70	<i>Hypsypops rubicundus</i>	11
29	<i>Cyclothona signata</i>	54	70	<i>Nannobrachium regale</i>	11
29	<i>Hexagrammos decagrammus</i>	54	73	<i>Anisotremus davidsoni</i>	10
31	<i>Citharichthys sordidus</i>	50	74	<i>Tactostoma macropus</i>	9
32	<i>Cheilopogon heterurus</i>	49	74	<i>Icosteus aenigmaticus</i>	9
33	<i>Genyonemus lineatus</i>	43	74	<i>Ophidion scrippsae</i>	9
34	<i>Nannobrachium ritteri</i>	39	74	<i>Hippoglossina stomata</i>	9
35	<i>Lampadena urophaos</i>	38	74	<i>Oneirodes</i> spp.	9
35	<i>Nannobrachium</i> spp.	38	79	<i>Parophrys vetulus</i>	8
37	<i>Pleuronichthys verticalis</i>	37	79	<i>Pleuronichthys ritteri</i>	8
37	<i>Hypsoblennius</i> spp.	37	79	<i>Synodus lucioceps</i>	8
39	<i>Cheilopogon</i> spp.	36	79	<i>Seriola lalandi</i>	8
39	<i>Oxylebius pictus</i>	36	79	<i>Hirundichthys marginatus</i>	8
41	<i>Aristostomias scintillans</i>	35	79	<i>Typhlogobius californiensis</i>	8
41	<i>Paralichthys californicus</i>	35	85	<i>Bathylagus ochotensis</i>	7
41	<i>Hypsoblennius gentilis</i>	35	85	<i>Leuroglossus stilbius</i>	7

Rank	Taxon	Total	Rank	Taxon	Total
85	<i>Cyclothona acclinidens</i>	7	132	Kyphosidae	2
85	<i>Hemilepidotus spinosus</i>	7	132	<i>Argyropelecus sladeni</i>	2
85	Cottidae	7	132	<i>Icelinus</i> spp.	2
85	<i>Hermosilla azurea</i>	7	132	<i>Lythrypnus dalli</i>	2
85	<i>Howella</i> spp.	7	132	<i>Oligocottus</i> spp.	2
85	<i>Hygophum reinhardtii</i>	7	132	<i>Vinciguerria poweriae</i>	2
93	<i>Sebastes levis</i>	6	132	<i>Bolinichthys longipes</i>	2
93	<i>Atractoscion nobilis</i>	6	140	<i>Mugil cephalus</i>	1
93	<i>Bathylagus wesethi</i>	6	140	<i>Parvilux ingens</i>	1
93	<i>Neoclinus</i> spp.	6	140	<i>Lestidiops ringens</i>	1
97	<i>Brosmophycis marginata</i>	5	140	<i>Stemonosudis macrura</i>	1
97	<i>Hypsopsetta guttulata</i>	5	140	<i>Nannobrachium hawaiiensis</i>	1
97	<i>Syphurus atricaudus</i>	5	140	Scorpaenidae	1
97	<i>Microstomus pacificus</i>	5	140	<i>Sebastolobus altivelis</i>	1
97	<i>Rathbunella</i> spp.	5	140	<i>Lampanyctus bristori</i>	1
97	<i>Desmodema lorum</i>	5	140	<i>Ruscarius meanyi</i>	1
97	<i>Liparis mucosus</i>	5	140	<i>Diplophos taenia</i>	1
97	<i>Trachipterus altivelis</i>	5	140	<i>Artedius harringtoni</i>	1
97	<i>Citharichthys</i> spp.	5	140	<i>Zaniolepis frenata</i>	1
97	<i>Chiasmodon niger</i>	5	140	<i>Taaningichthys minimus</i>	1
107	<i>Tarletonbeania crenularis</i>	4	140	<i>Diogenichthys laternatus</i>	1
107	<i>Leptocottus armatus</i>	4	140	<i>Bathypterois</i> spp.	1
107	<i>Hexagrammos lagocephalus</i>	4	140	<i>Scopelogadus bispinosus</i>	1
107	<i>Sebastes goodei</i>	4	140	<i>Idiacanthus antrostomus</i>	1
107	<i>Scorpaena guttata</i>	4	140	<i>Umbrina roncador</i>	1
107	<i>Cheilotrema saturnum</i>	4	140	<i>Cyclothona pseudopallida</i>	1
107	<i>Sarda chiliensis</i>	4	140	<i>Scopelosaurus</i> spp.	1
107	<i>Ruscarius creaseri</i>	4	140	Stomiiformes	1
107	<i>Lyopsetta exilis</i>	4	140	<i>Dolichopteryx longipes</i>	1
107	Sciaenidae	4	140	<i>Artedius lateralis</i>	1
107	<i>Odontopyxis trispinosa</i>	4	140	<i>Melamphaes lugubris</i>	1
107	<i>Lepidogobius lepidus</i>	4	140	<i>Pleuronichthys</i> spp.	1
107	<i>Cryptotrema corallinum</i>	4	140	<i>Etrumeus teres</i>	1
107	<i>Chilara taylori</i>	4	140	<i>Argentina sialis</i>	1
107	Myctophidae	4	140	<i>Roncador stearnsii</i>	1
107	<i>Hirundichthys</i> spp.	4	140	<i>Coryphaena hippurus</i>	1
107	<i>Sternoptyx</i> spp.	4	140	<i>Pteraclis aesticola</i>	1
107	Clupeidae	4	140	<i>Psenes pellucidus</i>	1
107	<i>Rathbunella alleni</i>	4	140	Carangidae	1
126	<i>Sebastolobus</i> spp.	3	140	Atherinidae	1
126	<i>Clinocottus analis</i>	3	140	Trachipteridae	1
126	<i>Myctophum nitidulum</i>	3	140	<i>Caulophryne pelagica</i>	1
126	Exocoetidae	3	140	<i>Diplospinus multistriatus</i>	1
126	<i>Brama japonica</i>	3	140	<i>Antennarius avalonis</i>	1
126	<i>Menticirrhus undulatus</i>	3	140	Cyclopteridae	1
132	<i>Notoscopelus resplendens</i>	2	140	<i>Lepidopsetta bilineata</i>	1

Rank	Taxon	Total	Rank	Taxon	Total
140	<i>Glyptocephalus zachirus</i>	1	140	Labrisomidae	1
140	<i>Xystreurus liolepis</i>	1	140	Gobiidae	1
140	<i>Cataetyx rubrirostris</i>	1	140	<i>Liparis fucensis</i>	1
140	Stichaeidae	1	140	<i>Liparis florae</i>	1
140	Clinidae	1	140	<i>Caulophryne</i> spp.	1

Table 6. Total counts of fifty most abundant larval fish from CalCOFI Manta net catches, 1977-2000, compared with standardized total counts of the same taxa from concurrent oblique net catches.

Taxon	Family	Manta		Oblique	
		Rank	Total	Rank	Total
<i>Engraulis mordax</i>	Engraulidae	1	149,592	1	1,182,698.5
<i>Sardinops sagax</i>	Clupeidae	2	27,247	6	120,223.8
<i>Cololabis saira</i>	Scomberesocidae	3	7,459	87	404.5
<i>Hypsoblennius jenkinsi</i>	Blenniidae	4	6,155	71	824.9
<i>Scomber japonicus</i>	Scombridae	5	5,633	15	16,564.5
<i>Sebastes</i> spp.	Sebastidae	6	5,348	4	128,896.8
<i>Merluccius productus</i>	Merlucciidae	7	3,501	2	439,873.4
<i>Hexagrammos decagrammus</i>	Hexagrammidae	8	2,190	197	28.0
<i>Chromis punctipinnis</i>	Pomacentridae	9	2,155	50	1,560.0
<i>Atherinopsis californiensis</i>	Atherinidae	10	1,996	162	70.0
<i>Scorpaenichthys marmoratus</i>	Cottidae	11	1,868	105	256.5
<i>Trachurus symmetricus</i>	Carangidae	12	1,526	10	21,414.7
<i>Leuresthes tenuis</i>	Atherinidae	13	1,384	215	19.2
<i>Vinciguerria lucetia</i>	Phosichthyidae	14	1,086	3	216,998.1
<i>Sebastes jordani</i>	Sebastidae	15	683	9	23,546.5
<i>Oxyjulis californica</i>	Labridae	16	642	33	3,990.3
<i>Sphyraena argentea</i>	Sphyraenidae	17	620	54	1,484.6
<i>Hypsoblennius gilberti</i>	Blenniidae	18	592	193	30.2
<i>Sebastes diploproa</i>	Sebastidae	19	560	60	1,121.6
<i>Ceratoscopelus townsendi</i>	Myctophidae	20	506	13	20,504.3
<i>Scorpaena guttata</i>	Scorpaenidae	21	463	210	20.5
<i>Xenistius californiensis</i>	Haemulidae	22	398	161	70.3
<i>Paralabrax</i> spp.	Serranidae	23	389	40	2,153.3
<i>Girella nigricans</i>	Kyphosidae	24	360	168	65.4
<i>Hypsoblennius</i> spp.	Blenniidae	25	307	61	1,091.1

Taxon	Family	Manta		Oblique	
		Rank	Total	Rank	Total
<i>Genyonemus lineatus</i>	Sciaenidae	26	291	18	14,236.6
<i>Ophiodon elongatus</i>	Hexagrammidae	27	255	160	75.4
<i>Stenobrachius leucopsarus</i>	Myctophidae	28	229	7	87,912.6
<i>Medialuna californiensis</i>	Kyphosidae	29	223	163	69.1
<i>Tetragonurus cuvieri</i>	Tetragonuridae	30	187	43	1,931.4
<i>Pleuronichthys coenosus</i>	Pleuronectidae	31	185	123	163.9
<i>Citharichthys stigmaeus</i>	Paralichthyidae	32	158	19	13,351.1
<i>Triphoturus mexicanus</i>	Myctophidae	33	151	14	19,847.8
<i>Seriphis politus</i>	Sciaenidae	34	142	44	1,900.8
<i>Cheilopogon heterurus</i>	Exocoetidae	35	119	—	0
<i>Hypsoblennius gentilis</i>	Blenniidae	36	116	150	93.6
<i>Hypsopops rubicundus</i>	Pomacentridae	37	102	234	12.1
<i>Cheilopogon pinnatibarbus</i>	Exocoetidae	38	99	—	0
<i>Cheilopogon</i> spp.	Exocoetidae	39	88	—	0
<i>Lampadена urophaos</i>	Myctophidae	40	81	93	342.8
<i>Paralichthys californicus</i>	Paralichthyidae	41	76	46	1,781.0
<i>Cyclothone signata</i>	Gonostomatidae	42	75	16	15,868.2
<i>Citharichthys sordidus</i>	Paralichthyidae	43	75	21	11,726.9
<i>Pleuronichthys verticalis</i>	Pleuronectidae	44	71	58	1,296.3
<i>Icichthys lockingtoni</i>	Centrolophidae	45	66	48	1,589.5
<i>Neoclinus blanchardi</i>	Chaenopsidae	46	64	255	5.6
<i>Oxylebius pictus</i>	Hexagrammidae	47	62	108	233.0
<i>Anistremus davidsoni</i>	Haemulidae	48	60	190	33.1
<i>Nannobrachium</i> spp.	Myctophidae	49	52	24	9,447.3
<i>Stomias atriventer</i>	Stomiidae	50	49	55	1,369.0
Proportion of Total for All Taxa			0.978		0.865

Table 7. Results of recurrent group analyses of larval fish in Manta and oblique net samples from CalCOFI surveys (current station pattern) during 1977–2000. Each taxon is preceded by its taxonomic code and followed by the affinity index and taxa to which it is linked.

Manta Recurrent Group 1					
31	<i>Engraulis mordax</i>	Has and	0.462 0.334	link to link to	683 736
683	<i>Sebastes</i> spp.	Has	0.339	link to	736
736	<i>Scorpaenichthys marmoratus</i>				
Associates					
853	<i>Hypsoblennius jenkinsi</i>	Has	0.311	link to	31
19	<i>Sardinops sagax</i>	Has	0.362	link to	31
Manta Recurrent Group 2					
486	<i>Sphyraena argentea</i>	Has and	0.316 0.332	link to link to	626 853
626	<i>Chromis punctipinnis</i>	Has	0.381	link to	853
853	<i>Hypsoblennius jenkinsi</i>				
Associates					
574	<i>Scomber japonicus</i>	Has	0.357	link to	486
431	<i>Paralabrax</i> spp.	Has	0.408	link to	486
Manta Recurrent Group 3					
93	<i>Vinciguerria lucetia</i>	Has	0.331	link to	226
226	<i>Ceratoscopelus townsendi</i>				
Manta Recurrent Group 4					
601	<i>Anisotremus davidsoni</i>	Has	0.416	link to	452
452	<i>Xenistius californiensis</i>				
Associates					
628	<i>Hypsopops rubicundus</i>	Has	0.326	link to	601

Table 8. Results of recurrent group analyses of larval fish in oblique net samples from CalCOFI surveys (current station pattern) during 1977-2000. Each taxon is preceded by its taxonomic code and followed by the affinity index and taxa to which it is linked.

Oblique Recurrent Group 1					
72	<i>Leuroglossus stilbius</i>	Has and and and and	0.624 0.615 0.333 0.319 0.579	link to link to link to link to link to	683 901 923 925 72
31	<i>Engraulis mordax</i>	Has and and and and and	0.667 0.537 0.345 0.365 0.537	link to link to link to link to link to	683 901 923 925 901
901	<i>Merluccius productus</i>	Has and	0.356 0.315	link to link to	923 925
683	<i>Sebastes</i> spp.	Has and and	0.546 0.356 0.356	link to link to link to	901 923 925
923	<i>Citharichthys sordidus</i>	Has	0.420	link to	925
925	<i>Citharichthys stigmaeus</i>				
Associates					
68	<i>Bathylagus ochotensis</i>	Has and and and	0.429 0.546 0.588 0.470	link to link to link to link to	31 901 72 683
19	<i>Sardinops sagax</i>	Has and and and	0.352 0.313 0.316 0.315	link to link to link to link to	31 901 72 683
292	<i>Stenobrachius leucopsarus</i>	Has and and and	0.546 0.610 0.660 0.584	link to link to link to link to	31 901 72 683
684	<i>Sebastes jordani</i>	Has and and and	0.398 0.389 0.385 0.426	link to link to link to link to	31 901 72 683
686	<i>Sebastes paucispinis</i>	Has and and	0.318 0.338 0.317	link to link to link to	72 901 683
924	<i>Citharichthys</i> spp.	Has and	0.327 0.306	link to link to	31 683

Oblique Recurrent Group 2

71	<i>Bathylagus wesethi</i>	Has and and and and and and and	0.522 0.560 0.511 0.489 0.346 0.387 0.500	link to link to link to link to link to link to link to	78 93 226 239 261 288 296
78	<i>Cyclothona signata</i>	Has and and and and and and	0.617 0.611 0.558 0.357 0.379 0.489	link to link to link to link to link to link to	93 226 239 261 288 296
93	<i>Vinciguerria lucetia</i>	Has and and and and and	0.627 0.553 0.310 0.407 0.454	link to link to link to link to link to	226 239 261 288 296
226	<i>Ceratoscopelus townsendi</i>	Has and and and and	0.536 0.326 0.345 0.447	link to link to link to link to	239 261 288 296
239	<i>Diogenichthys atlanticus</i>	Has and and	0.366 0.445 0.497	link to link to link to	261 288 296
261	<i>Nannobrachium</i> spp.	Has and	0.367 0.697	link to link to	288 296
288	<i>Protomyctophum crockeri</i>	Has	0.413	link to	296
296	<i>Symbolophorus californiensis</i>				

Associates

68	<i>Bathylagus ochotensis</i>	Has and	0.317 0.417	link to	261 288
72	<i>Leuroglossus stilbius</i>	Has	0.338	link to	288
76	<i>Cyclothona</i> spp.	Has and and	0.346 0.341 0.322	link to link to link to	93 226 239
94	<i>Vinciguerria poweriae</i>	Has and	0.332 0.347	link to link to	78 226
105	<i>Argyropelecus sladeni</i>	Has	0.321	link to	78
107	<i>Sternopyx</i> spp.	Has and	0.324 0.317	link to link to	78 226
161	<i>Idiacanthus antrostomus</i>	Has and and and and	0.397 0.365 0.433 0.396 0.304 0.344	link to link to link to link to link to link to	71 78 93 226 296 239

901	<i>Merluccius productus</i>	Has	0.320	link to	288
311	<i>Lestidiops ringens</i>	Has	0.336	link to	71
		and	0.320	link to	93
252	<i>Hygophum reinhardtii</i>	Has	0.310	link to	78
		and	0.348	link to	226
273	<i>Myctophum nitidulum</i>	Has	0.301	link to	93
		and	0.332	link to	226
260	<i>Nannobrachium ritteri</i>	Has	0.354	link to	71
		and	0.339	link to	78
		and	0.318	link to	296
		and	0.359	link to	239
		and	0.306	link to	261
		and	0.419	link to	288
283	<i>Notoscopelus resplendens</i>	Has	0.318	link to	226
292	<i>Stenobrachius leucopsarus</i>	Has	0.376	link to	288
301	<i>Triphoturus mexicanus</i>	Has	0.381	link to	71
		and	0.333	link to	78
		and	0.444	link to	93
		and	0.338	link to	226
514	<i>Trachurus symmetricus</i>	Has	0.331	link to	71
		and	0.322	link to	296
		and	0.303	link to	261
546	<i>Tetragonurus cuvieri</i>	Has	0.314	link to	78
		and	0.328	link to	93
		and	0.306	link to	226

Oblique Recurrent Group 3

68	<i>Bathylagus ochotensis</i>	Has	0.339	link to	260
		and	0.612	link to	292
260	<i>Nannobrachium ritteri</i>	Has	0.322	link to	292
292	<i>Stenobrachius leucopsarus</i>				

Associates

19	<i>Sardinops sagax</i>	Has	0.327	link to	292
684	<i>Sebastes jordani</i>	Has	0.400	link to	292
686	<i>Sebastes paucispinis</i>	Has	0.310	link to	292
514	<i>Trachurus symmetricus</i>	Has	0.346	link to	260

Oblique Recurrent Group 4

431	<i>Paralabrax</i> spp.	Has	0.374	link to	486
486	<i>Sphyraena argentea</i>				

Oblique Recurrent Group 5

531	<i>Peprilus simillimus</i>	Has	0.354	link to	610
610	<i>Sciaenidae</i>				

Table 9. List of taxa with station and month ranges used in calculating averages for atlas graphs.

Taxon	Stations	Months	Taxon	Stations	Months
<i>Anisotremus davidsoni</i>	27–33	6–8	<i>Neoclinus blanchardi</i>	27–55	1–12
<i>Anoplopoma fimbria</i>	42–90	1–5	<i>Neoclinus stephensae</i>	27–55	1–12
<i>Aristostomias scintillans</i>	53–120	1–12	<i>Ophidion scrippsae</i>	27–60	3–8
<i>Atherinops affinis</i>	27–51	3–11	<i>Ophiodon elongatus</i>	27–55	1–4
<i>Atherinopsis californiensis</i>	27–55	11–7	<i>Oxyjulis californica</i>	27–70	4–10
<i>Atractoscion nobilis</i>	27–55	4–8	<i>Oxylebius pictus</i>	28–90	10–7
<i>Bathophilus flemingi</i>	80–120	1–12	<i>Paralabrax</i> spp.	27–60	6–10
<i>BathyLAGUS ochotensis</i>	50–100	1–5	<i>Paralichthys californicus</i>	27–60	1–10
<i>Bolinichthys longipes</i>	90–120	9–11	<i>Parophrys vetulus</i>	30–60	1–5
<i>Ceratoscopelus townsendi</i>	53–120	1–12	<i>Peprius simillimus</i>	27–70	3–11
<i>Cheilopogon heterurus</i>	27–80	6–10	<i>Pleuronichthys coenosus</i>	27–60	1–12
<i>Cheilopogon pinnatibarbatus</i>	27–53	5–11	<i>Pleuronichthys decurrens</i>	30–70	10–5
<i>Chromis punctipinnis</i>	27–60	7–10	<i>Pleuronichthys verticalis</i>	27–60	1–10
<i>Citharichthys sordidus</i>	28–90	1–12	<i>Protomyctophum crockeri</i>	51–120	1–7
<i>Citharichthys stigmaeus</i>	30–100	1–12	<i>Rathbunella allenii</i>	33–51	2–5
<i>Cololabis saira</i>	27–120	1–12	<i>Sarda chiliensis</i>	27–42	4–8
<i>Cololabis saira</i> eggs	27–120	1–12	<i>Sardinops sagax</i>	27–120	1–12
<i>Coryphopterus nicholsii</i>	33–70	1–9	<i>Sardinops sagax</i> eggs	27–120	1–12
<i>Cryptotrema corallinum</i> ^a	42–51	3–7	<i>Scomber japonicus</i>	27–100	3–10
<i>Cyclothone signata</i>	60–120	1–11	<i>Scorpaena guttata</i>	27–45	7–8
<i>Diaphus</i> spp.	60–120	4–10	<i>Scorpaenichthys marmoratus</i> ^d	27–70	10–5
<i>Diogenichthys atlanticus</i>	45–120	1–12	<i>Sebastes aurora</i>	30–80	1–8
<i>Engraulis mordax</i>	27–120	1–12	<i>Sebastes diploproa</i>	27–100	1–12
<i>Engraulis mordax</i> eggs	27–120	1–12	<i>Sebastes jordani</i>	27–60	1–5
<i>Etrumeus teres</i>	27–51	4–10	<i>Sebastes levius</i>	27–60	1–5
<i>Fodiator acutus</i>	27–60	7–10	<i>Sebastes paucispinis</i>	28–60	1–5
<i>Genyonemus lineatus</i>	27–55	10–3	<i>Sebastes</i> spp.	27–80	1–12
<i>Gigantactis</i> spp.	70–120	7–1	<i>Sebastolobus</i> spp.	55–60	3–7
<i>Girella nigricans</i>	27–60	4–11	<i>Semicossyphus pulcher</i>	40–60	4–10
<i>Halichoeres semicinctus</i>	27–70	4–10	<i>Seriola lalandi</i>	27–60	8–10
<i>Hemilepidotus spinosus</i> ^b	46.9–60	1–3	<i>Seriphis politus</i>	27–51	2–8
<i>Hermosilla azurea</i>	27–40	7–8	<i>Sphyraena argentea</i>	27–55	4–10
<i>Hexagrammos decagrammus</i>	33–60	1–4	<i>Stenobrachius leucopsarus</i>	27–120	12–5
<i>Hexagrammos lagocephalus</i> ^c	33–55	1–3	<i>Stomias atriventer</i>	45–120	1–12
<i>Hypsoblennius gentilis</i>	27–51	4–12	<i>Symbolophorus californiensis</i>	55–120	1–7
<i>Hypsoblennius gilberti</i>	27–60	4–10	<i>Synodus lucioceps</i>	27–60	7–1
<i>Hypsoblennius jenkinsi</i>	27–55	4–11	<i>Tactostoma macropus</i>	50–120	1–10
<i>Hypsypops rubicundus</i>	27–53	6–9	<i>Tetragonurus cuvieri</i>	35–120	1–12
<i>Icichthys lockingtoni</i>	51–110	1–12	Total eggs	27–120	1–12
<i>Icosteus aenigmaticus</i>	55–80	1–5	Total larvae	27–120	1–12
<i>Lampadena urophaos</i>	80–120	7–11	<i>Trachurus symmetricus</i>	27–110	2–10
<i>Leuresthes temnus</i>	27–51	4–8	<i>Triphoturus mexicanus</i>	28–120	3–10
<i>Leuroglossus stibius</i>	33–100	1–4	<i>Vinciguerria lucetia</i>	55–120	1–12
<i>Macroramphosus gracilis</i>	28–70	1–12	<i>Xenistius californiensis</i>	27–51	6–10
<i>Medialuna californiensis</i>	27–100	5–11			
<i>Merluccius productus</i>	28–110	10–4	^a lines 80–83 ^c lines 77–80		
<i>Microstomus pacificus</i>	50–90	3–8	^b lines 77–82 ^d lines 77–87		
<i>Nannobrachium regale</i>	55–120	1–10			
<i>Nannobrachium ritteri</i>	40–120	1–12			

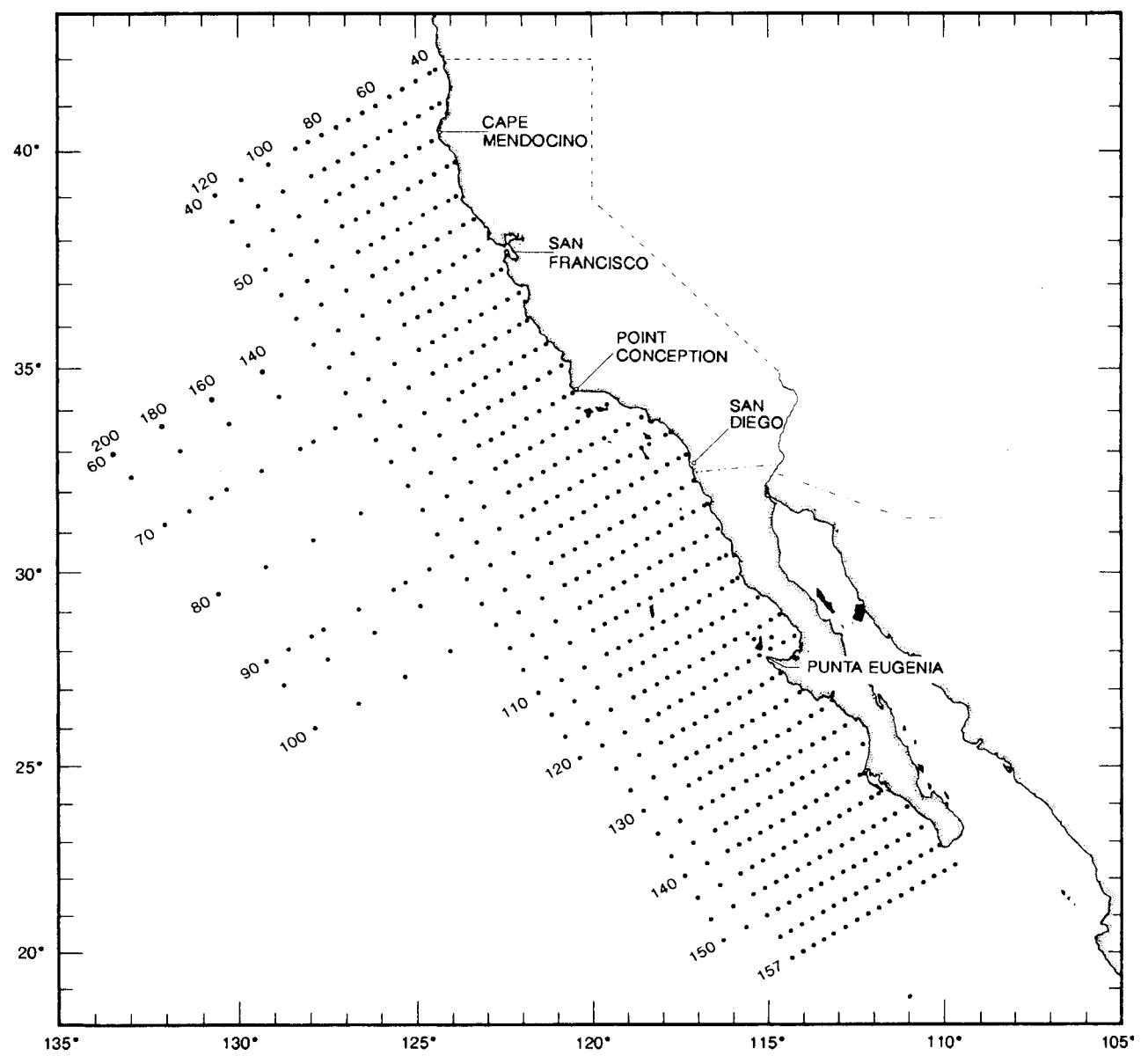


Figure 1. The basic CalCOFI station pattern occupied, in part, by cruises during 1951-1984.

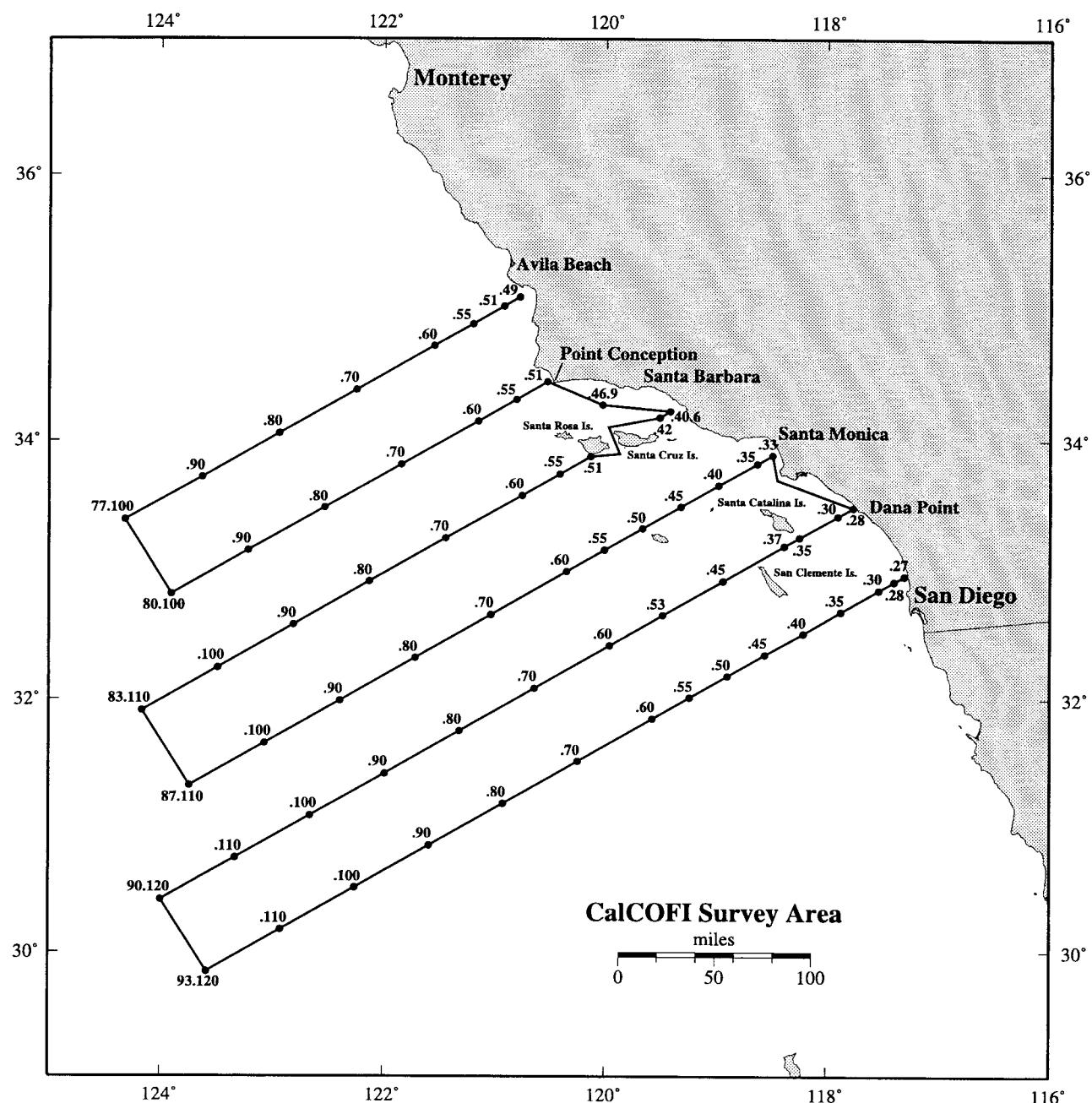


Figure 2. Basic station plan for CalCOFI survey cruises from 1985 to the present.

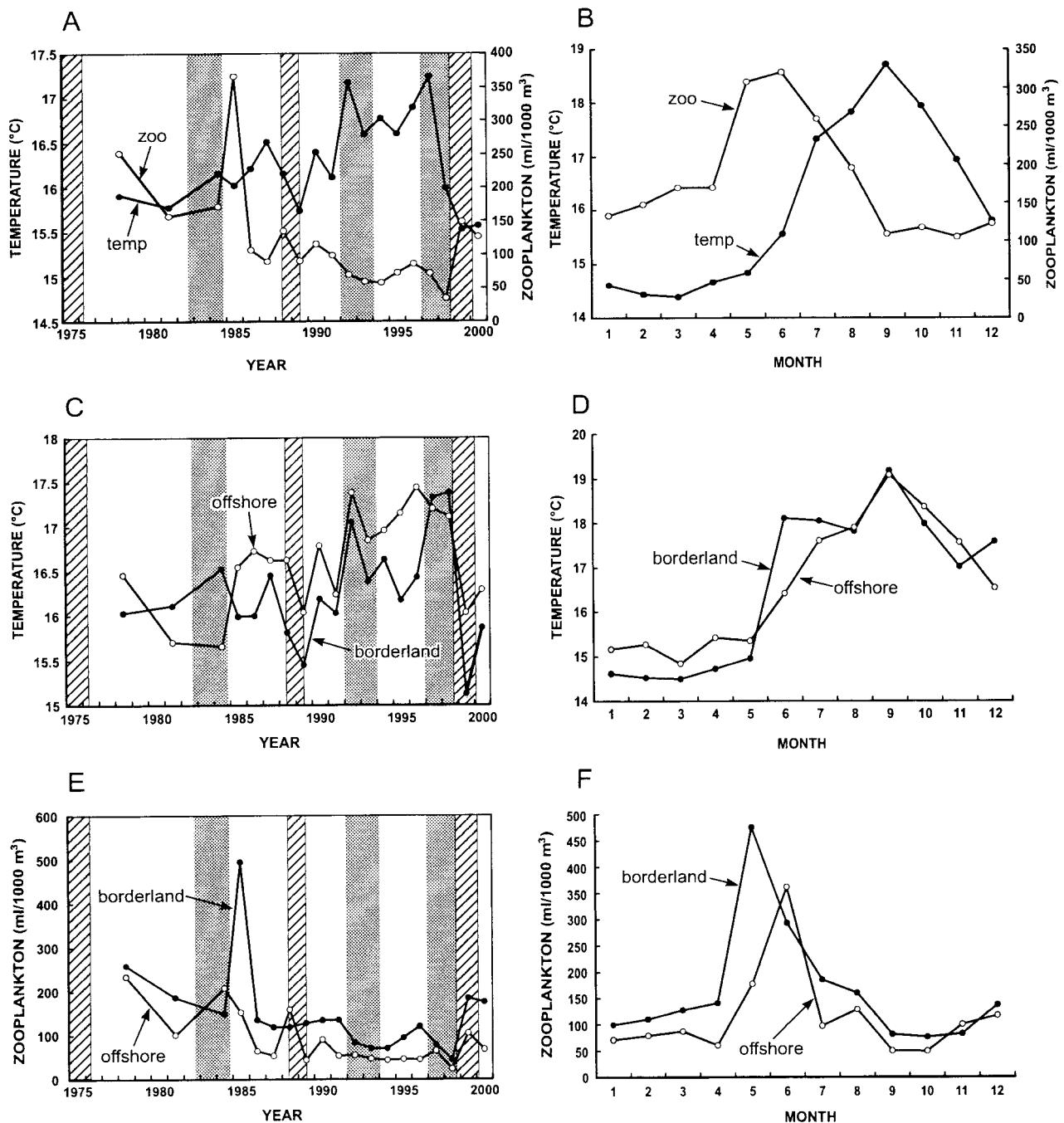


Figure 3. Average seasonal and annual temperatures and zooplankton volumes for CalCOFI stations used in this study. A) Annual temperatures (dots) and zooplankton volumes (circles) during 1978–2000; B) Monthly temperatures (dots) and zooplankton volumes (circles); C) Annual temperatures, borderland vs. offshore regions; D) Monthly temperatures, borderland vs. offshore regions; E) Annual zooplankton volumes, borderland vs. offshore regions; F) Monthly zooplankton volumes, borderland vs. offshore regions. ENSO events are indicated by vertical bars (El Niño, shading; La Niña, hatching.)

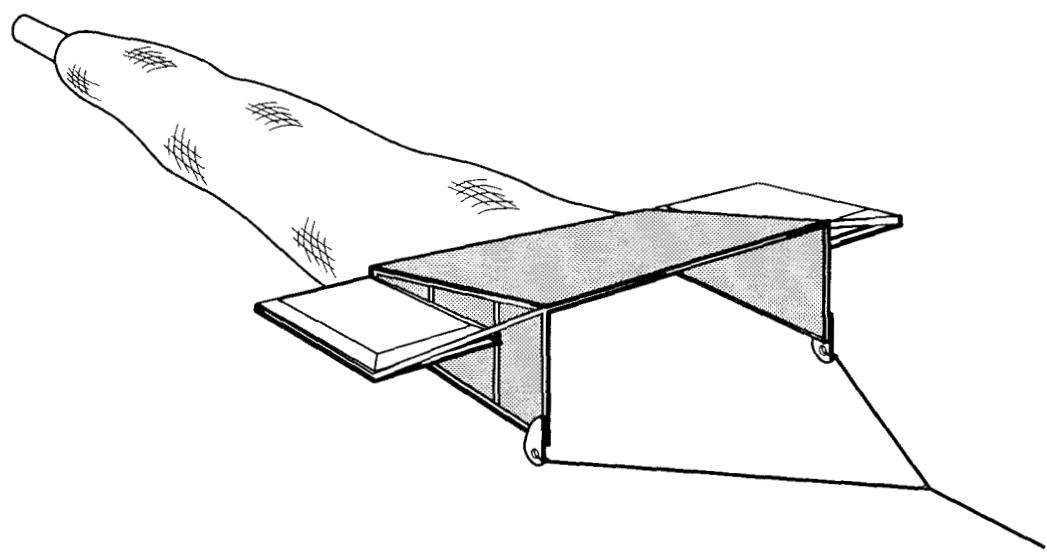
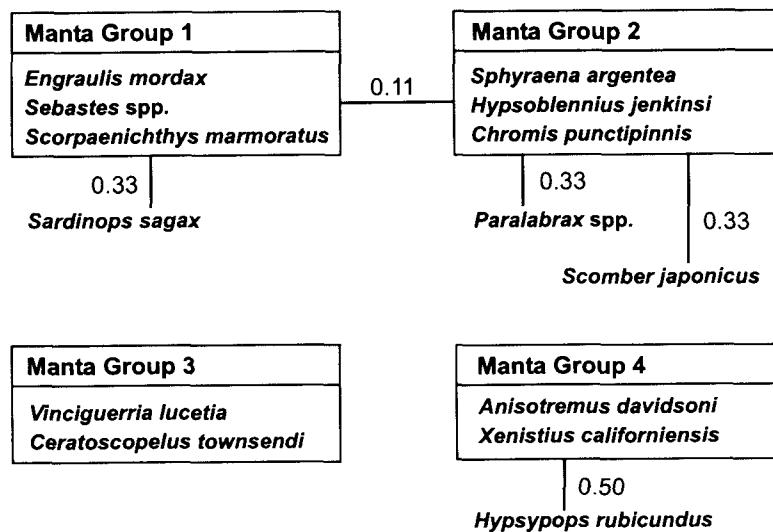


Figure 4. Diagram of the Manta net used on CalCOFI surveys, modified from the original Manta net developed by Brown and Cheng (1981).

A



B

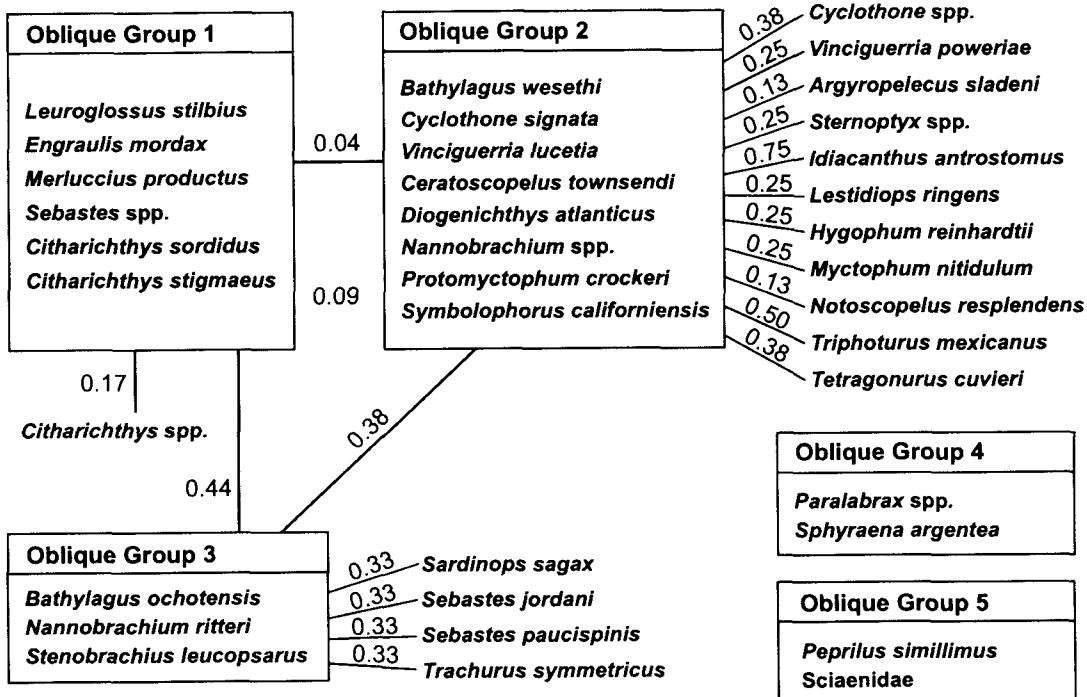


Figure 5. Recurrent groups of fish larvae from Manta net (A) and oblique net (B) samples taken on CalCOFI survey cruises during 1978-2000 in the SCB region. Fractions near lines connecting groups with other groups or with associate taxa indicate the number of significant pairs (affinity indices ≥ 0.3) divided by the number of possible pairs.

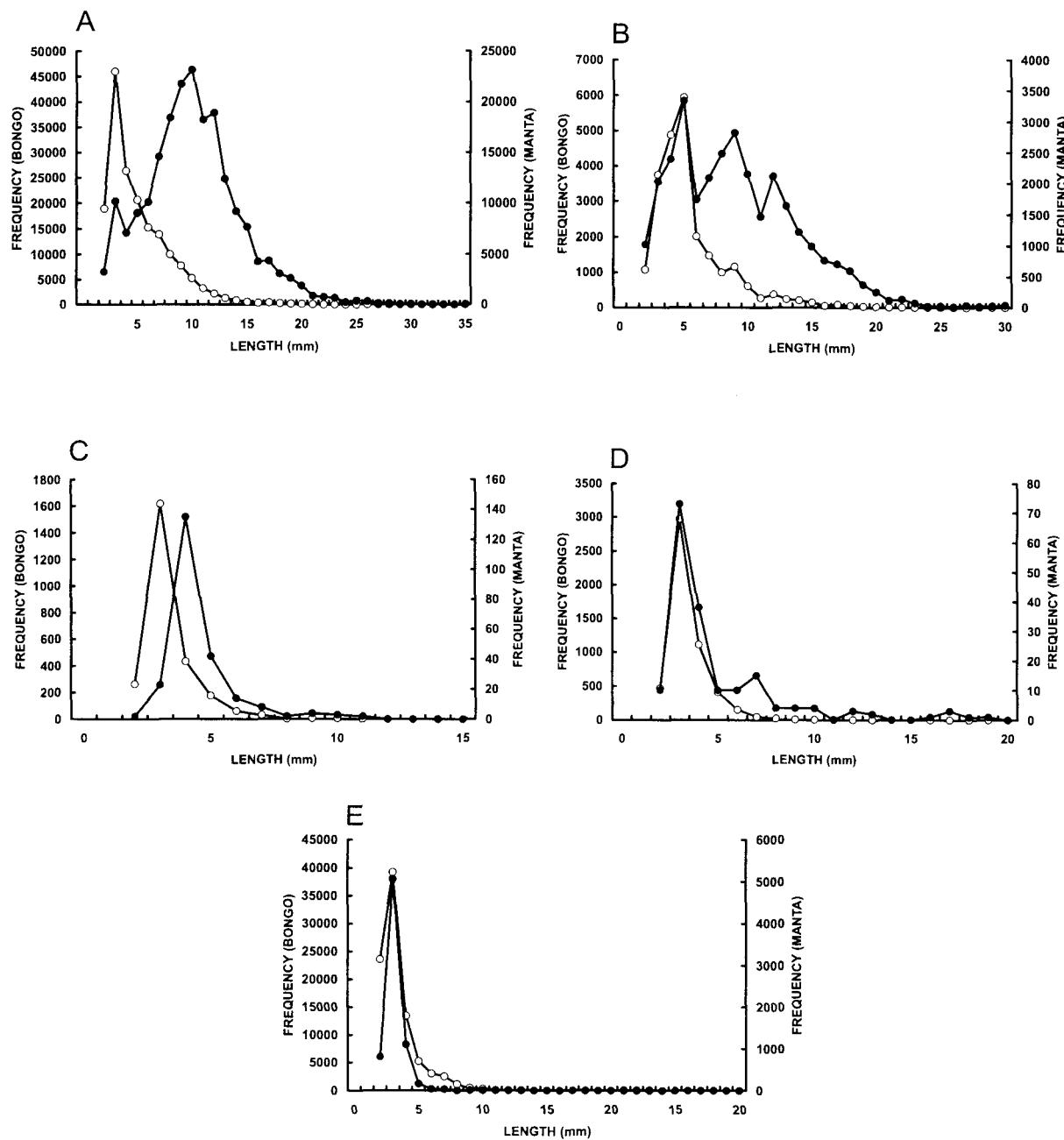


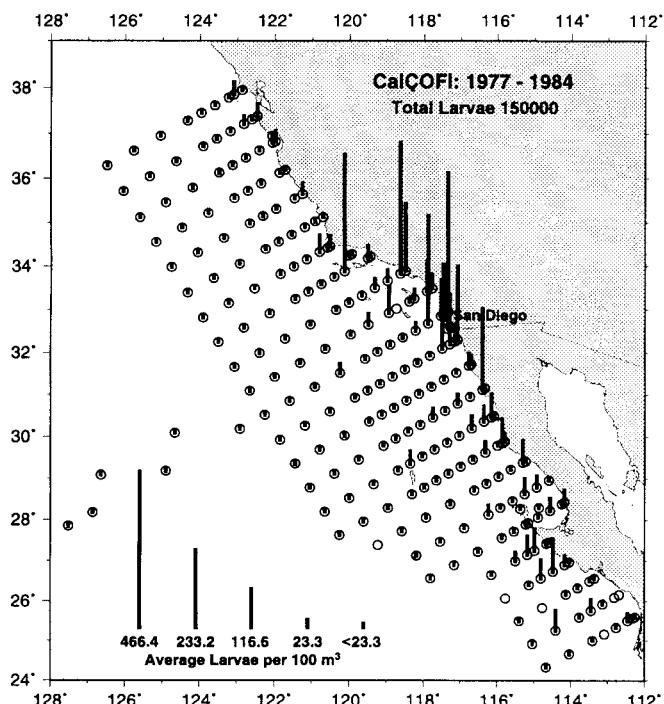
Figure 6. Length frequency distributions for larvae of five fish species taken in Manta (dots) and oblique (circles) net tows on CalCOFI surveys during 1978-2000 in the SCB region: A) northern anchovy *Engraulis mordax*; B) Pacific sardine *Sardinops sagax*; C) chub mackerel *Scomber japonicus*; D) jack mackerel *Trachurus symmetricus*; E) Pacific hake *Merluccius productus*.

List of Taxa

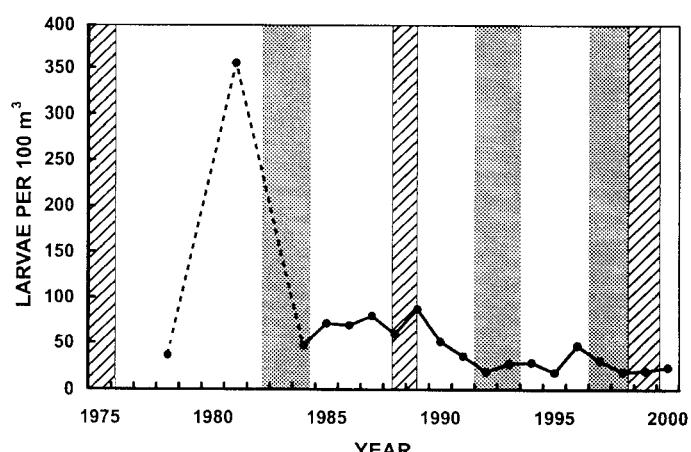
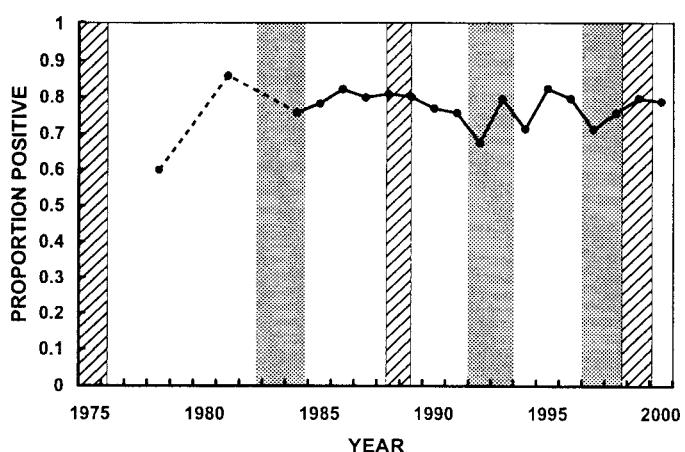
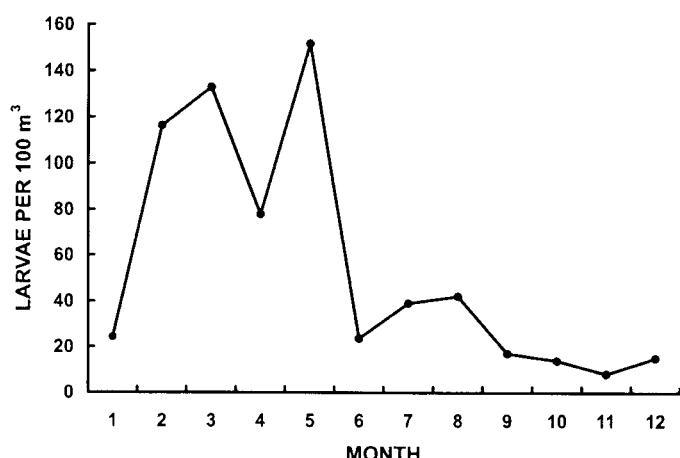
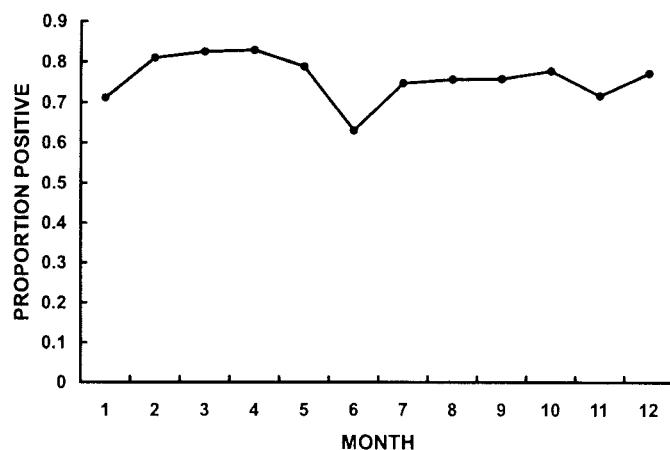
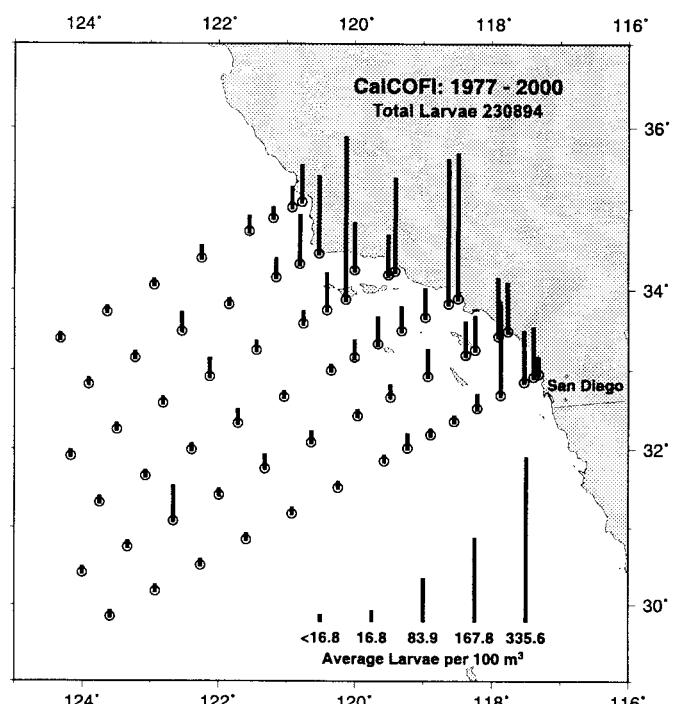
	Page		Page
Total larvae	2	<i>Icichthys lockingtoni</i>	49
Total eggs	3	<i>Neoclinus blanchardi</i>	50
<i>Engraulis mordax</i>	4	<i>Oxylebius pictus</i>	51
<i>Engraulis mordax</i> eggs	5	<i>Anisotremus davidsoni</i>	52
<i>Sardinops sagax</i>	6	<i>Stomias atriventer</i>	53
<i>Sardinops sagax</i> eggs	7	<i>Aristostomias scintillans</i>	54
<i>Cololabis saira</i>	8	<i>Nannobrachium ritteri</i>	55
<i>Cololabis saira</i> eggs	9	<i>Sebastes aurora</i>	56
<i>Hypsoblennius jenkinsi</i>	10	<i>Halichoeres semicinctus</i>	57
<i>Scomber japonicus</i>	11	<i>Semicossyphus pulcher</i>	58
<i>Sebastes</i> spp.	12	<i>Peprilus simillimus</i>	59
<i>Merluccius productus</i>	13	<i>Anoplopoma fimbria</i>	60
<i>Hexagrammos decagrammus</i>	14	<i>Atherinops affinis</i>	61
<i>Chromis punctipinnis</i>	15	<i>Coryphopterus nicholsii</i>	62
<i>Atherinopsis californiensis</i>	16	<i>Neoclinus stephensae</i>	63
<i>Scorpaenichthys marmoratus</i>	17	<i>Sebastes paucispinis</i>	64
<i>Trachurus symmetricus</i>	18	<i>Macroramphosus gracilis</i>	65
<i>Leuresthes tenuis</i>	19	<i>Fodiator acutus</i>	66
<i>Vinciguerria lucetia</i>	20	<i>Gigantactis</i> spp.	67
<i>Sebastes jordani</i>	21	<i>Bathylagus ochotensis</i>	68
<i>Oxyjulis californica</i>	22	<i>Pleuronichthys decurrens</i>	69
<i>Sphyraena argentea</i>	23	<i>Hermosilla azurea</i>	70
<i>Hypsoblennius gilberti</i>	24	<i>Diaphus</i> spp.	71
<i>Sebastes diploproa</i>	25	<i>Rathbunella alleni</i>	72
<i>Ceratoscopelus townsendi</i>	26	<i>Ophidion scrippsae</i>	73
<i>Scorpaena guttata</i>	27	<i>Symbolophorus californiensis</i>	74
<i>Xenistius californiensis</i>	28	<i>Bolinichthys longipes</i>	75
<i>Paralabrax</i> spp.	29	<i>Hemilepidotus spinosus</i>	76
<i>Girella nigricans</i>	30	<i>Leuroglossus stilibius</i>	77
<i>Genyonemus lineatus</i>	31	<i>Synodus lucioceps</i>	78
<i>Ophiodon elongatus</i>	32	<i>Diogenichthys atlanticus</i>	79
<i>Stenobrachius leucopsarus</i>	33	<i>Bathophilus flemingi</i>	80
<i>Medialuna californiensis</i>	34	<i>Protomyctophum crockeri</i>	81
<i>Tetragonurus cuvieri</i>	35	<i>Nannobrachium regale</i>	82
<i>Pleuronichthys coenosus</i>	36	<i>Icosteus aenigmaticus</i>	83
<i>Citharichthys stigmaeus</i>	37	<i>Cryptotrema corallinum</i>	84
<i>Triphoturus mexicanus</i>	38	<i>Tactostoma macropus</i>	85
<i>Seriphus politus</i>	39	<i>Parophrys vetulus</i>	86
<i>Cheilopogon heterurus</i>	40	<i>Sebastolobus</i> spp.	87
<i>Hypsoblennius gentilis</i>	41	<i>Seriola lalandi</i>	88
<i>Hypsypops rubicundus</i>	42	<i>Hexagrammos lagocephalus</i>	89
<i>Cheilopogon pinnatibarbus</i>	43	<i>Atractoscion nobilis</i>	90
<i>Lampadena urophaos</i>	44	<i>Etrumeus teres</i>	91
<i>Paralichthys californicus</i>	45	<i>Microstomus pacificus</i>	92
<i>Cyclothone signata</i>	46	<i>Sebastes levis</i>	93
<i>Citharichthys sordidus</i>	47	<i>Sarda chiliensis</i>	94
<i>Pleuronichthys verticalis</i>	48		

CHARTS AND GRAPHS

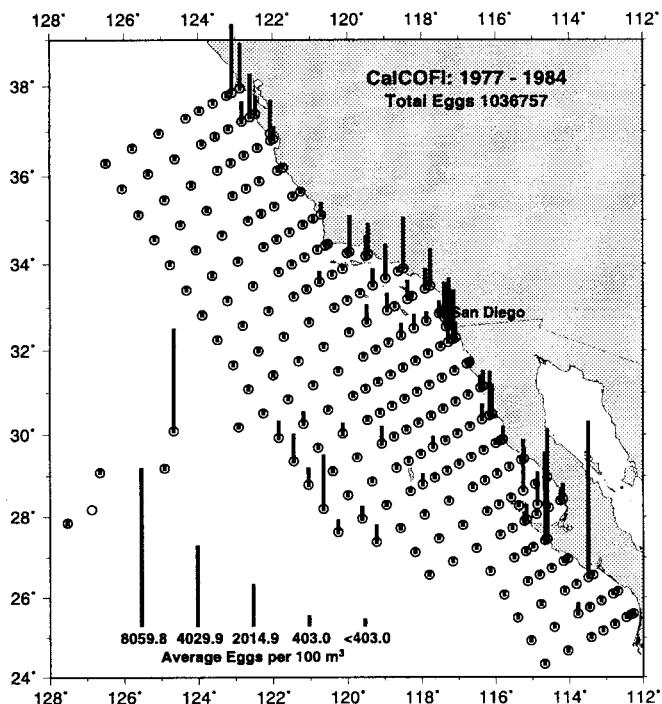
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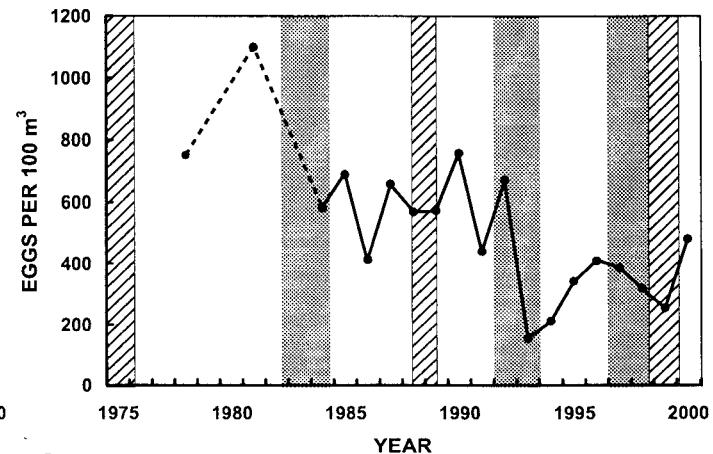
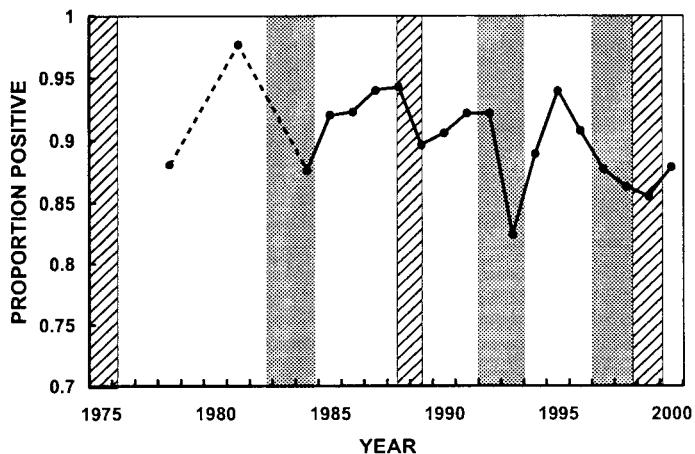
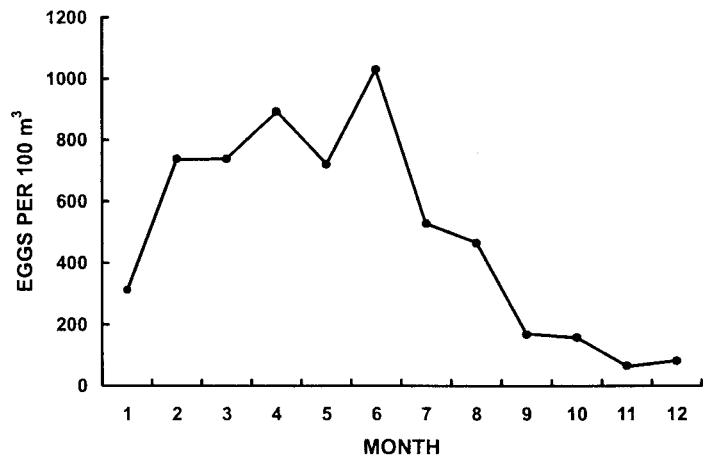
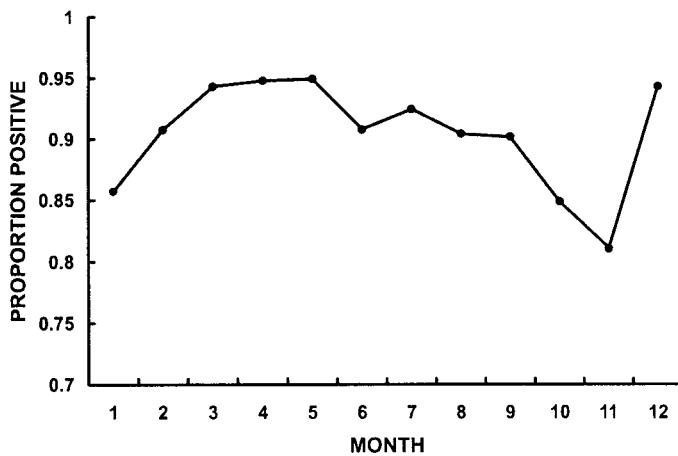
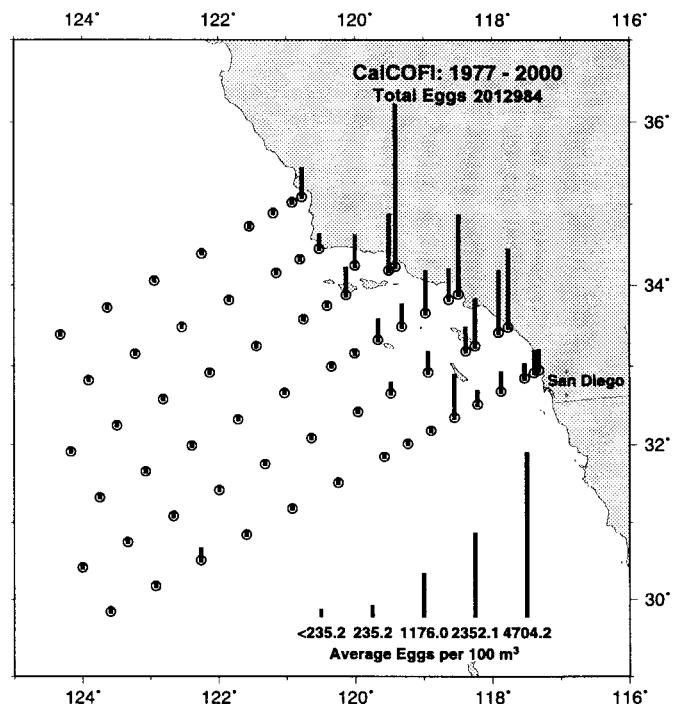
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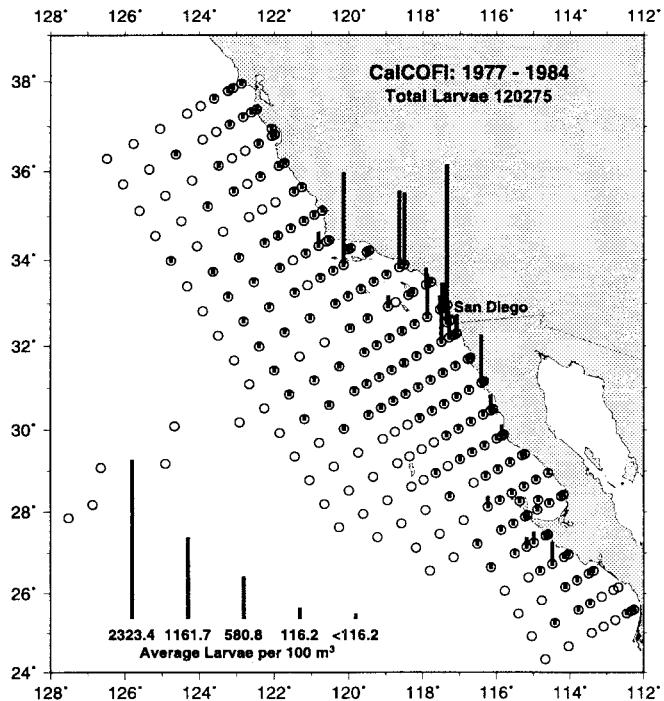
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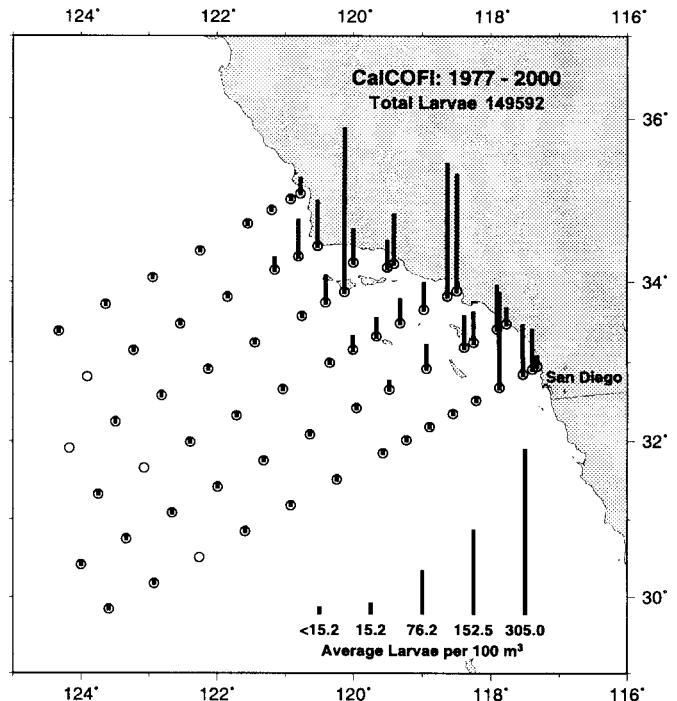
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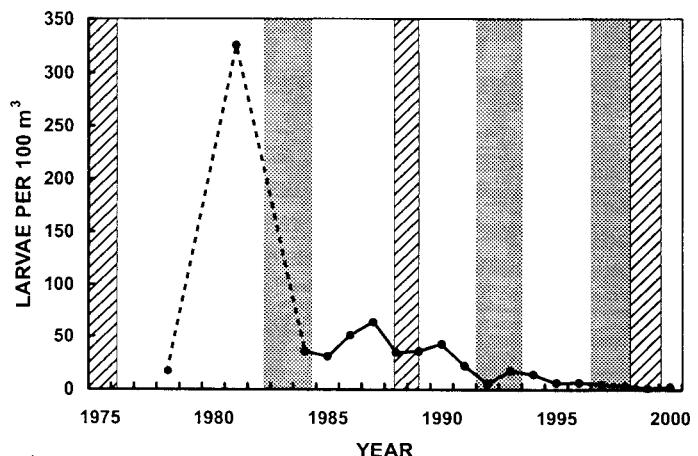
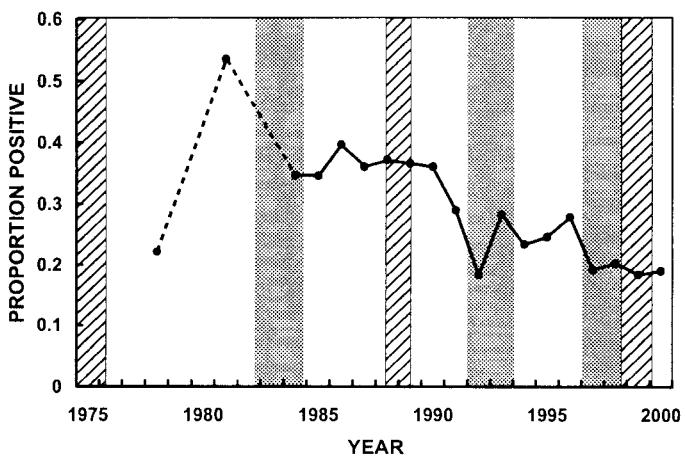
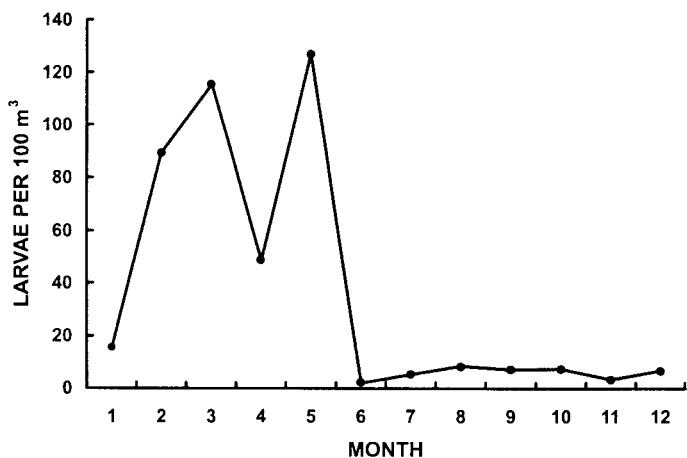
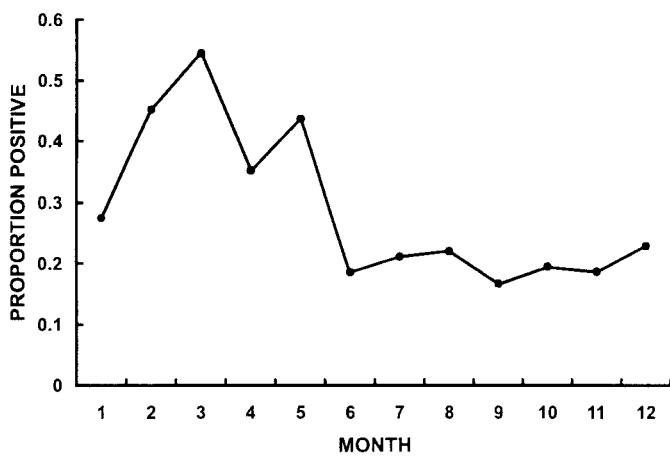
Engraulis mordax larvae



Northern anchovy larvae



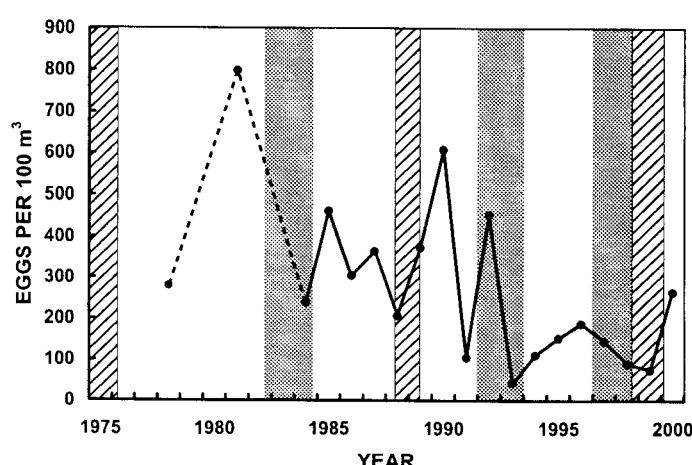
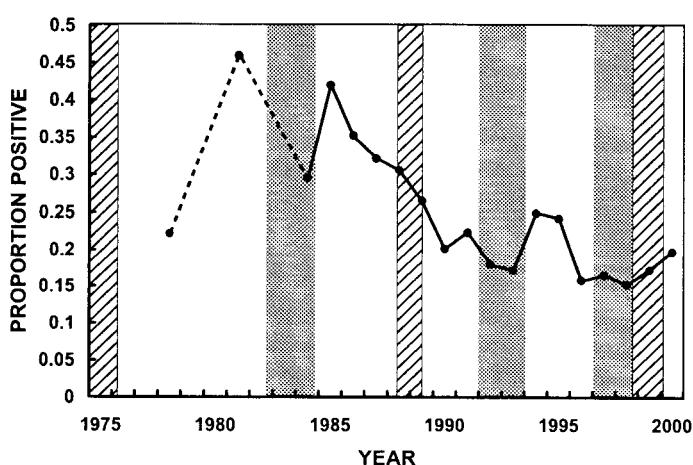
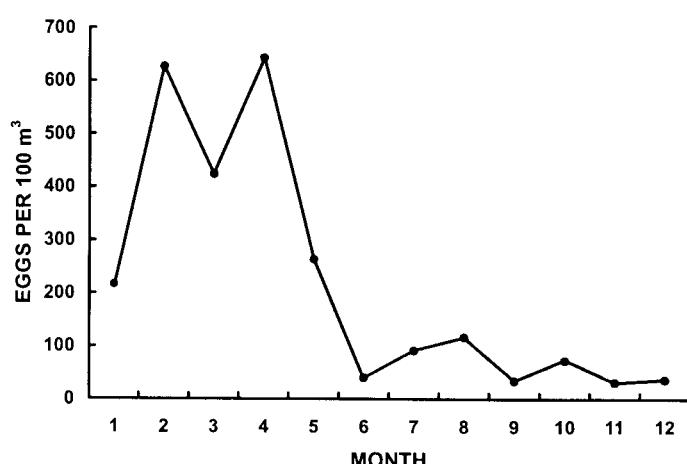
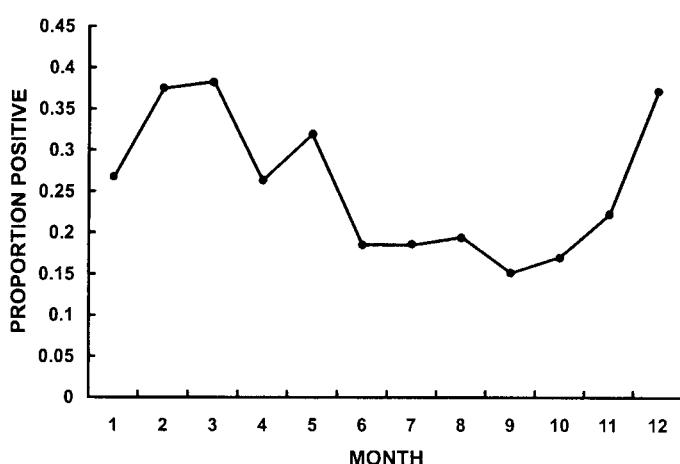
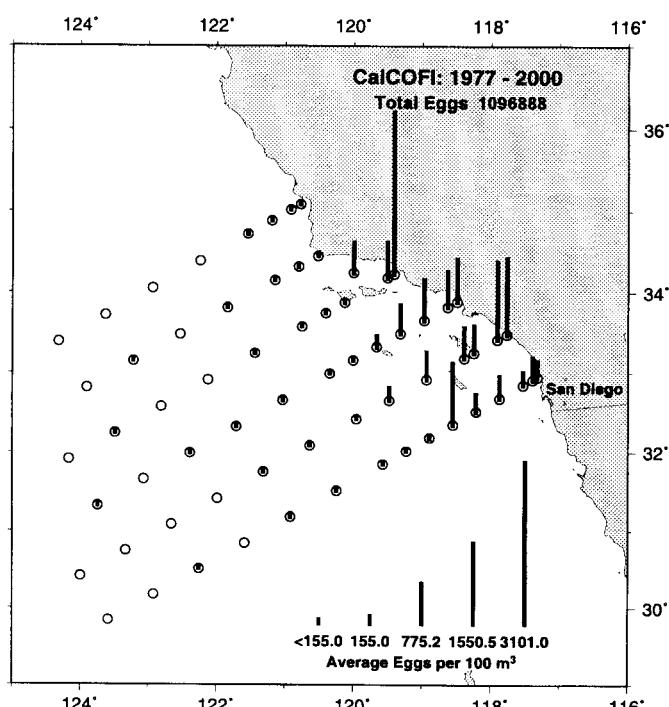
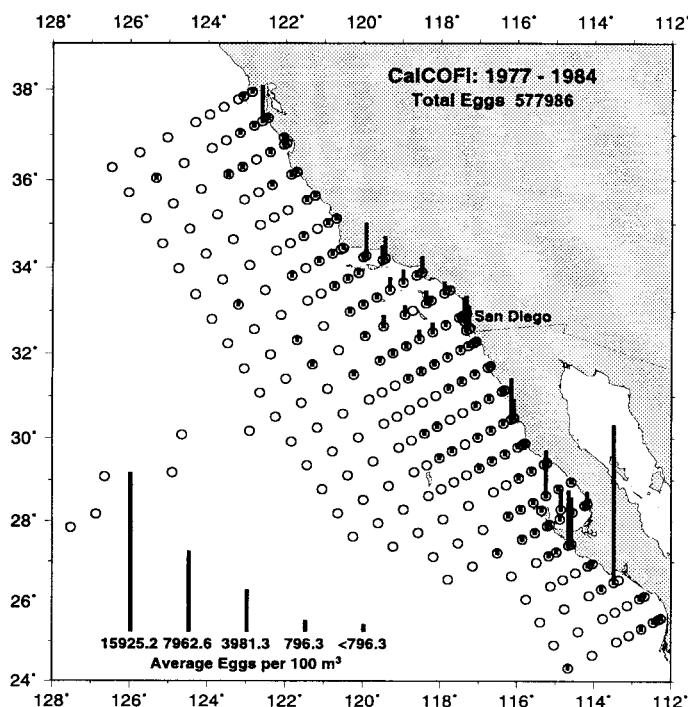
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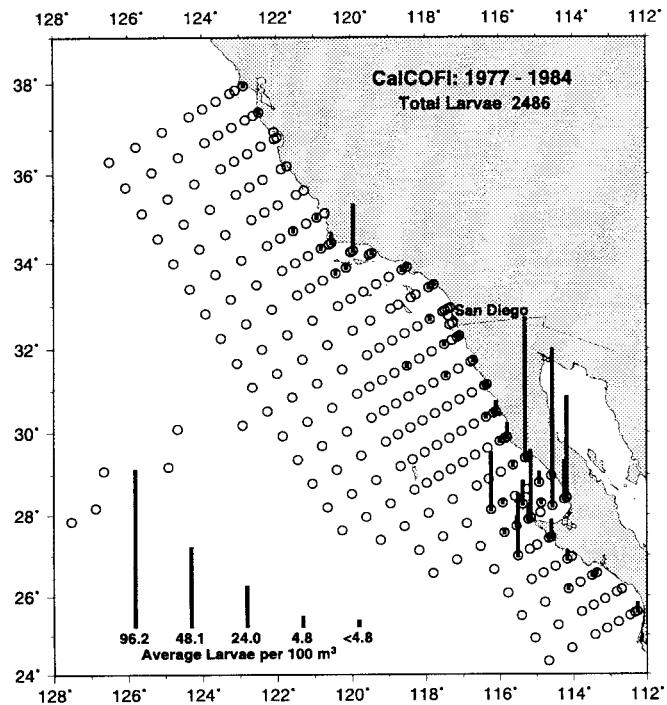
ENGRAULIDAE

Northern anchovy eggs

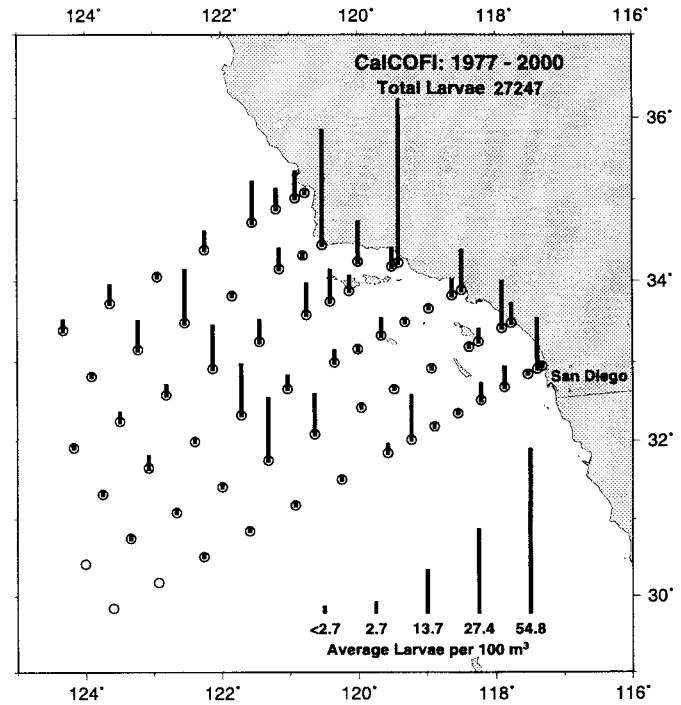
Engraulis mordax eggs



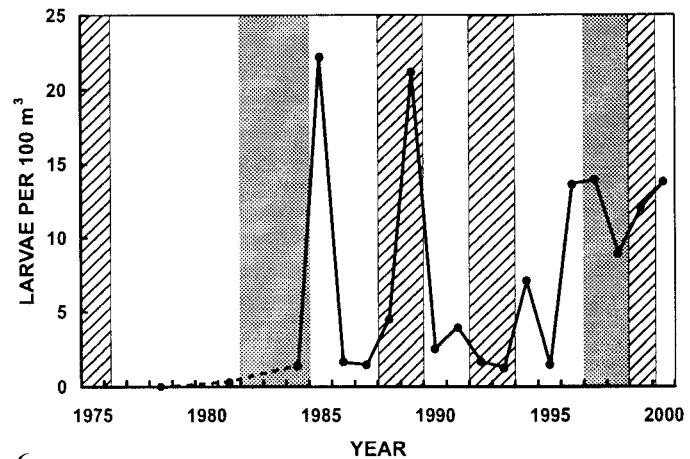
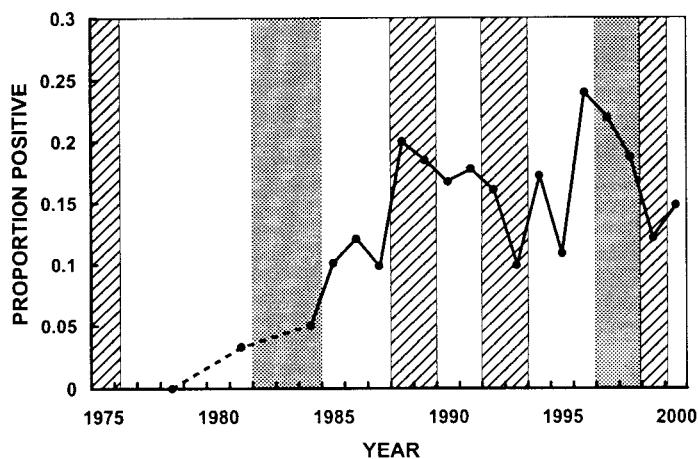
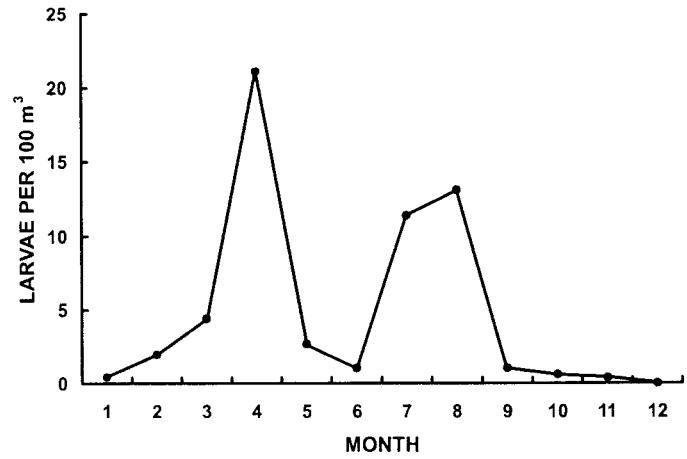
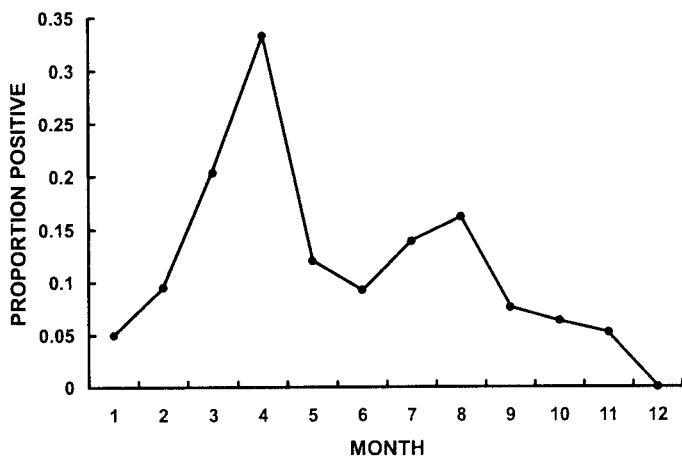
Sardinops sagax larvae



Pacific sardine larvae



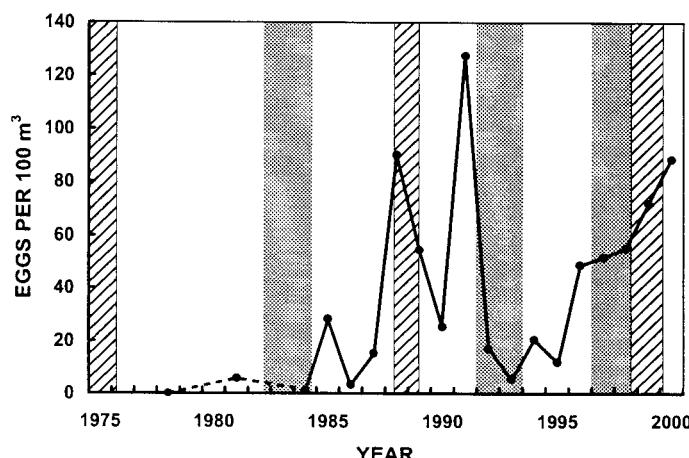
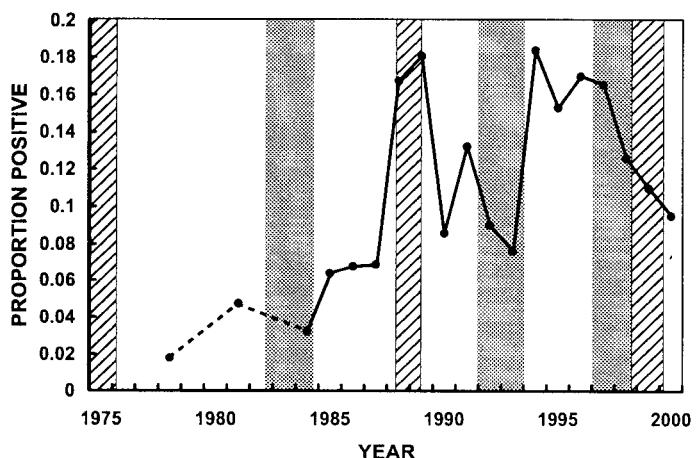
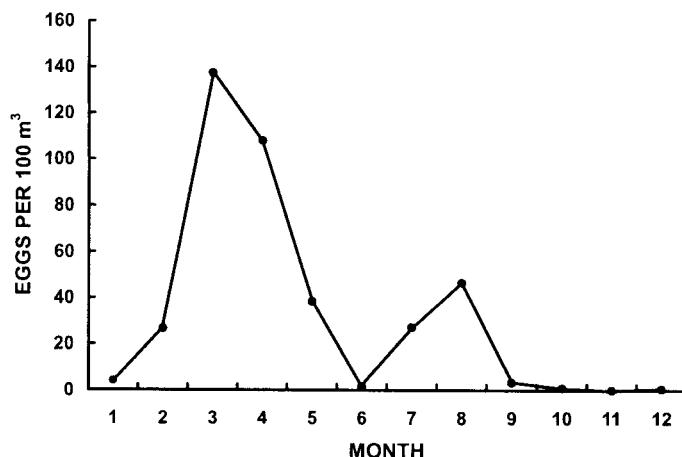
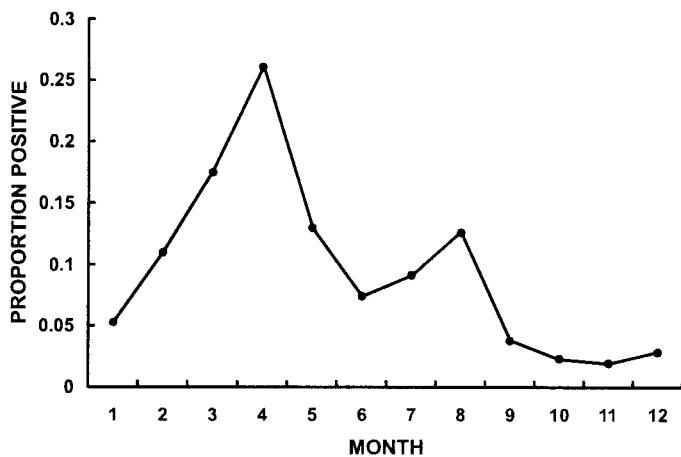
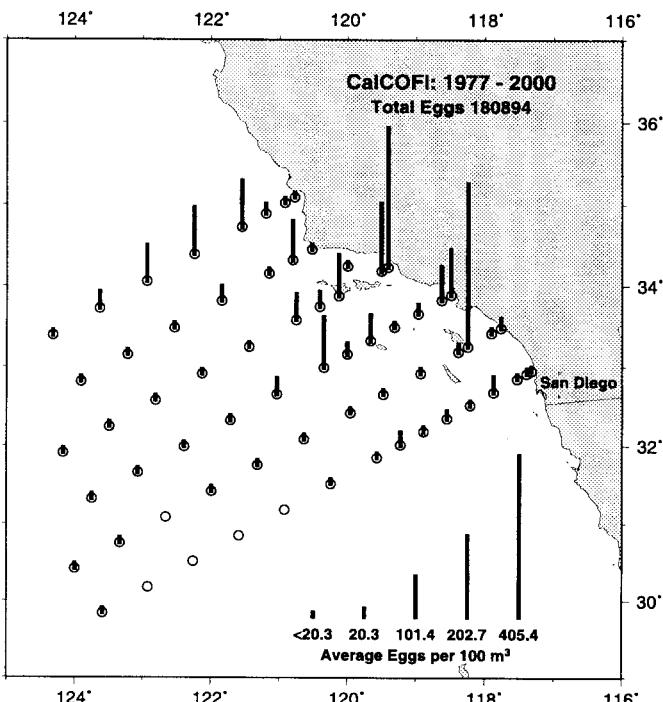
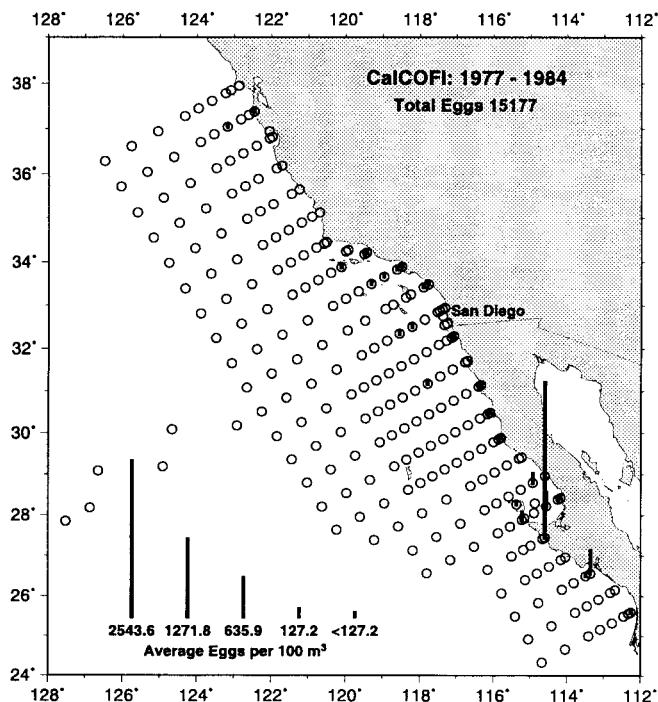
CLUPEIDAE



CLUPEIDAE

Pacific sardine eggs

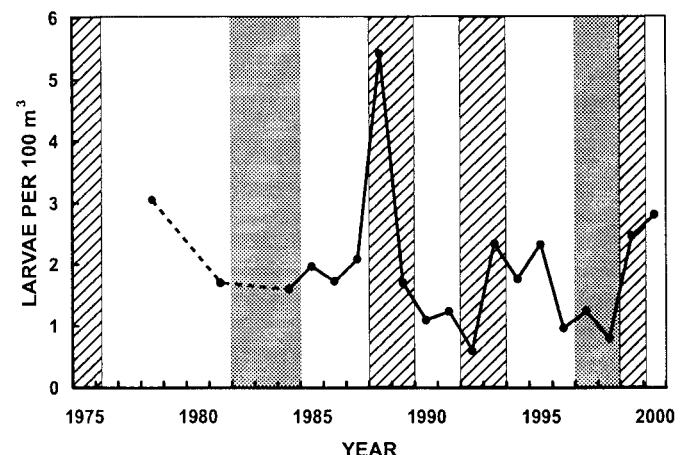
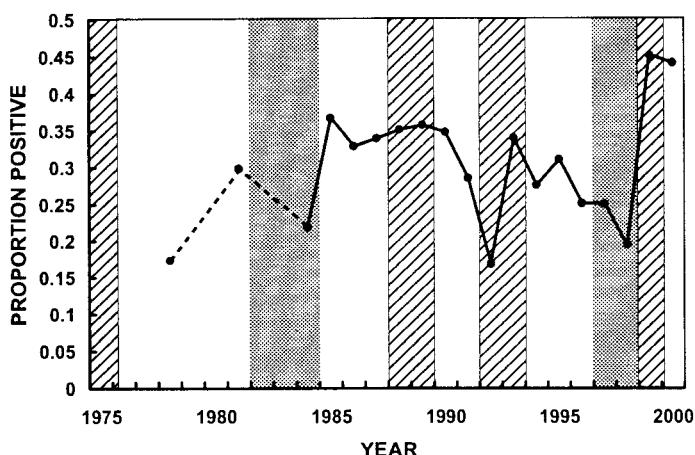
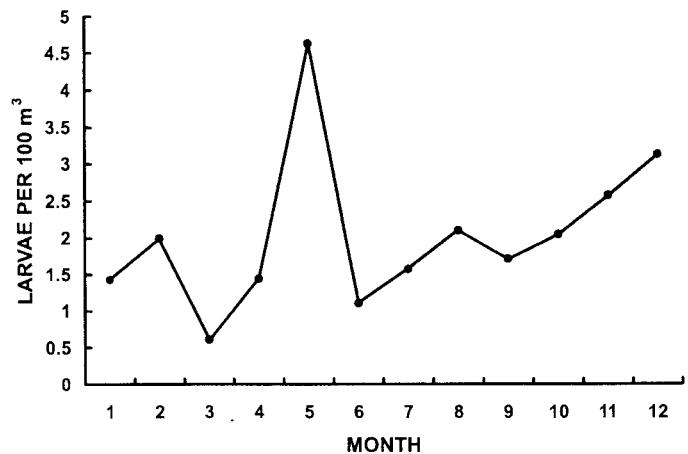
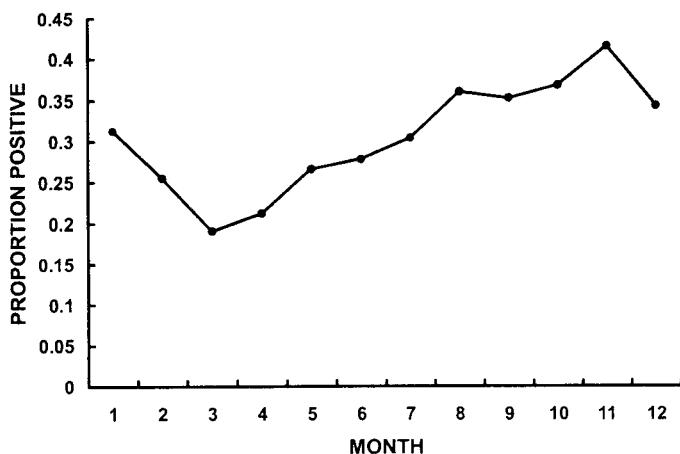
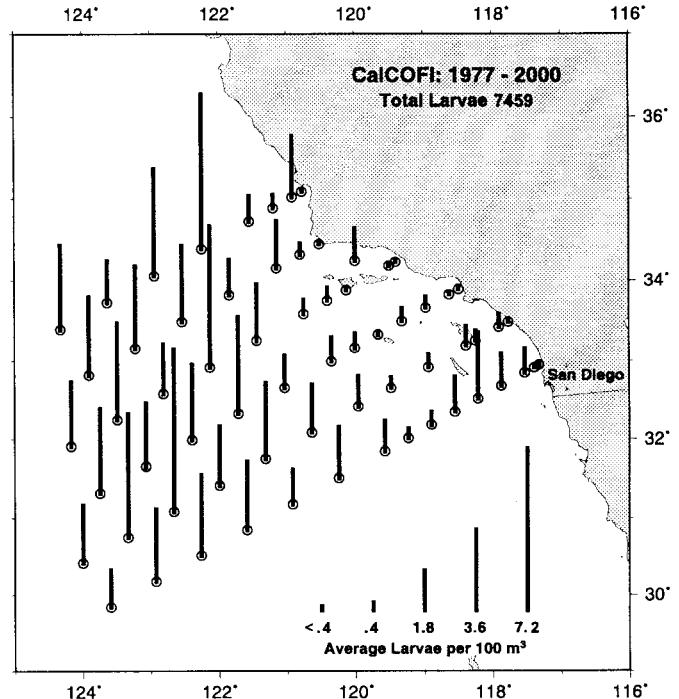
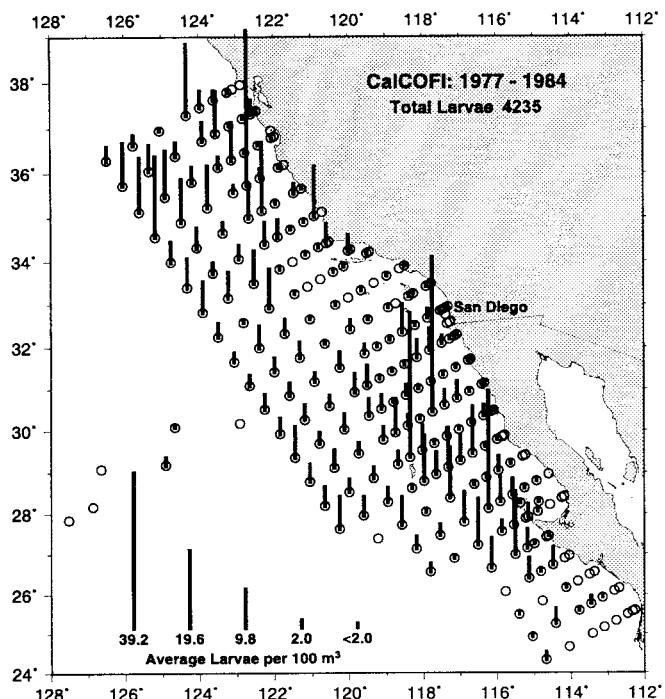
Sardinops sagax eggs



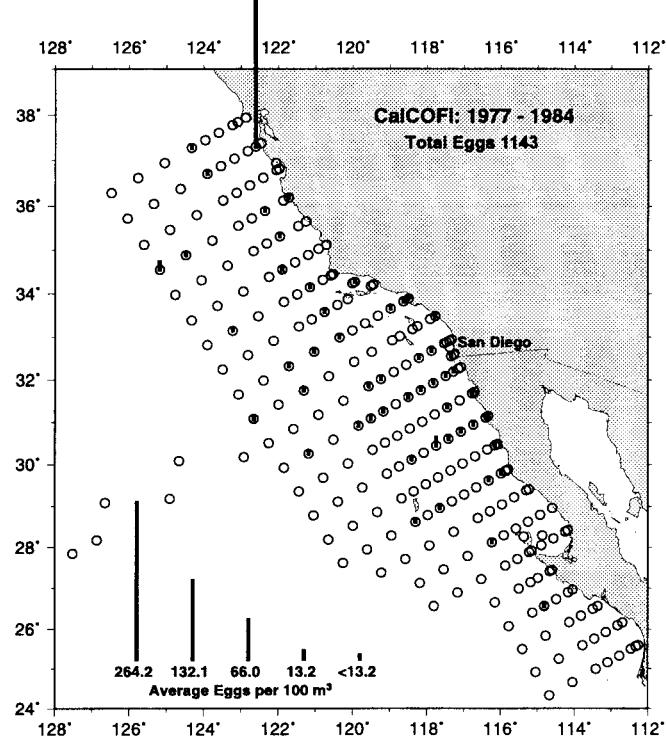
Cololabis saira larvae

Pacific saury larvae

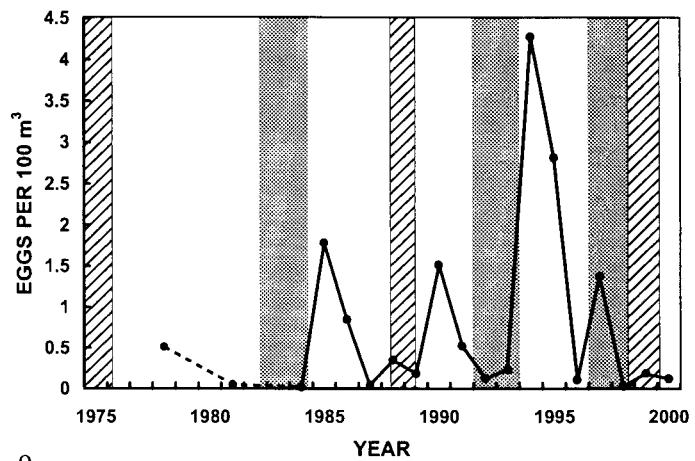
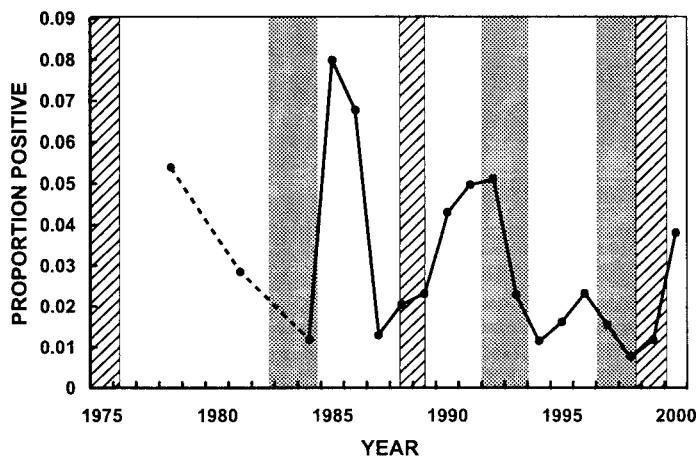
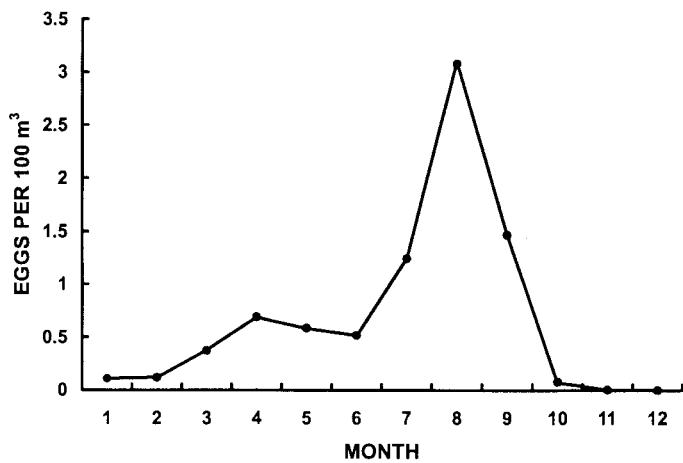
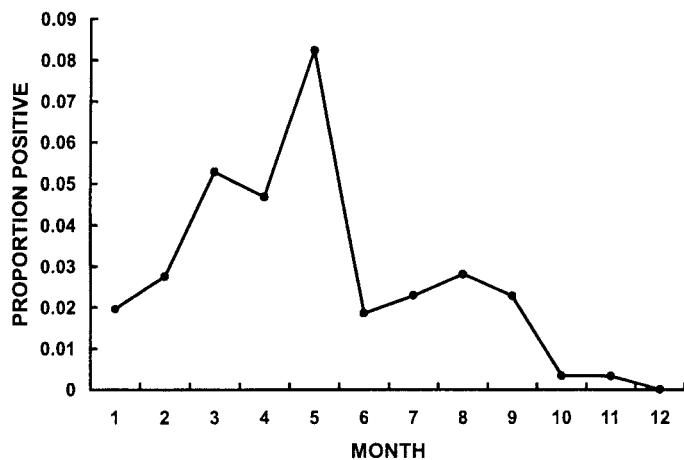
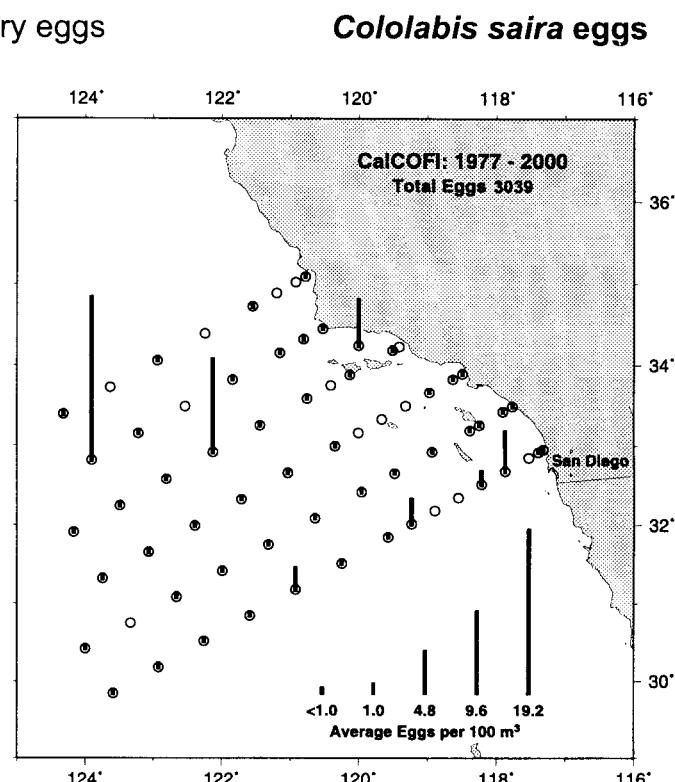
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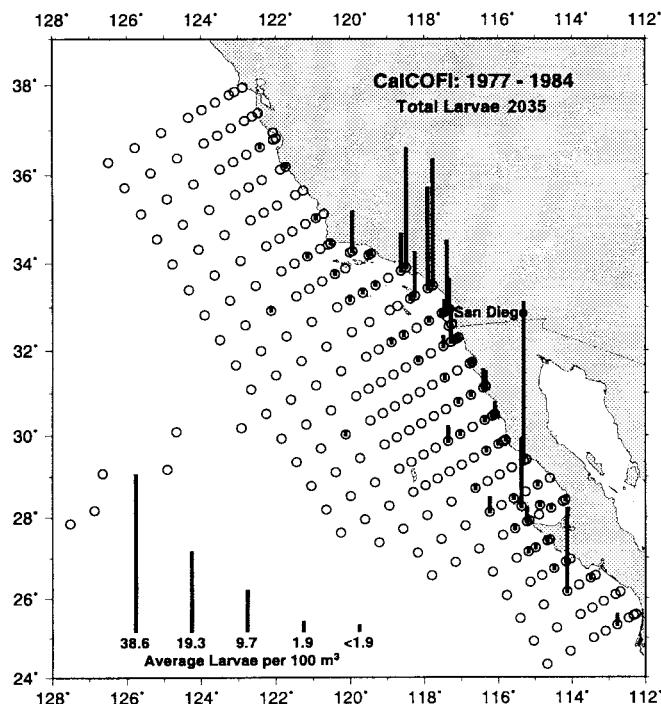
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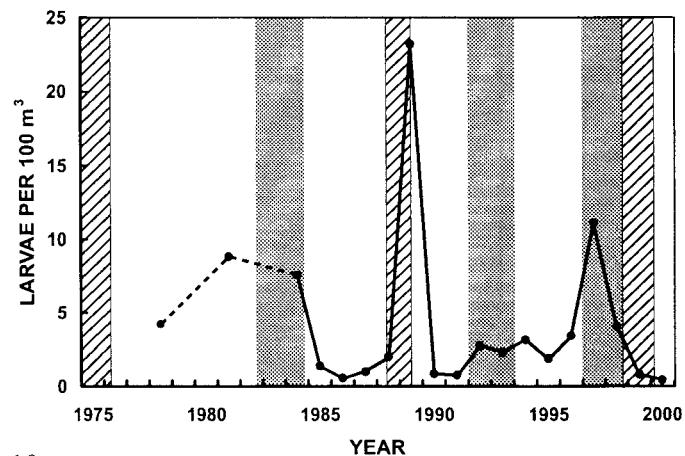
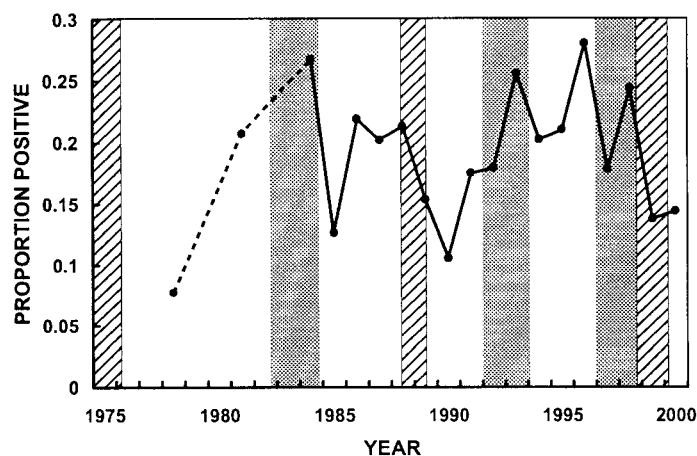
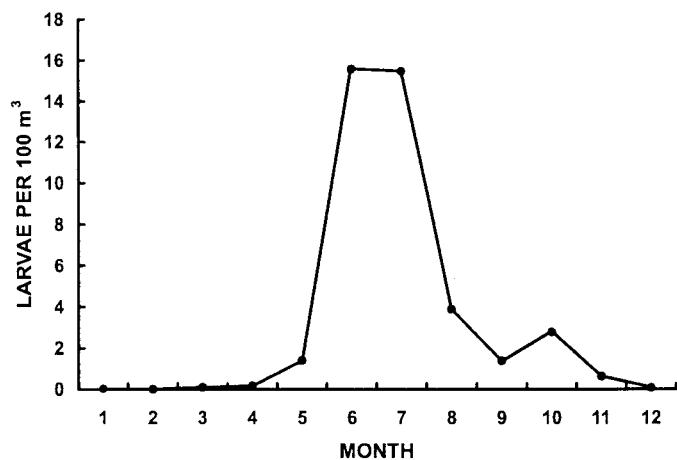
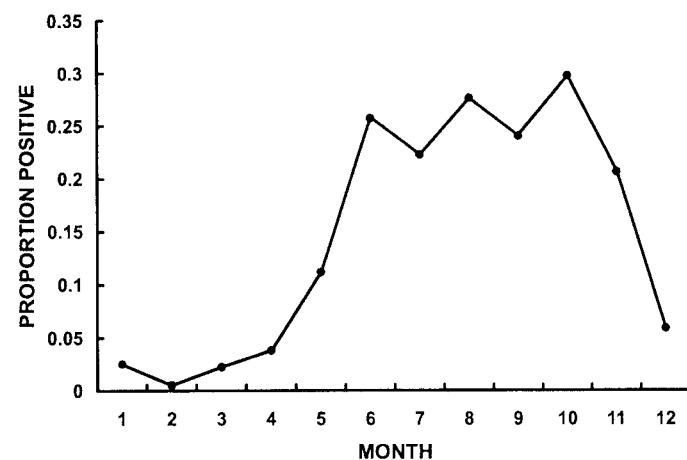
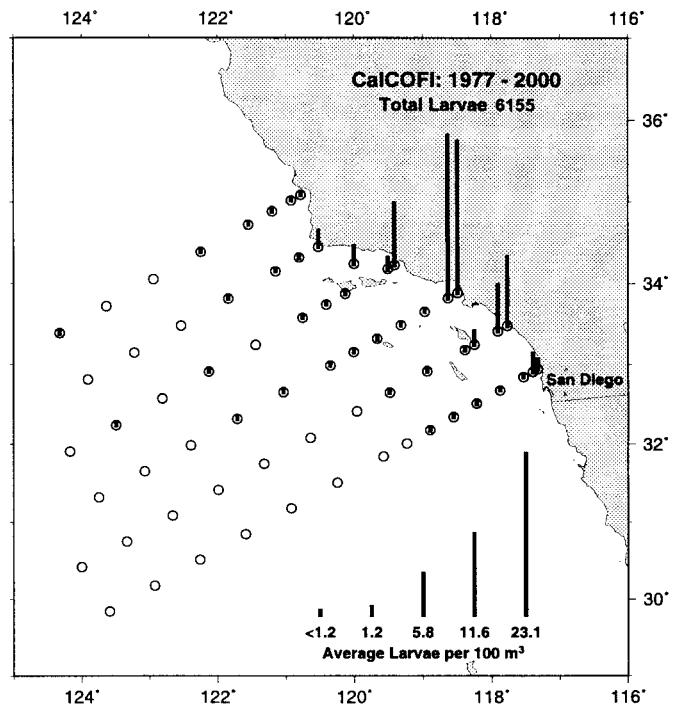
Pacific saury eggs



Hypsoblennius jenkinsi



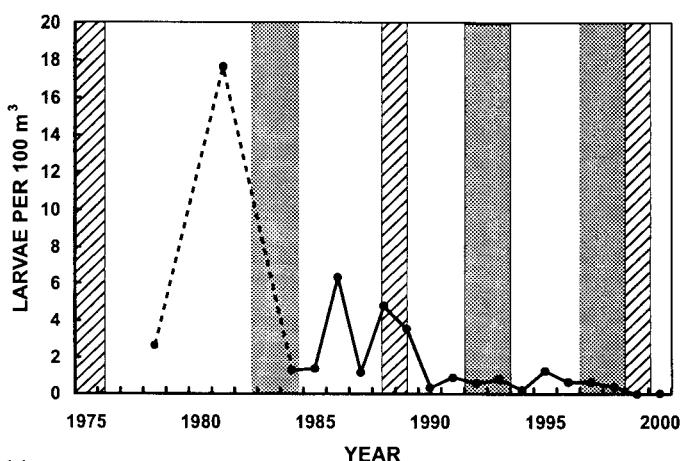
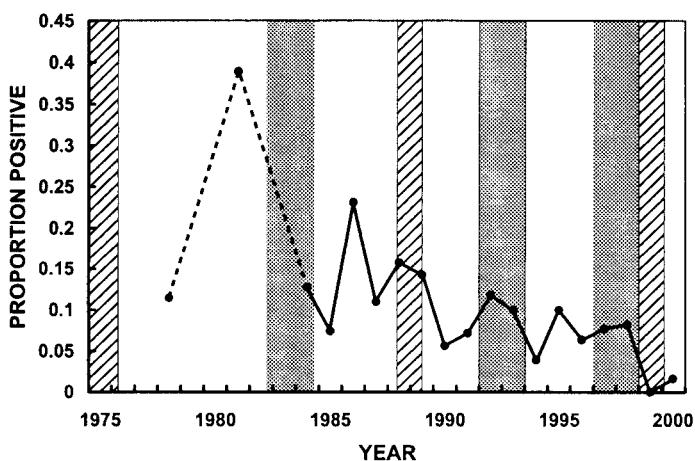
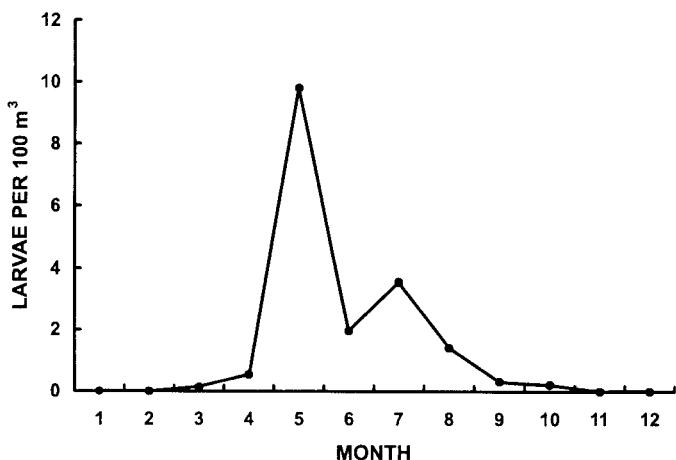
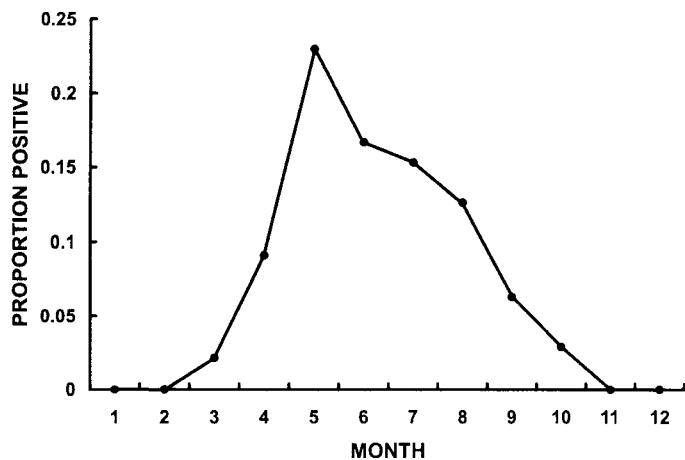
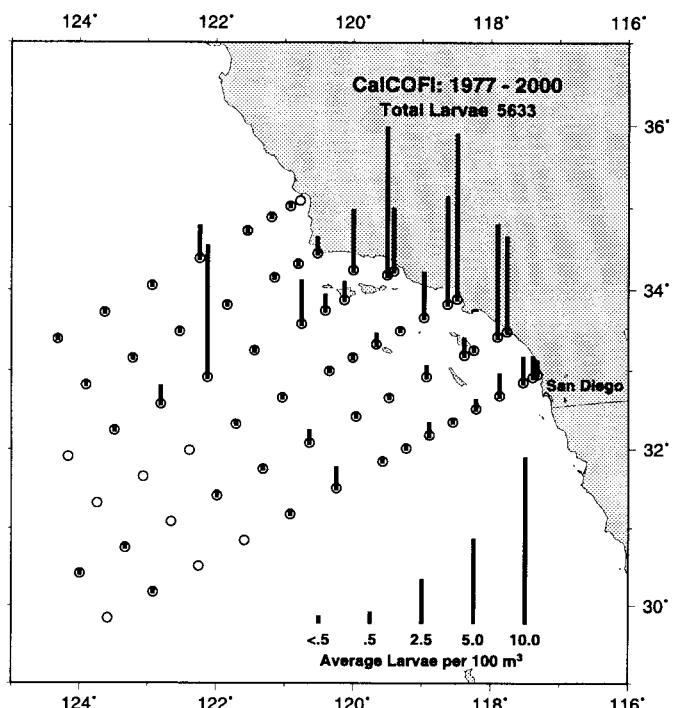
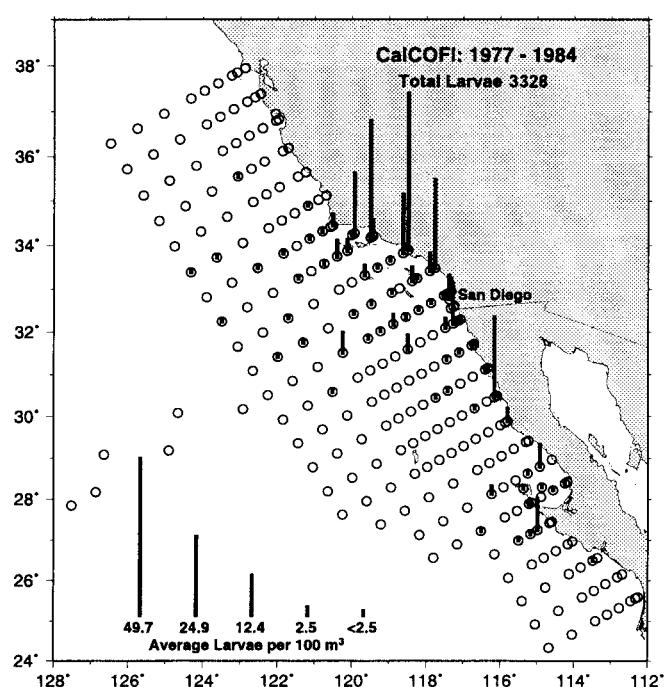
Mussel blenny



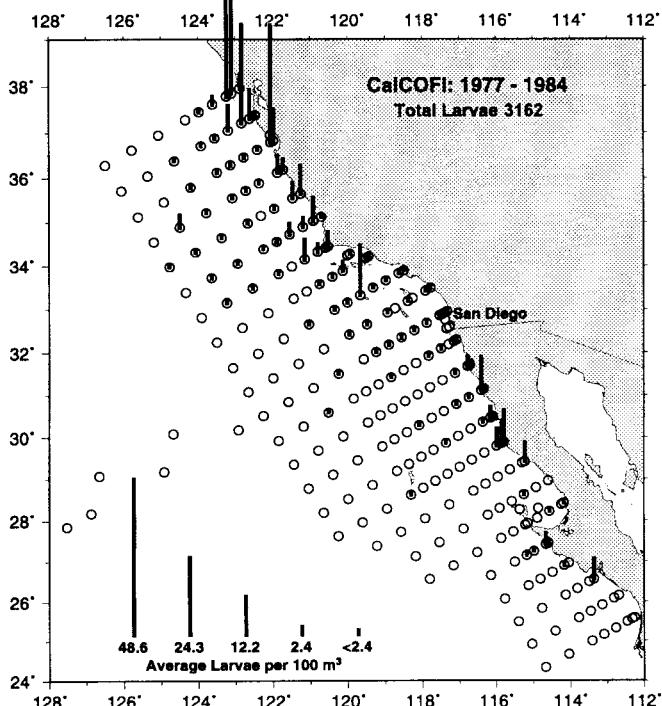
SCOMBRIDAE

Chub mackerel

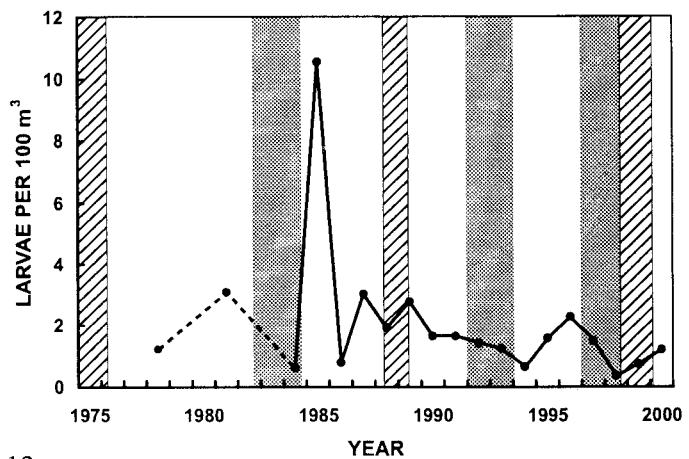
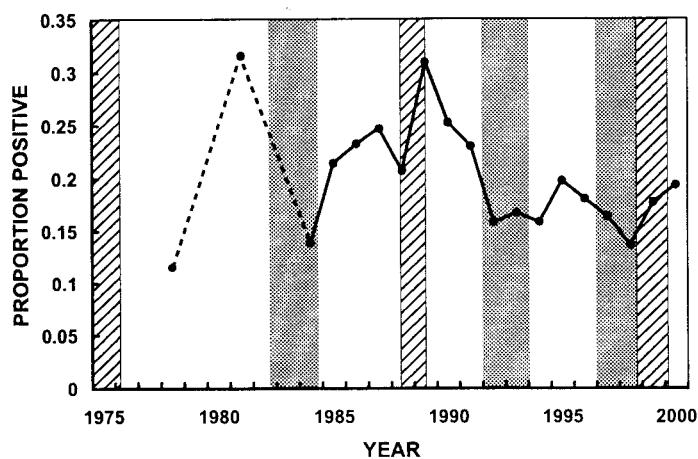
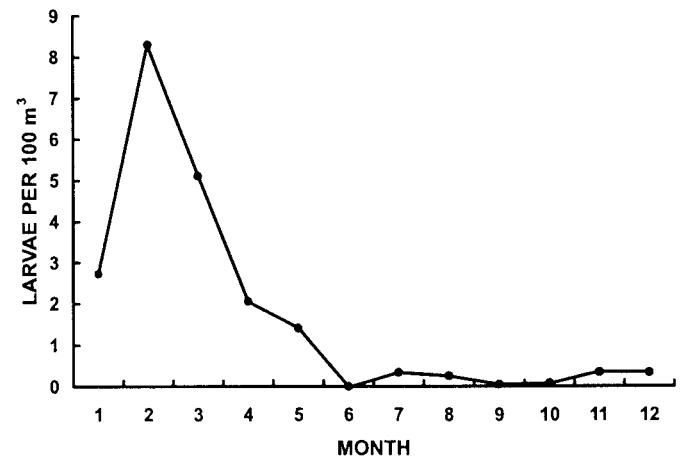
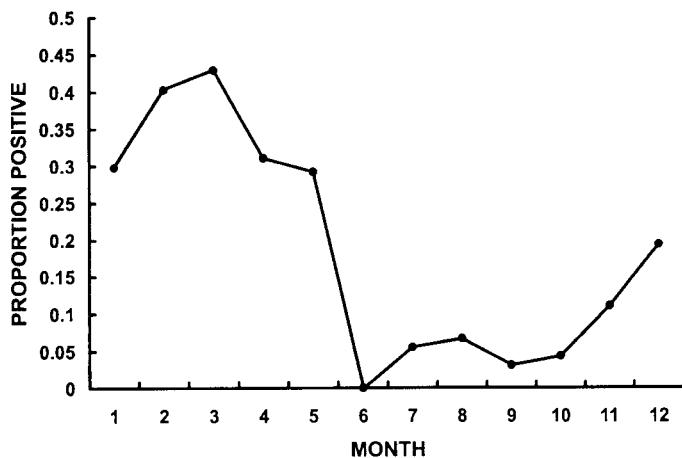
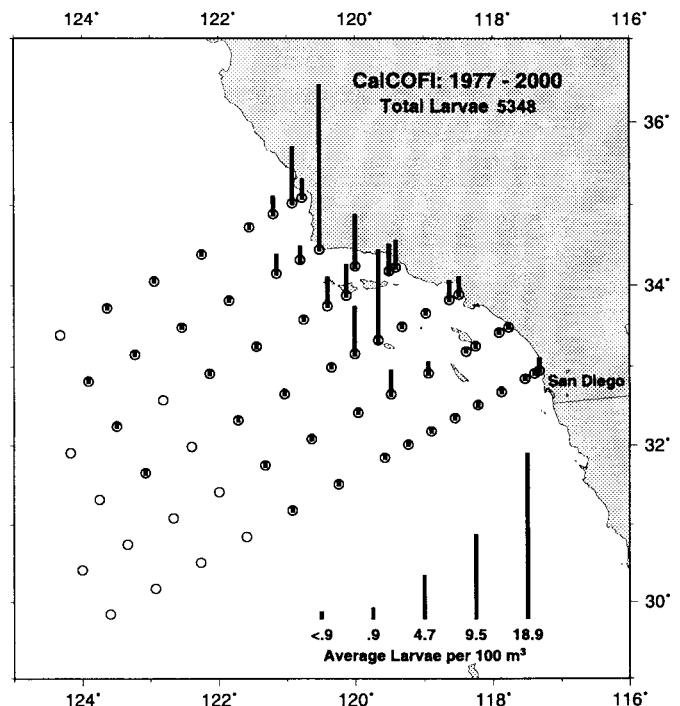
Scomber japonicus



Sebastes spp.



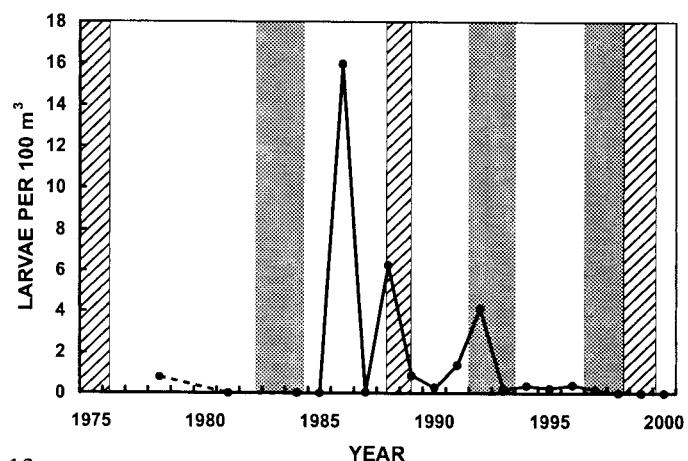
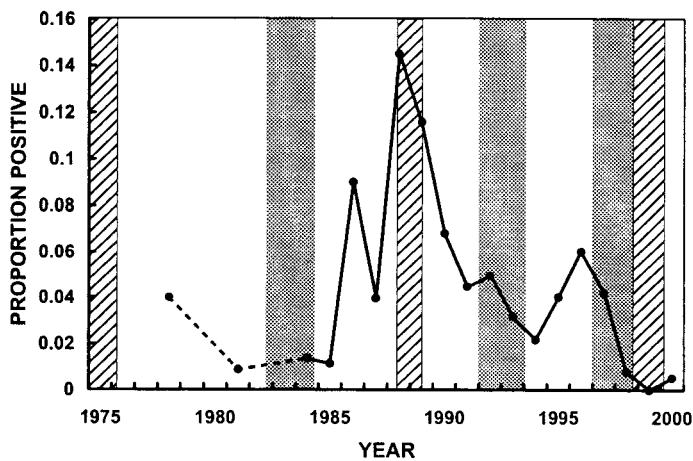
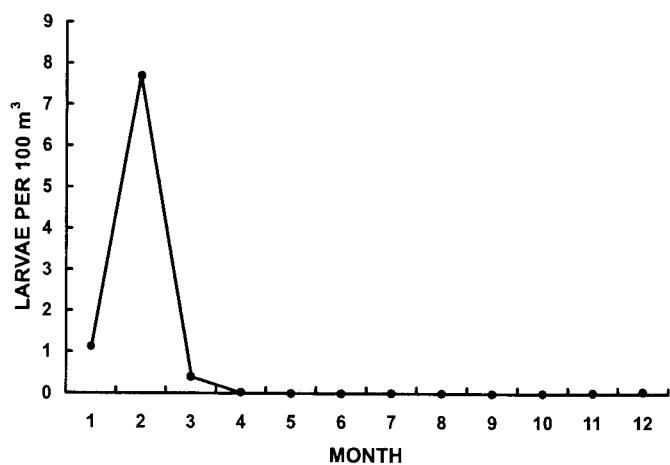
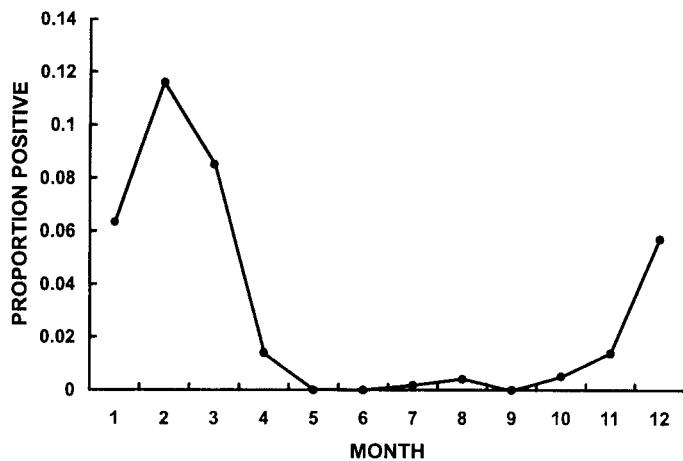
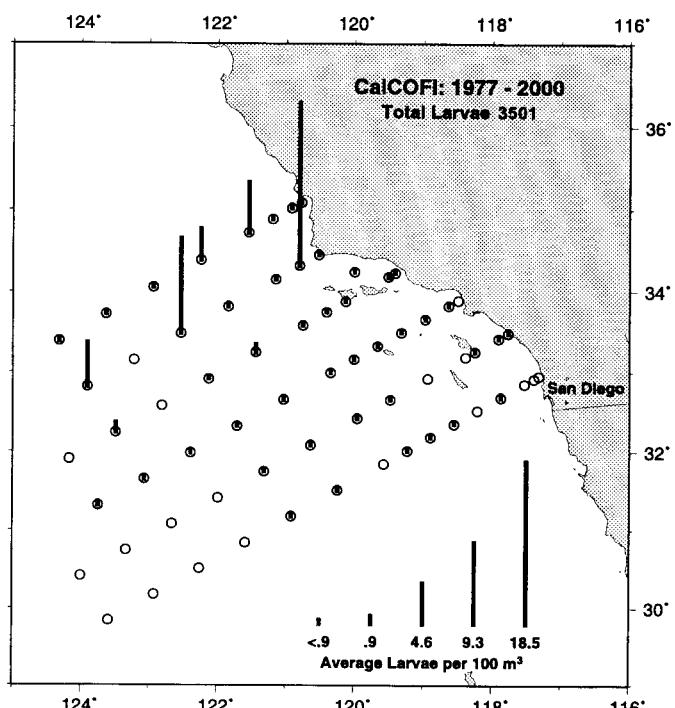
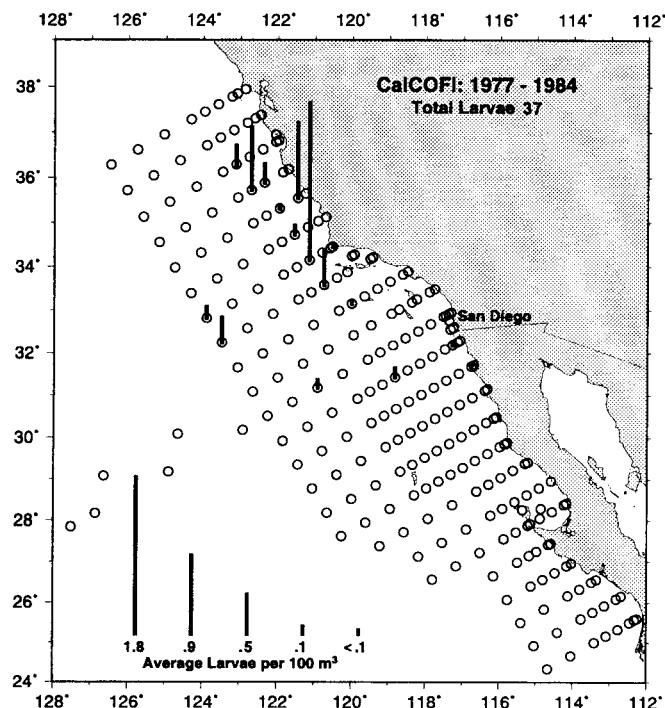
Rockfishes



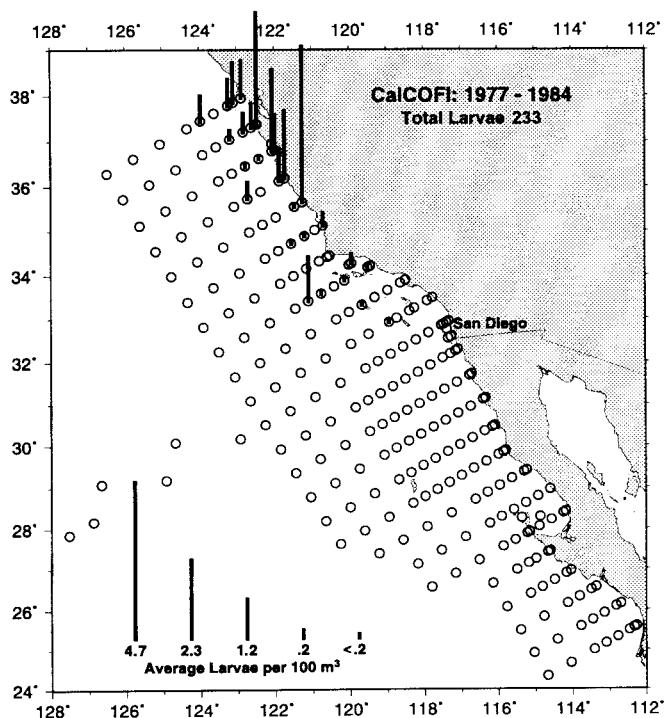
MERLUCCIIDAE

Pacific hake or whiting

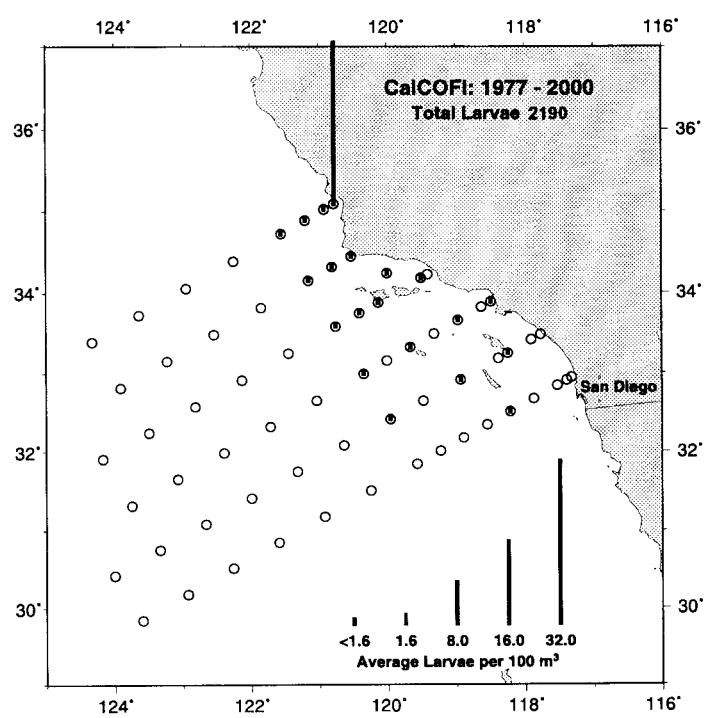
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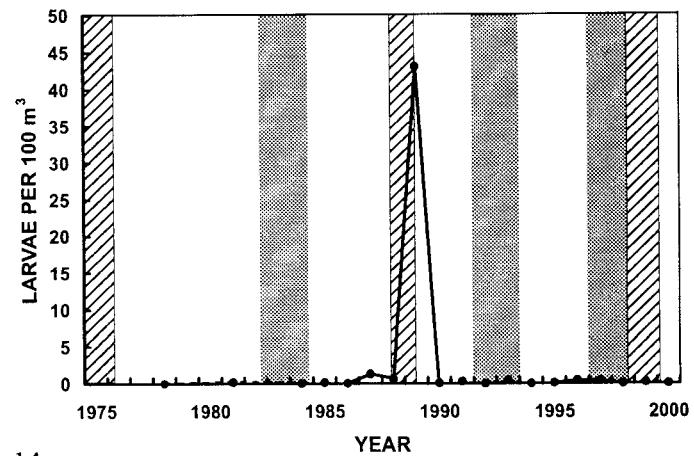
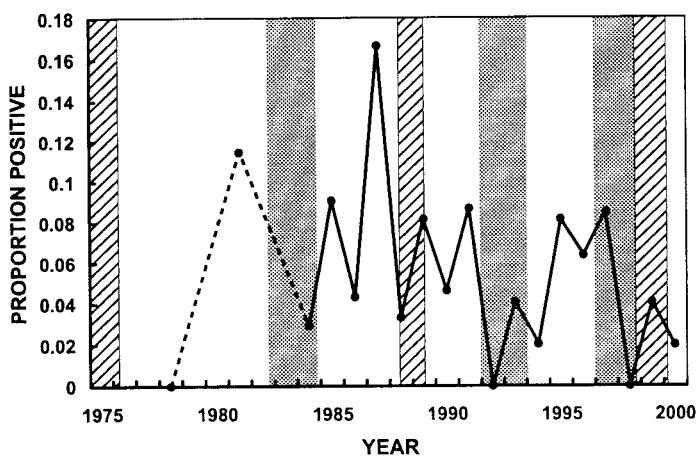
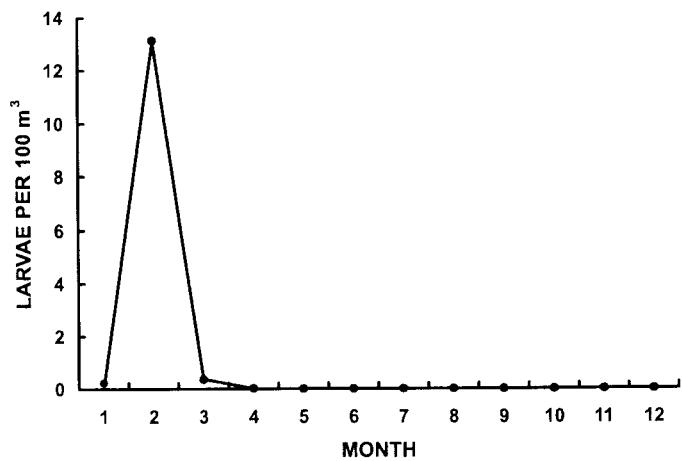
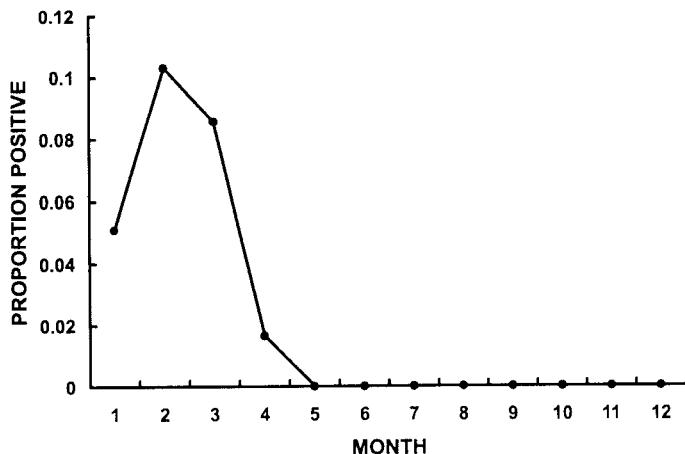
Hexagrammos decagrammus



Kelp greenling



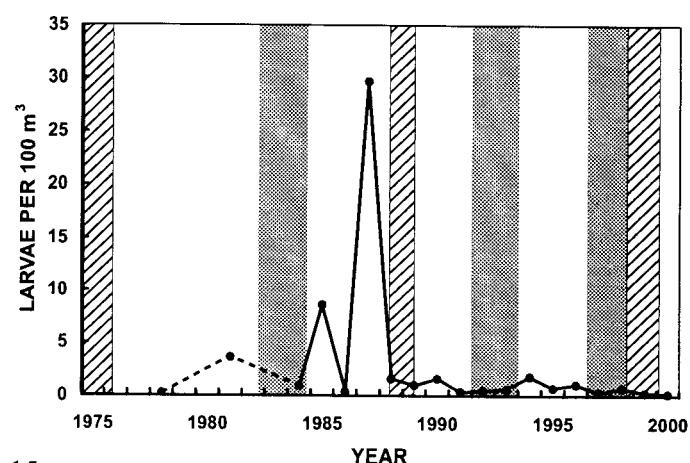
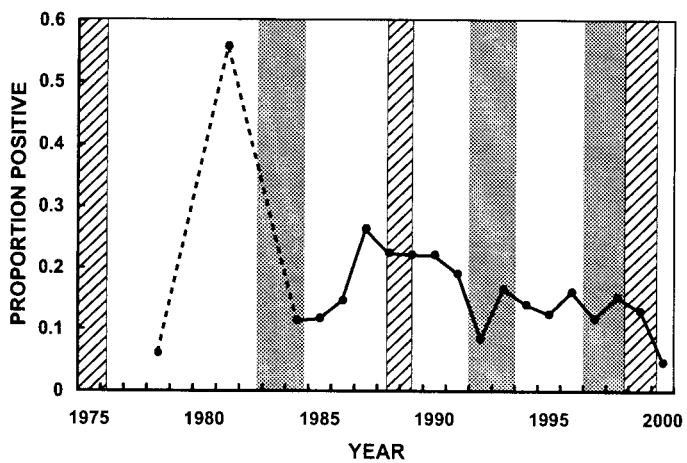
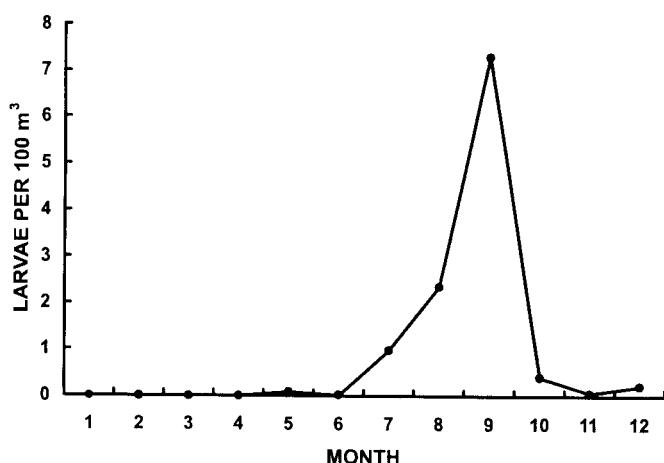
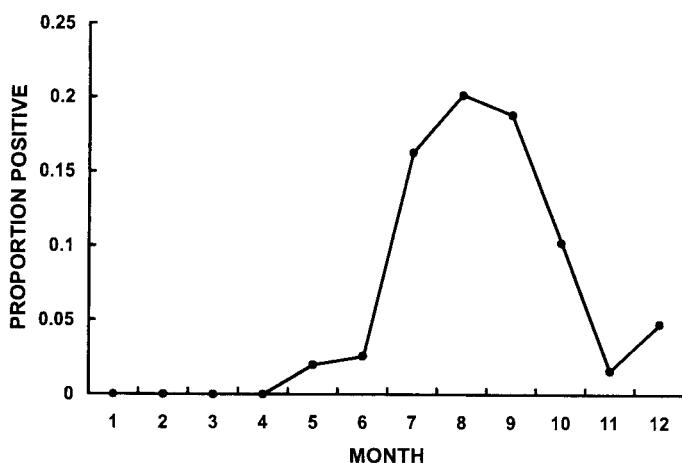
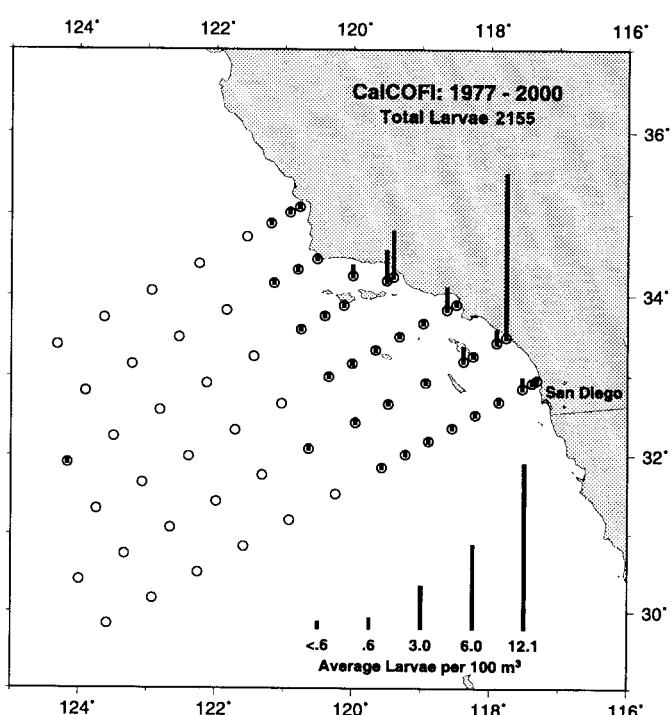
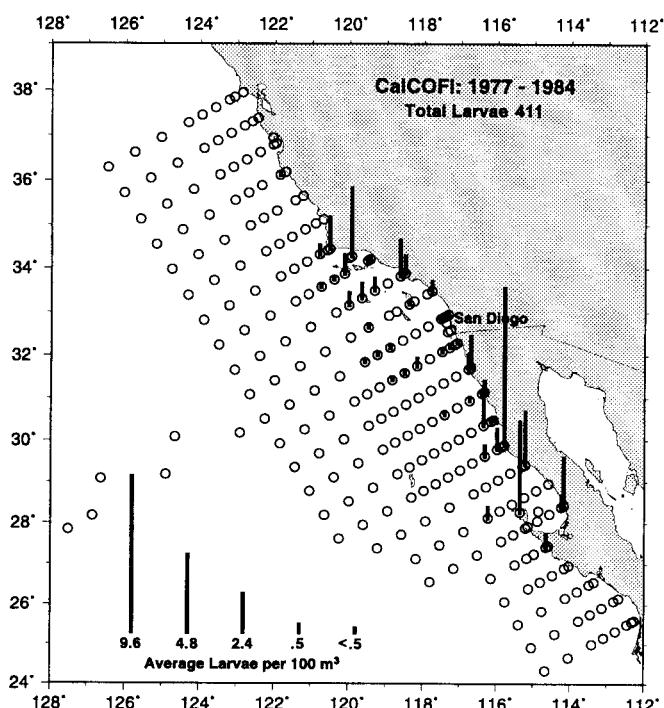
HEXAGRAMMIDAE



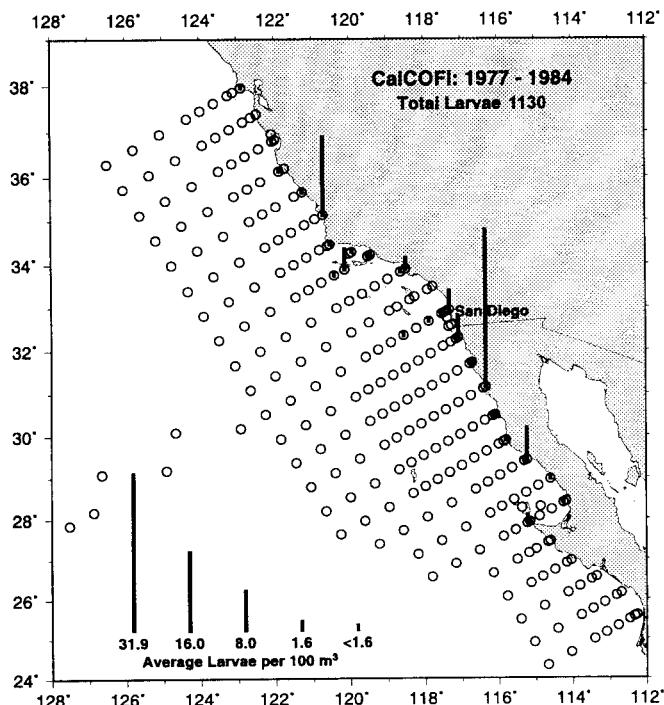
POMACENTRIDAE

Blacksmith

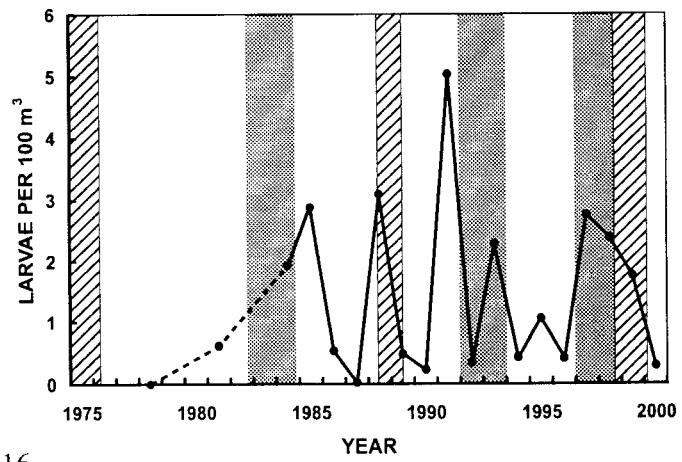
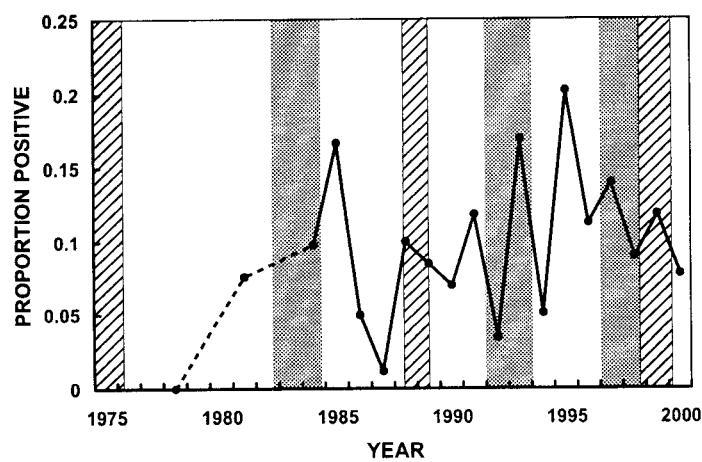
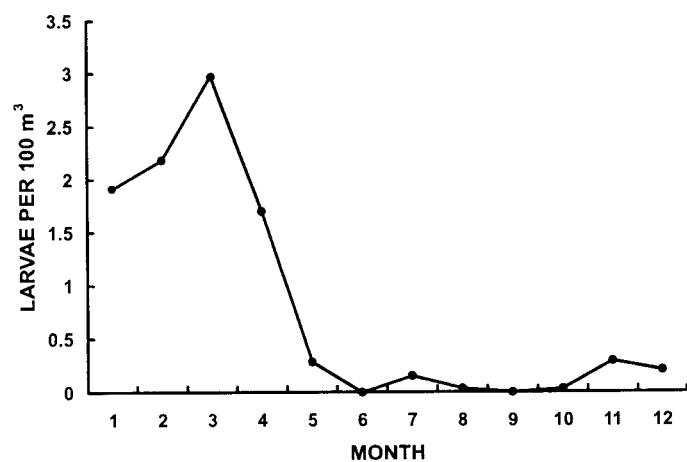
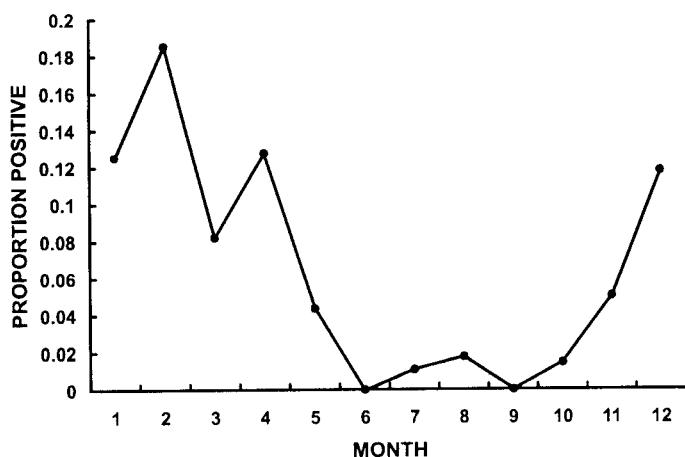
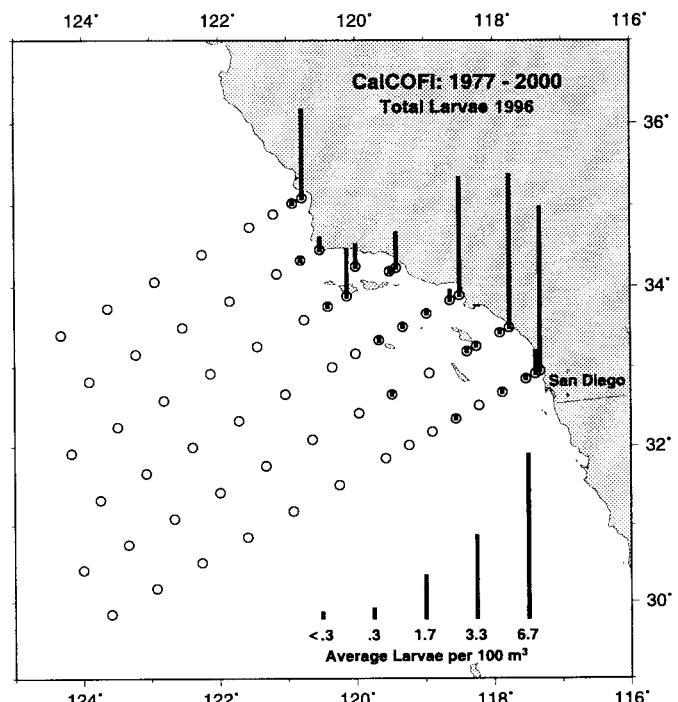
Chromis punctipinnis



Atherinopsis californiensis



Jacksmelt

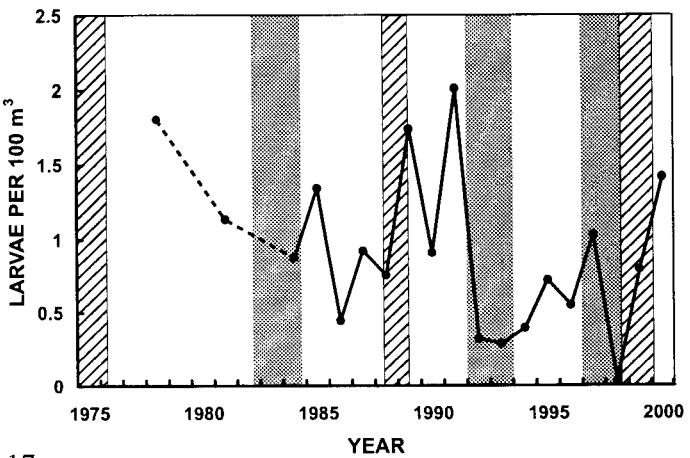
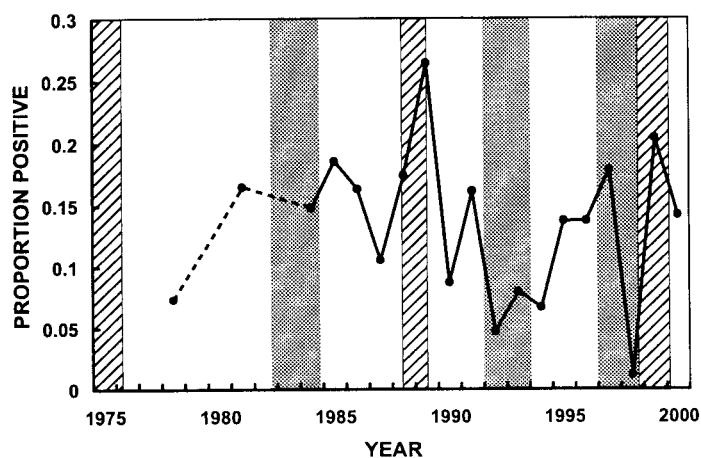
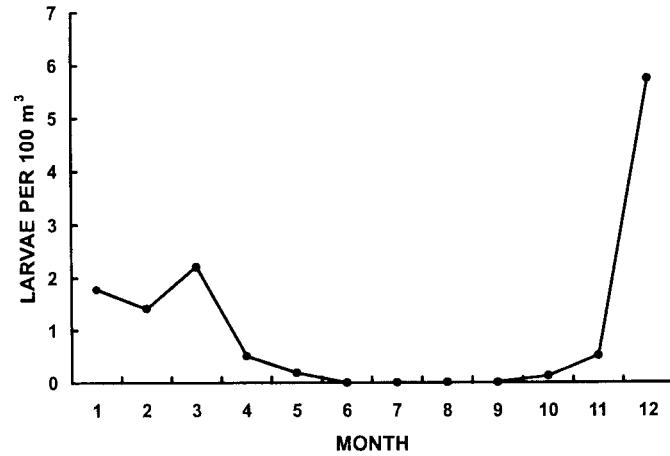
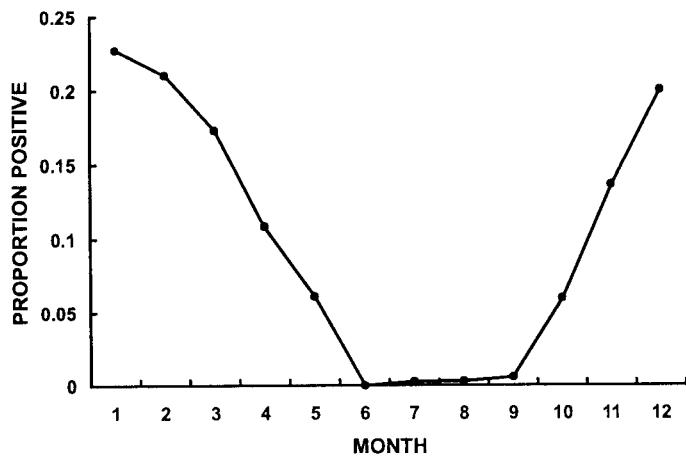
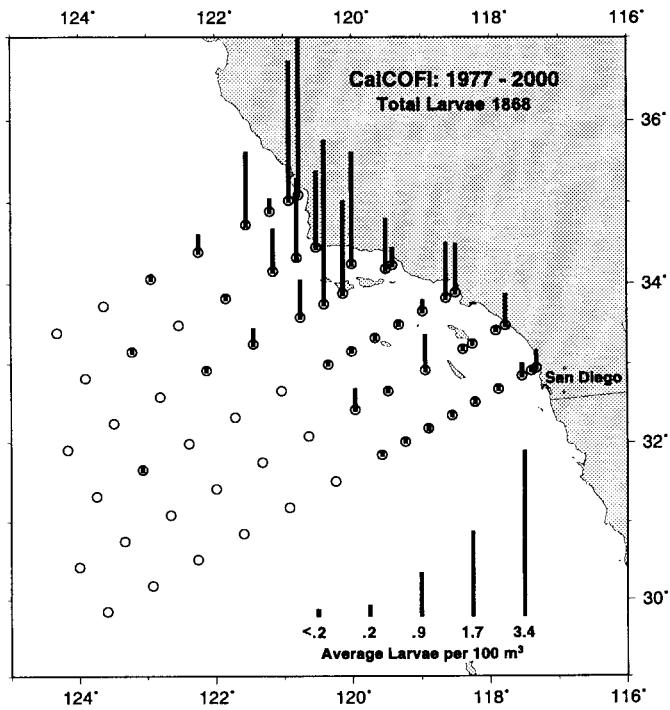
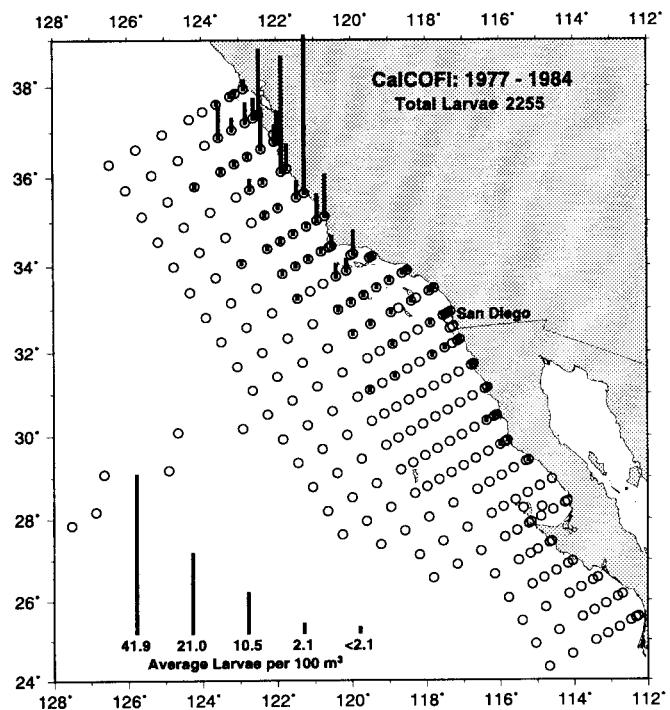


ATHERINIDAE

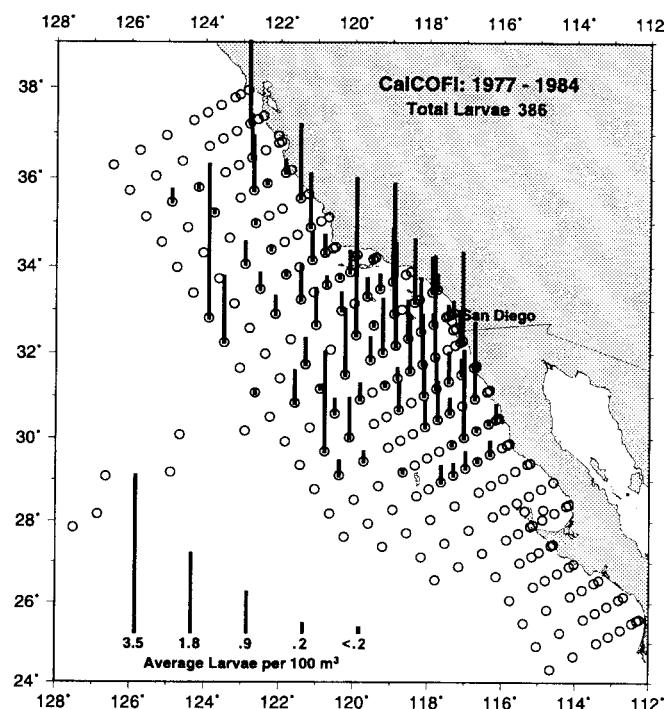
COTTIDAE

Cabezon

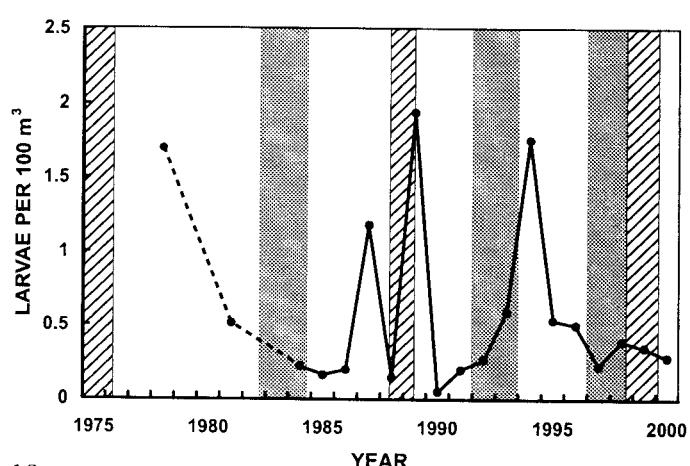
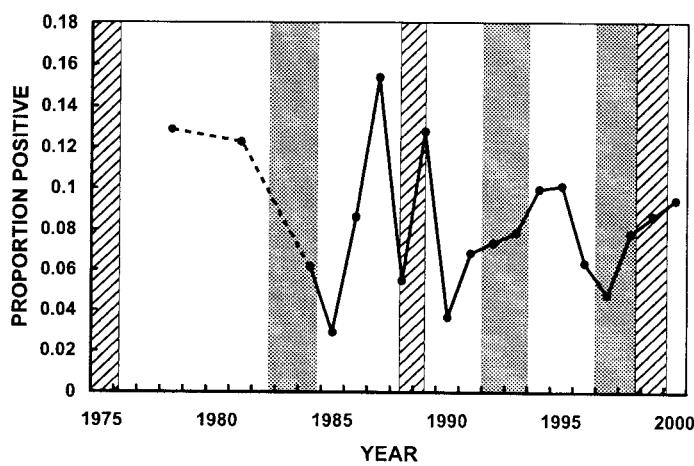
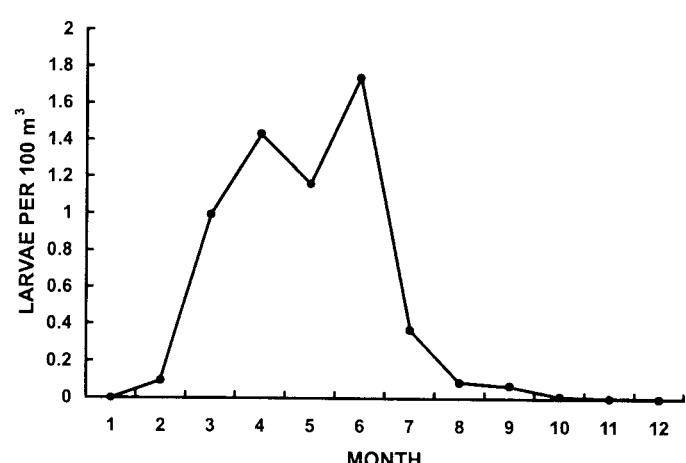
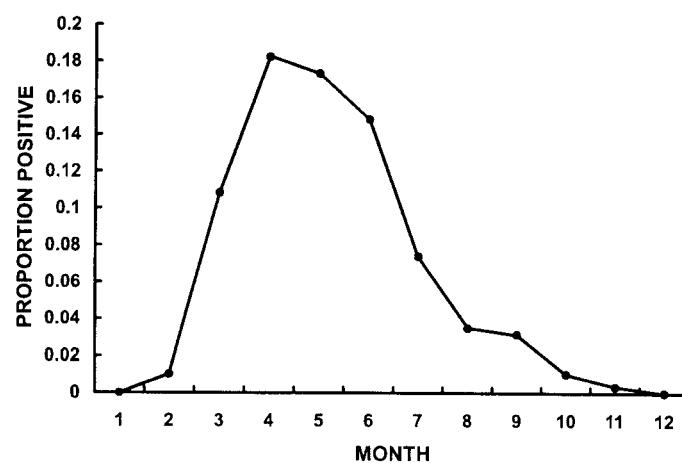
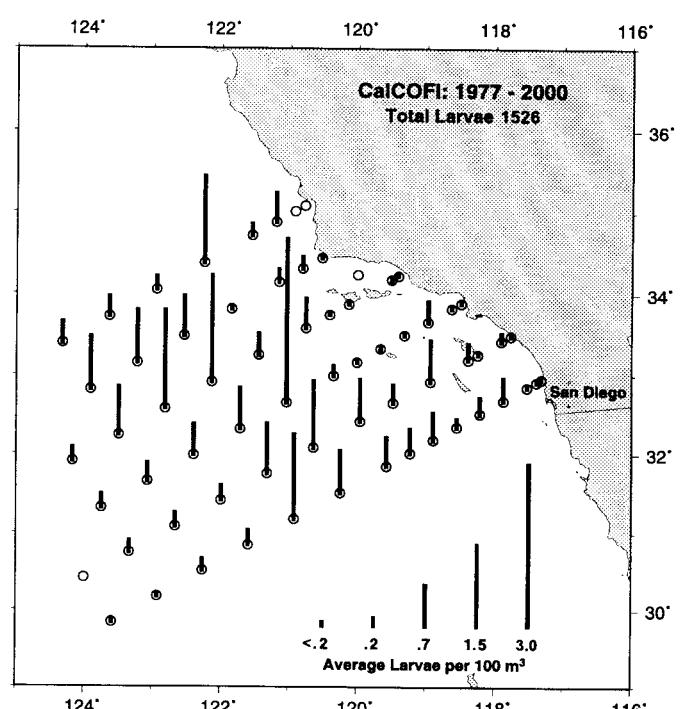
Scorpaenichthys marmoratus



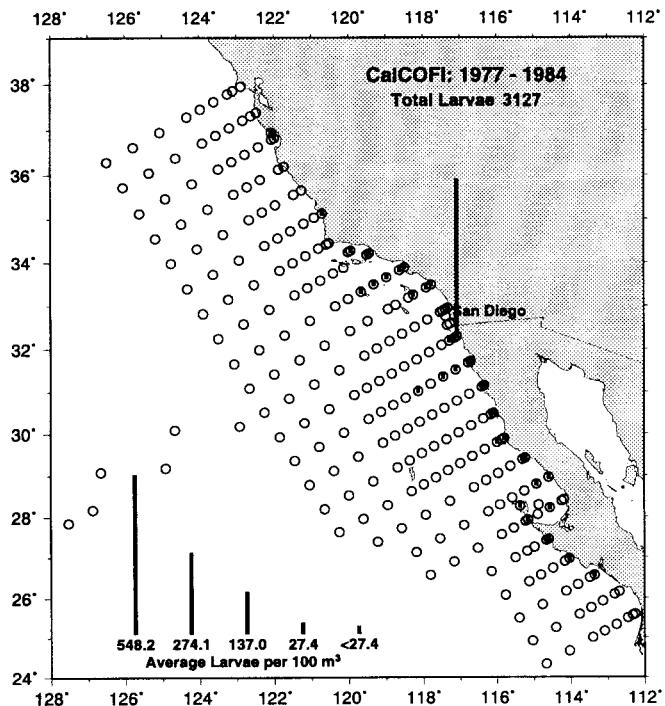
Trachurus symmetricus



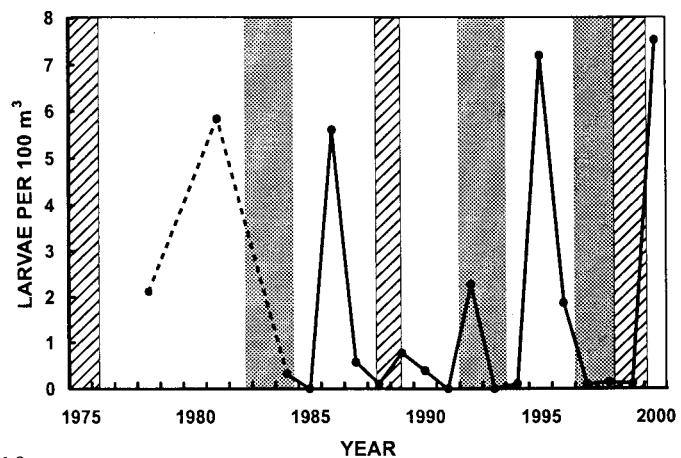
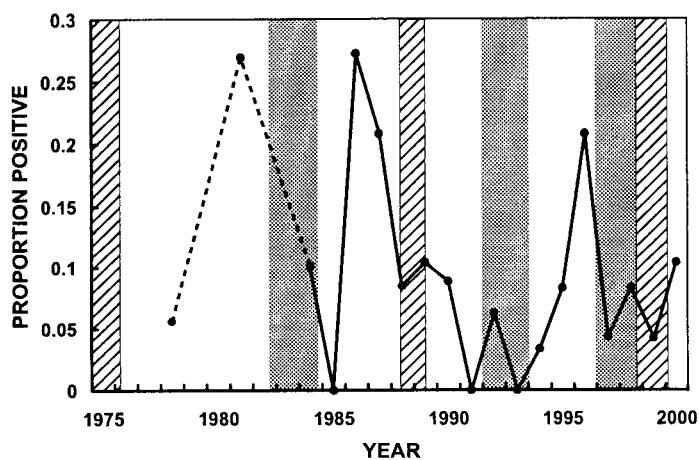
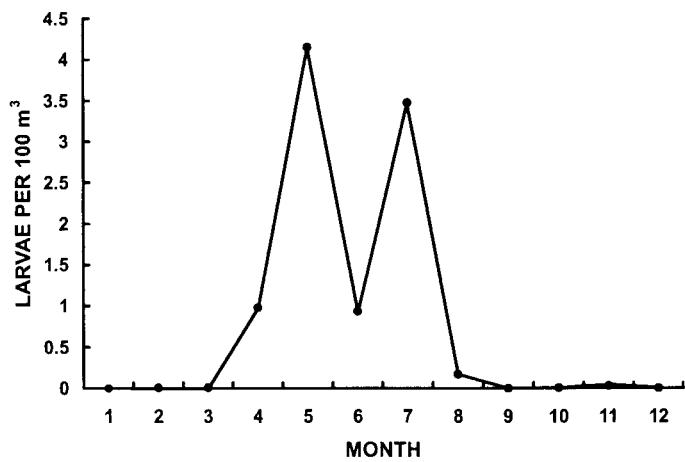
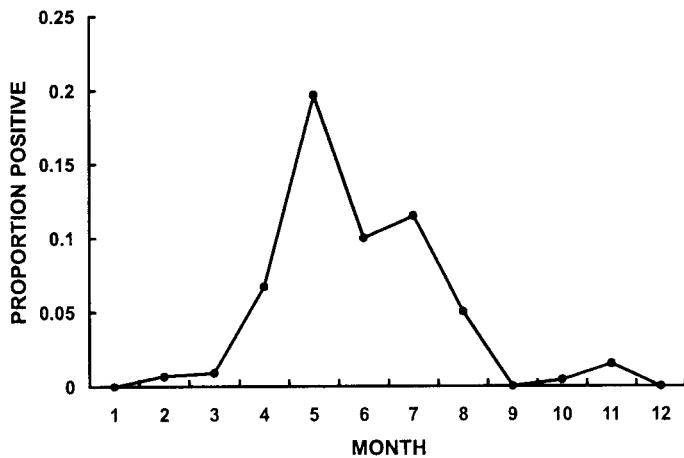
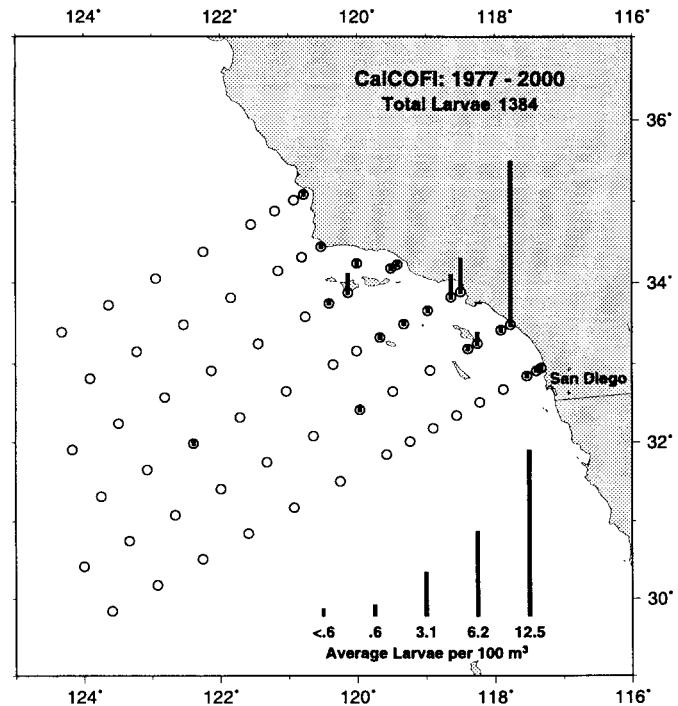
Jack mackerel



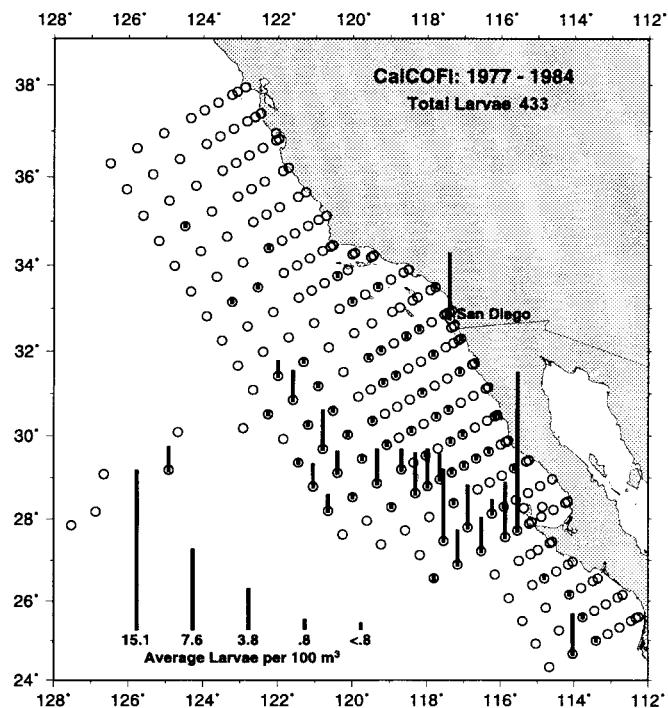
ATHERINIDAE



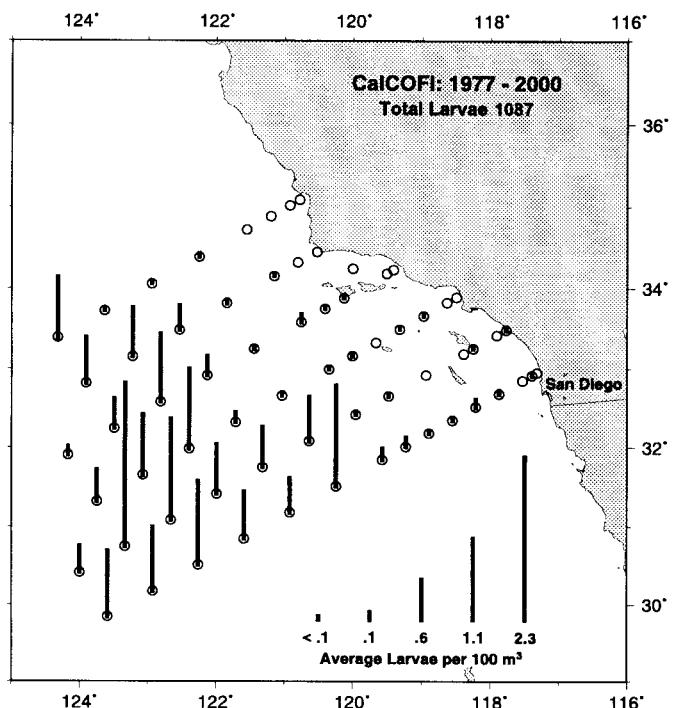
California grunion



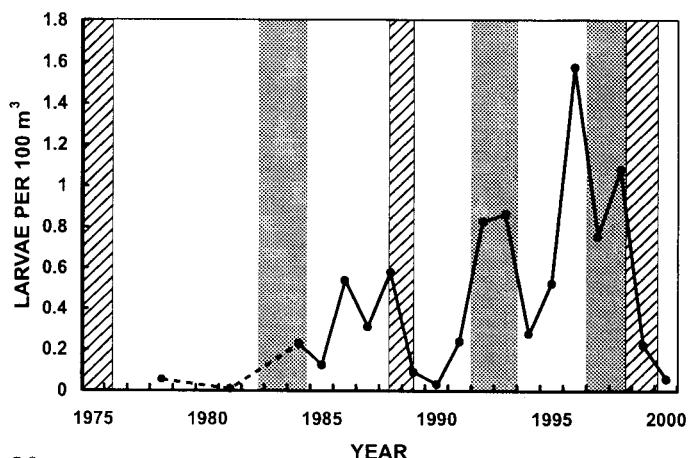
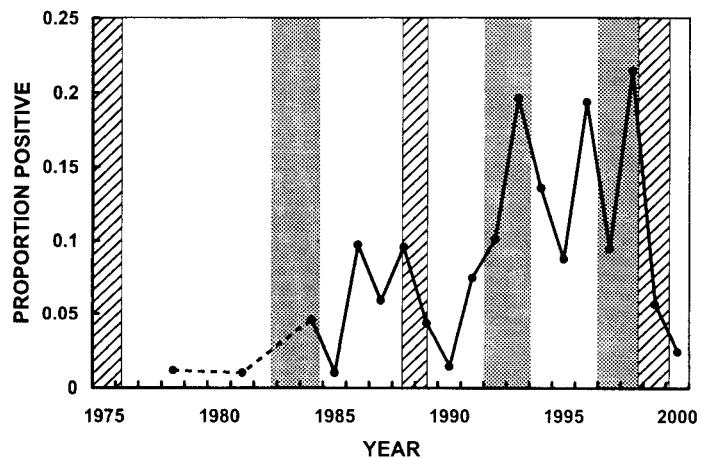
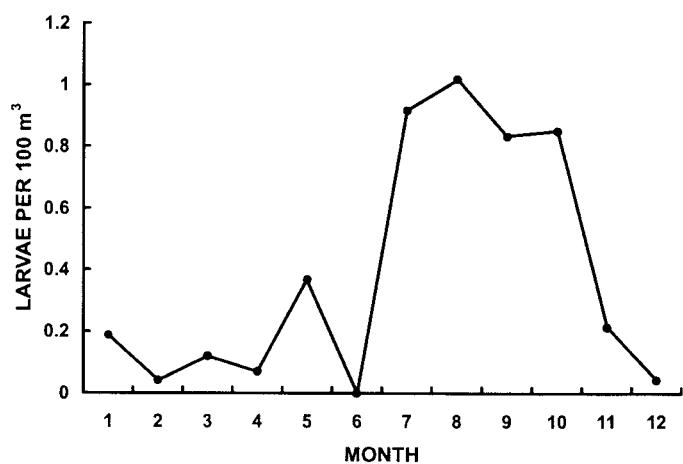
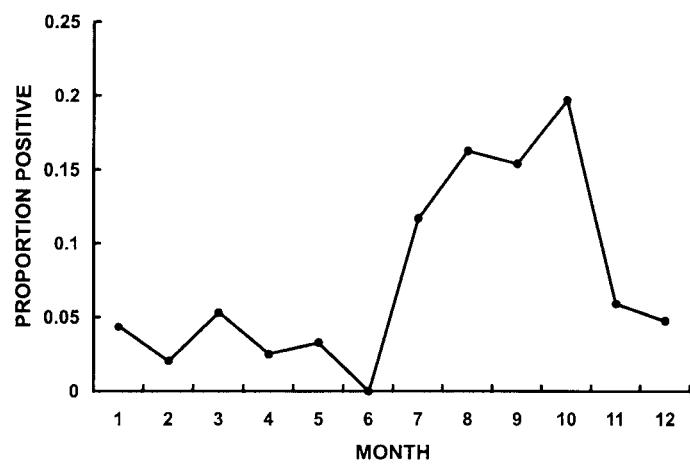
Vinciguerria lucetia



Panama lightfish



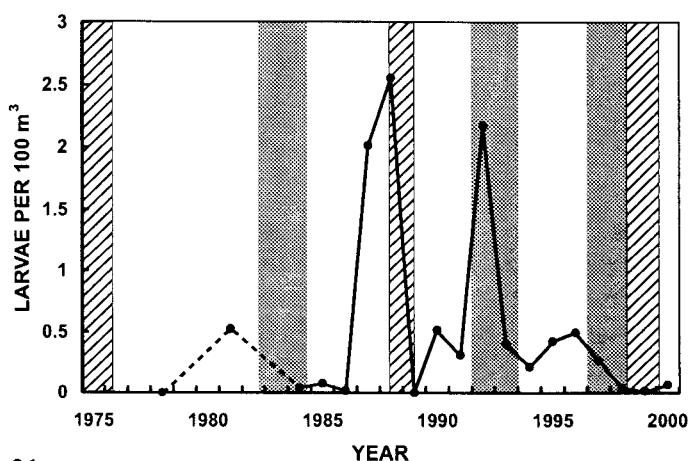
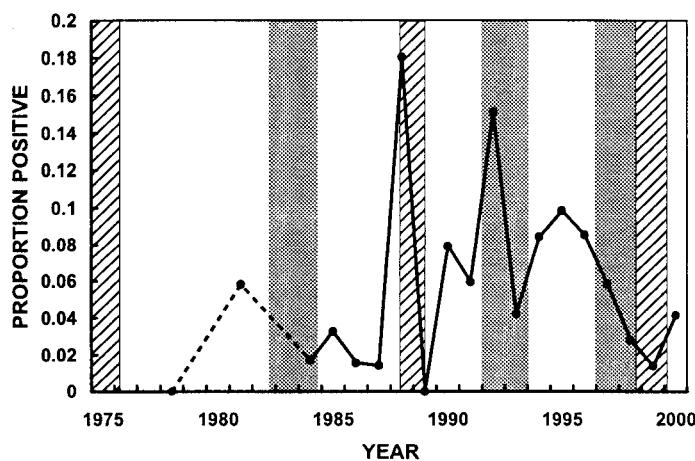
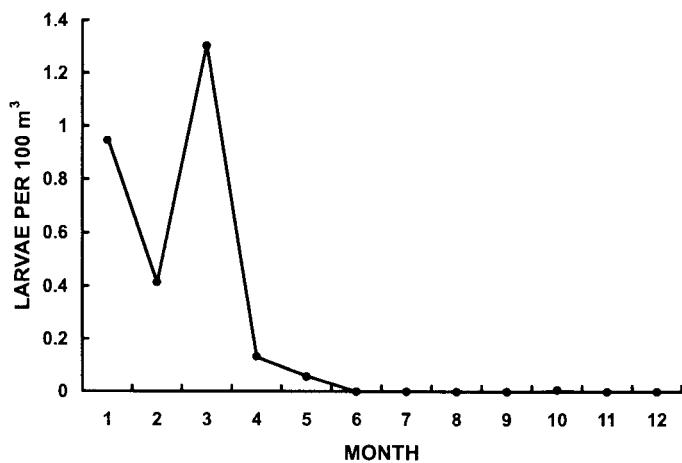
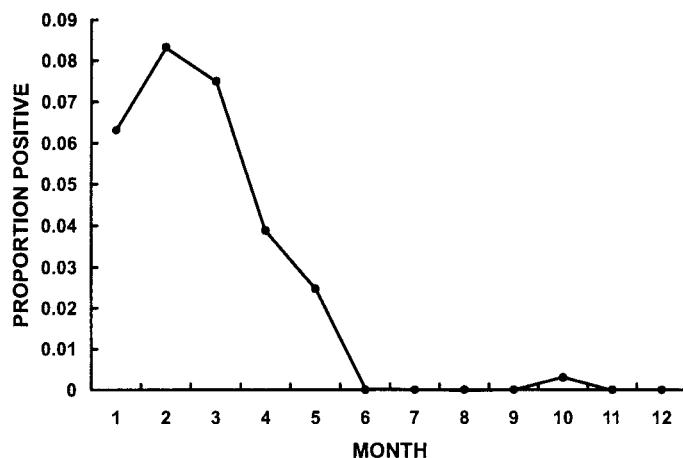
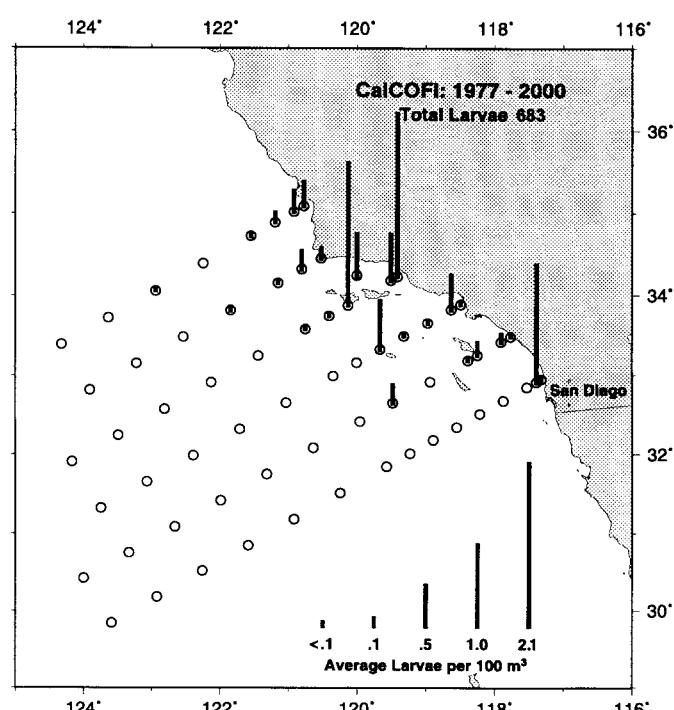
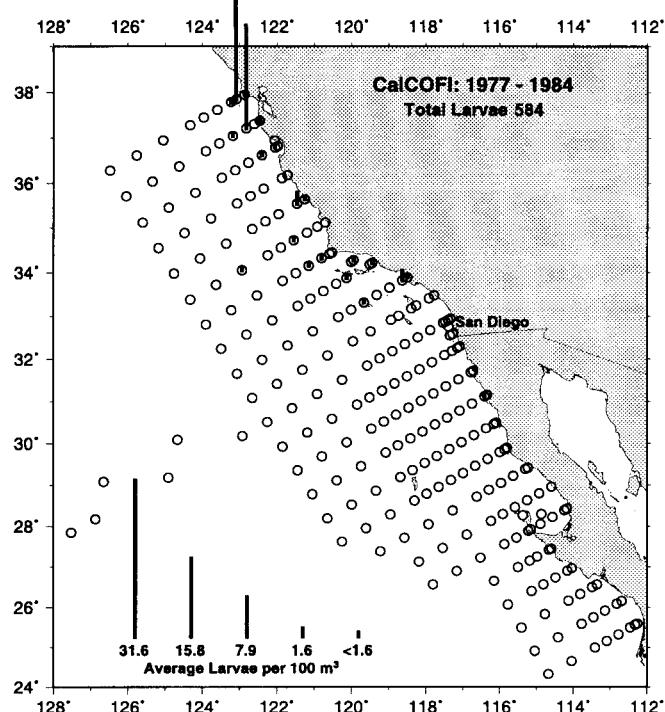
PHOSICHTHYIDAE



SEBASTIDAE

Shortbelly rockfish

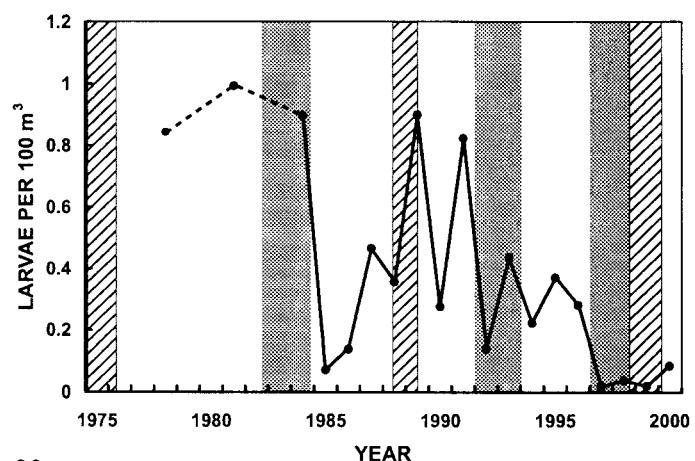
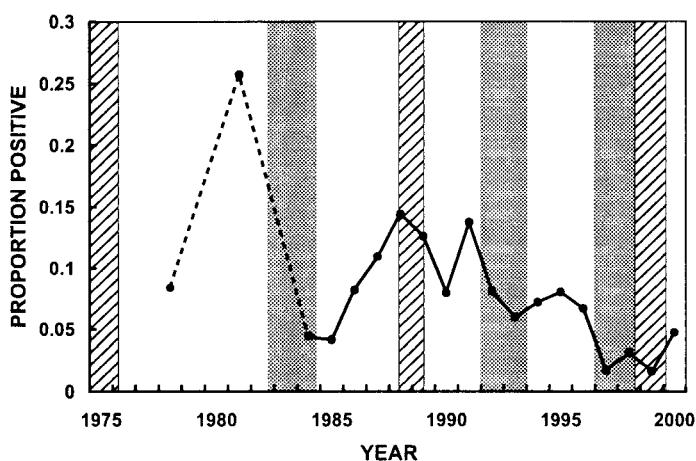
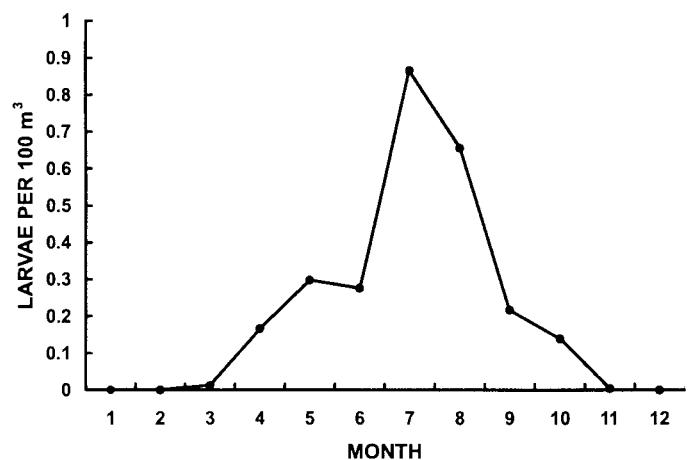
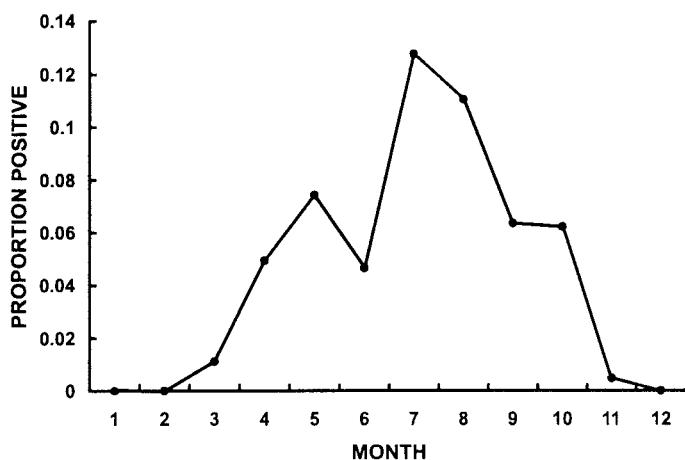
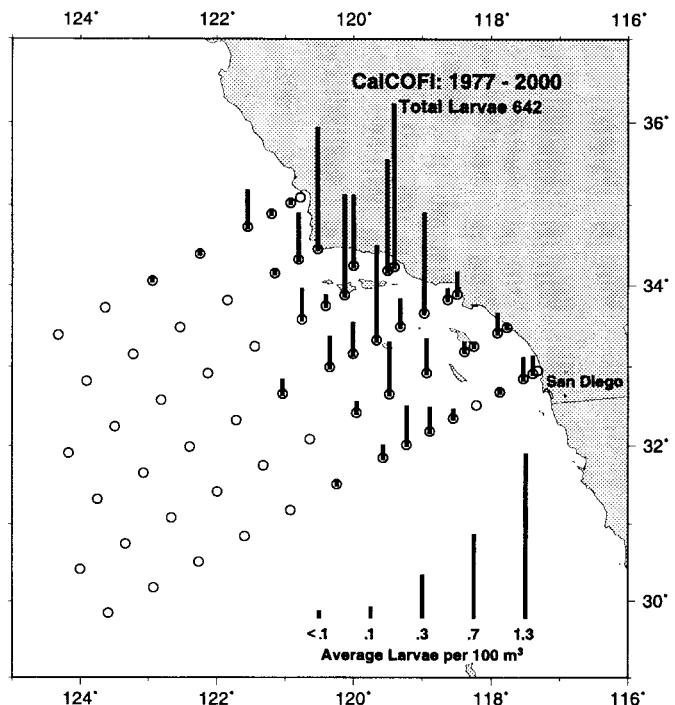
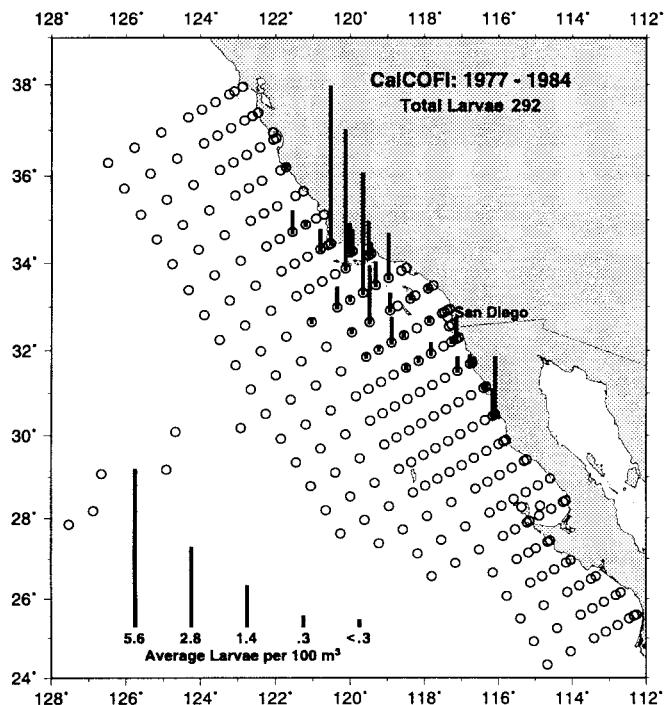
Sebastes jordani



Oxyjulis californica

Señorita

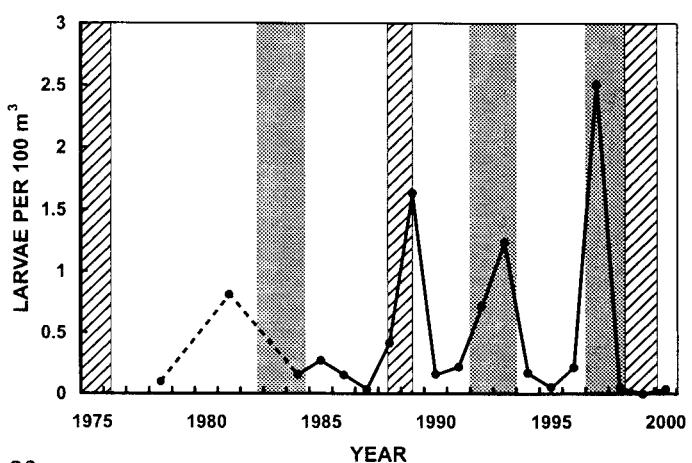
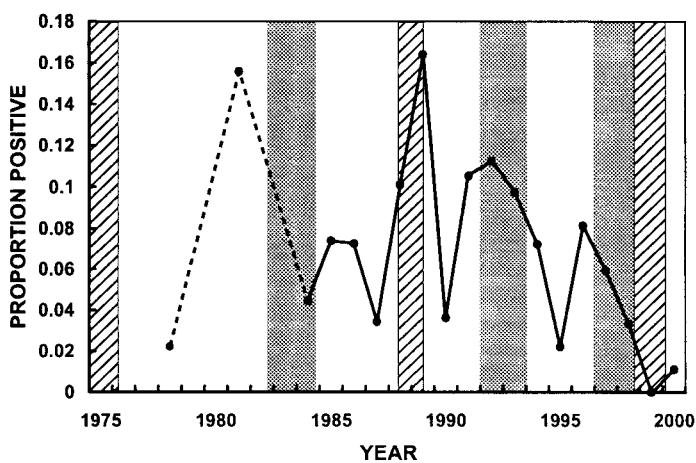
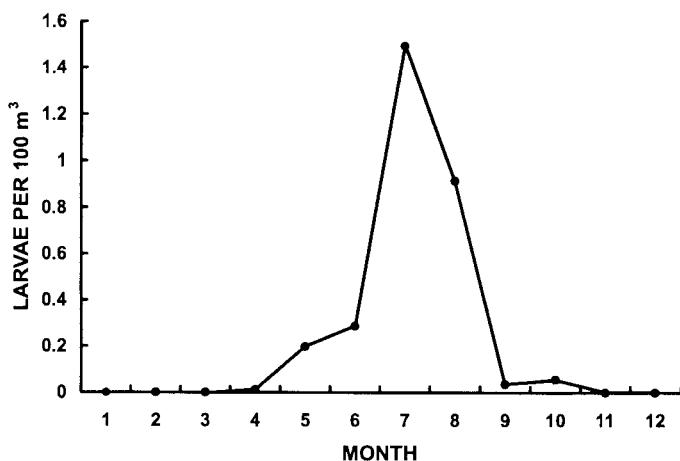
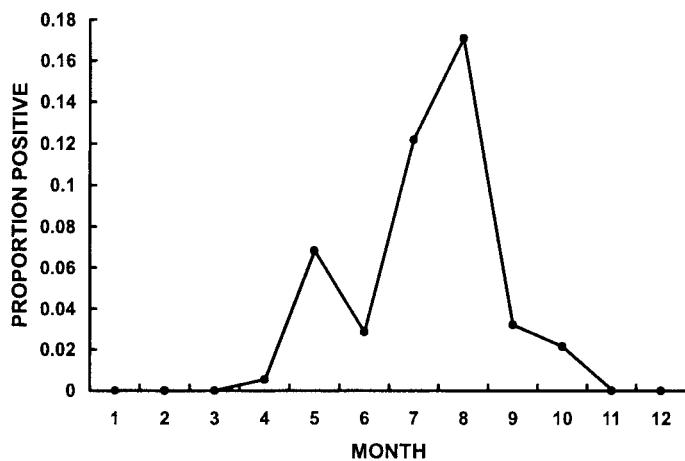
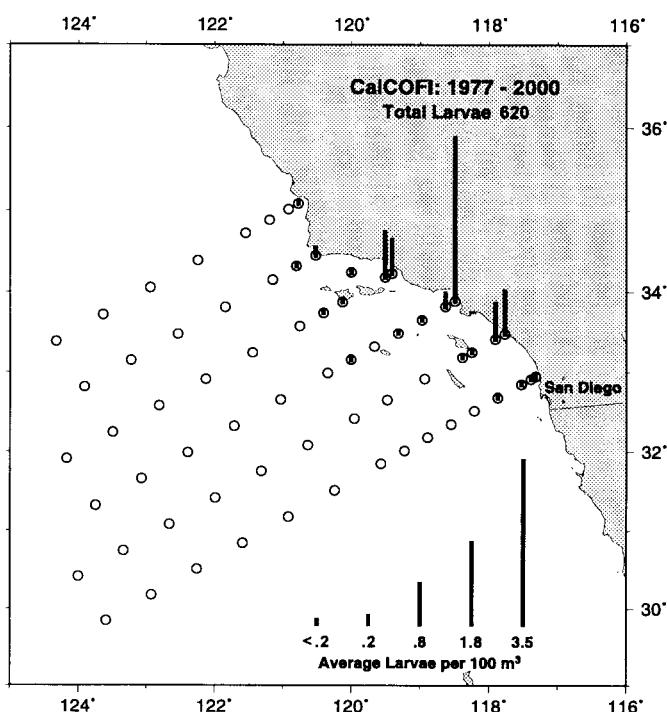
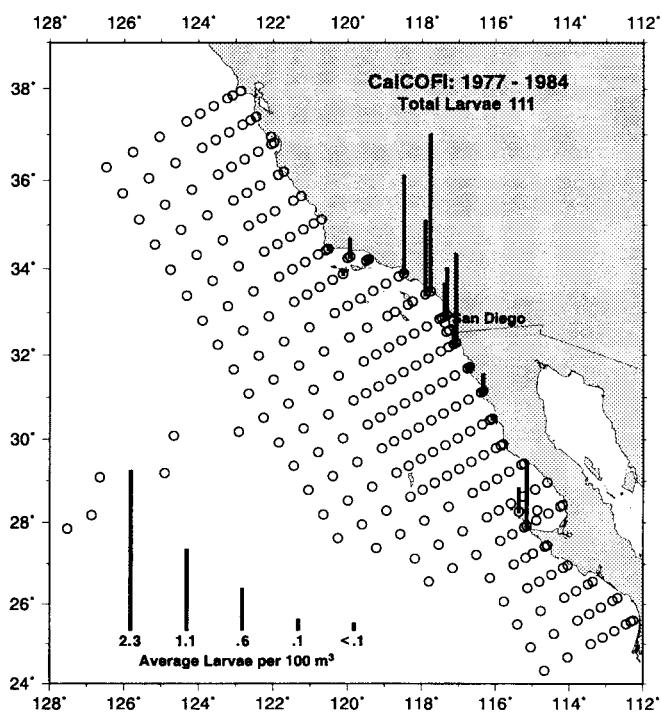
LABRIDAE



SPHYRAENIDAE

Pacific barracuda

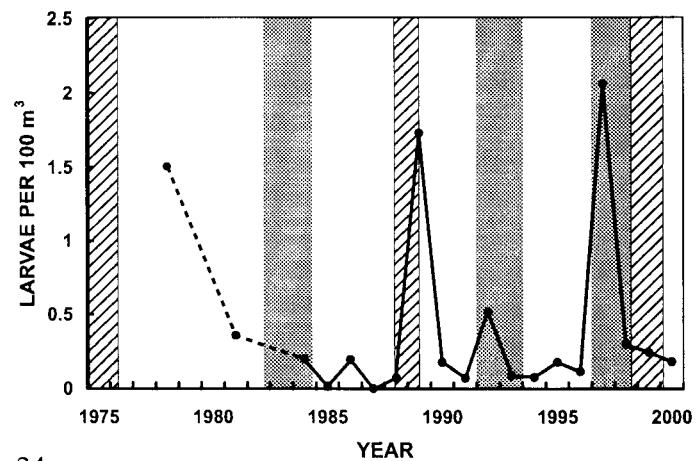
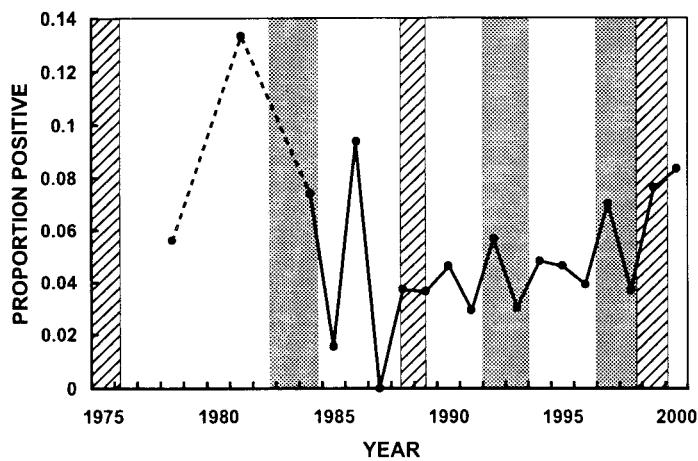
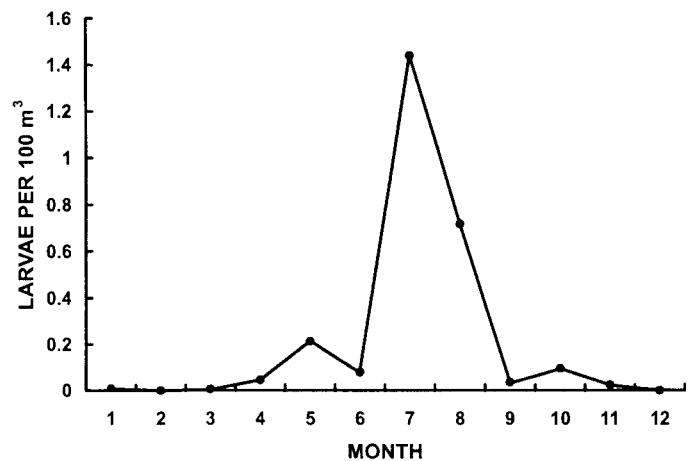
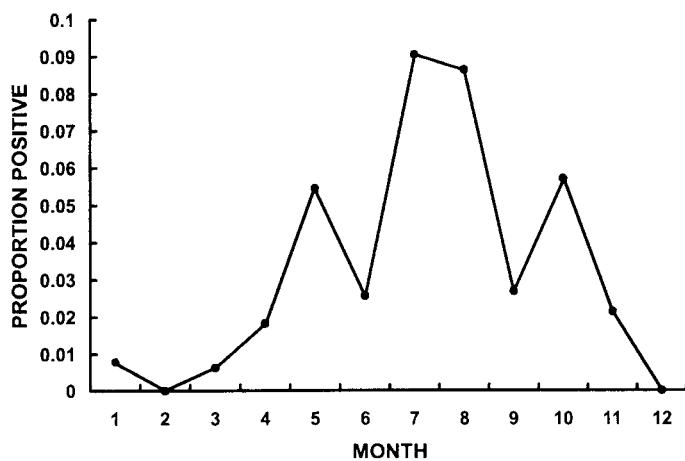
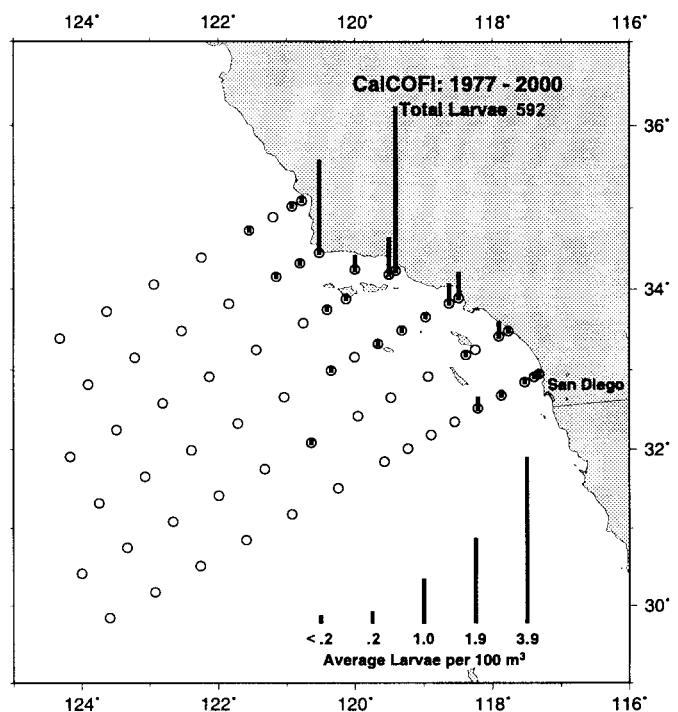
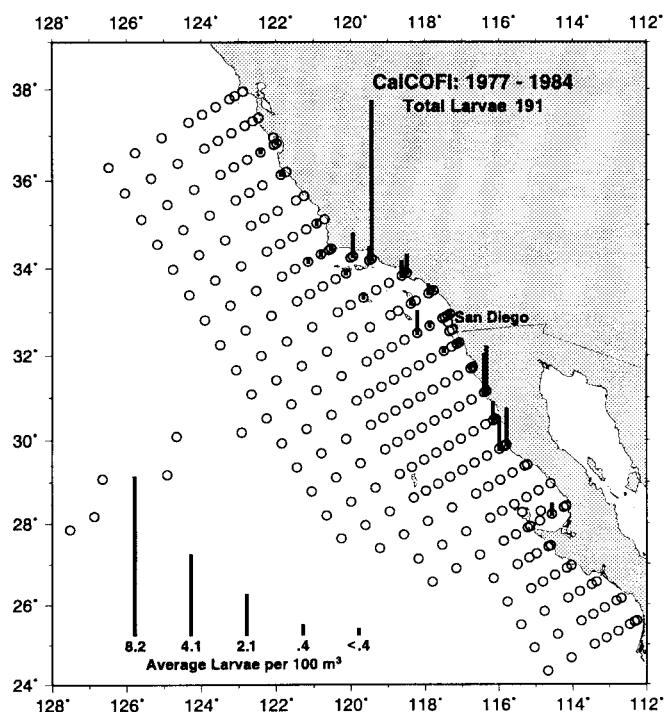
Sphyraena argentea



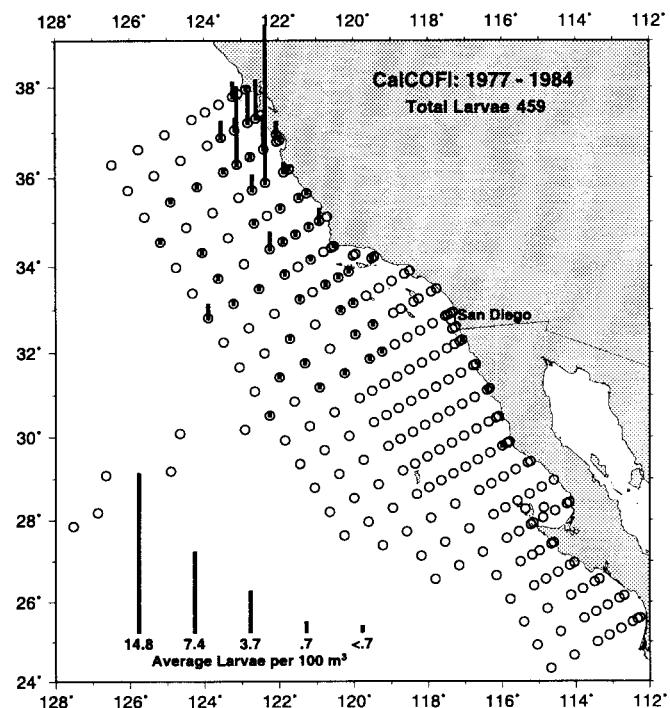
Hypsoblennius gilberti

Rockpool blenny

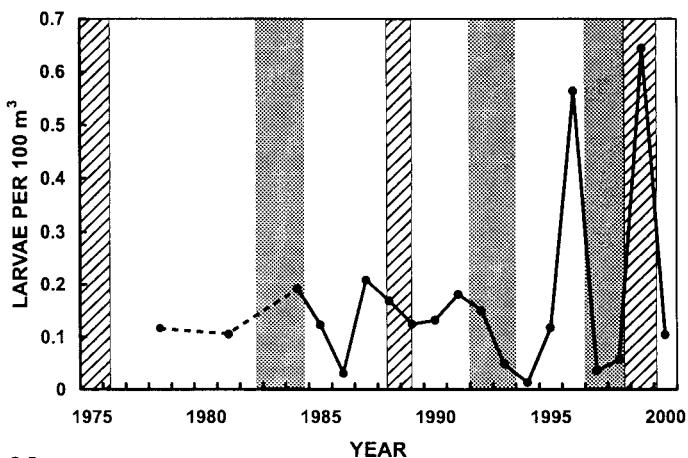
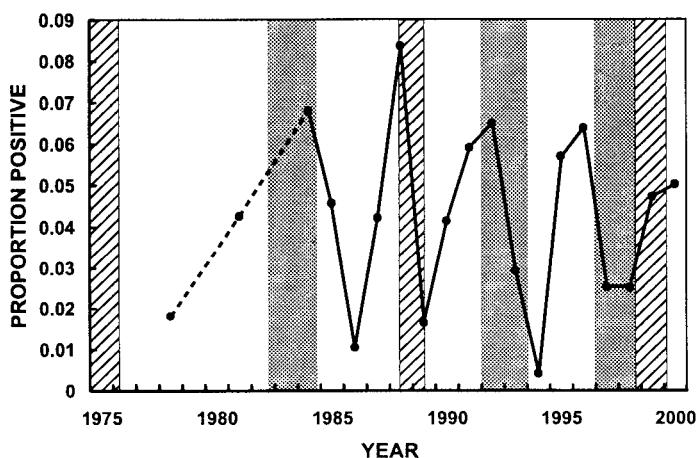
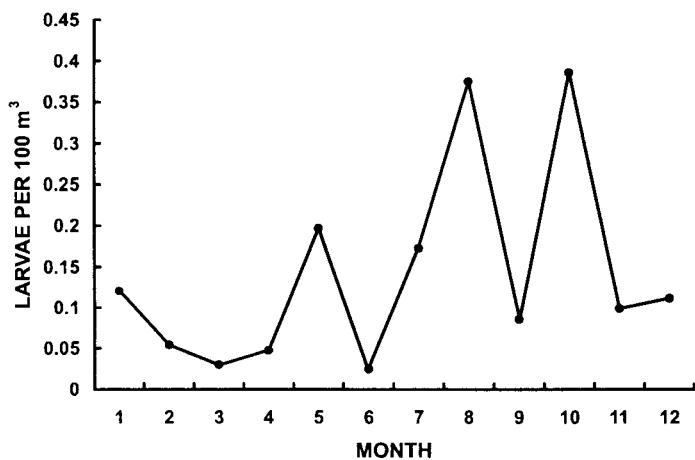
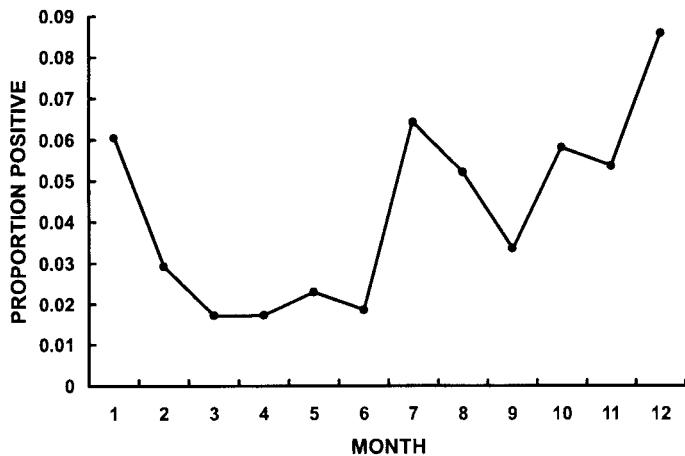
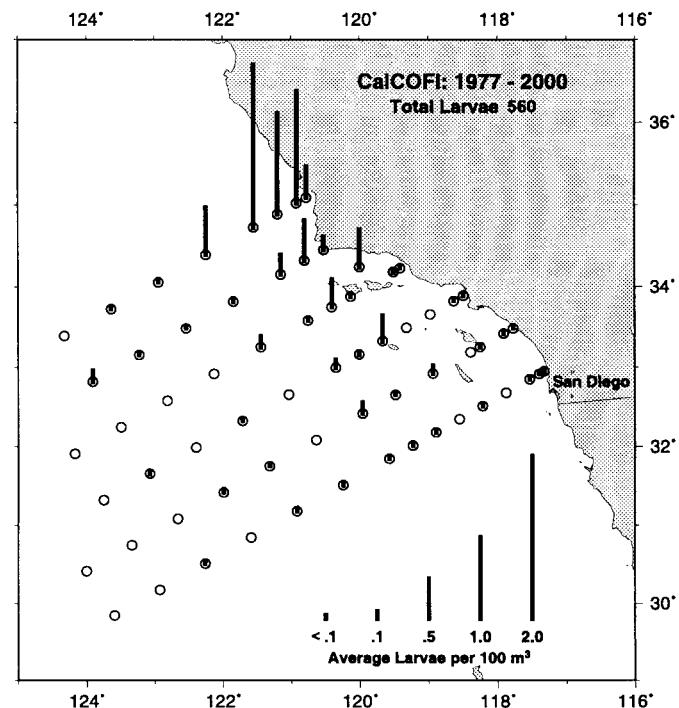
BLENNIIDAE



SEBASTIDAE



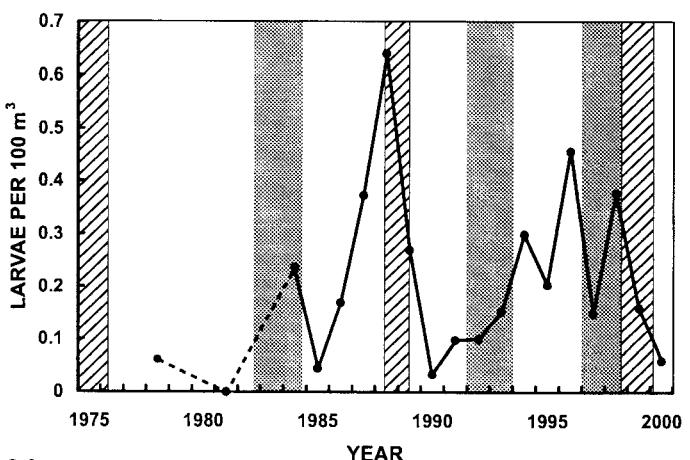
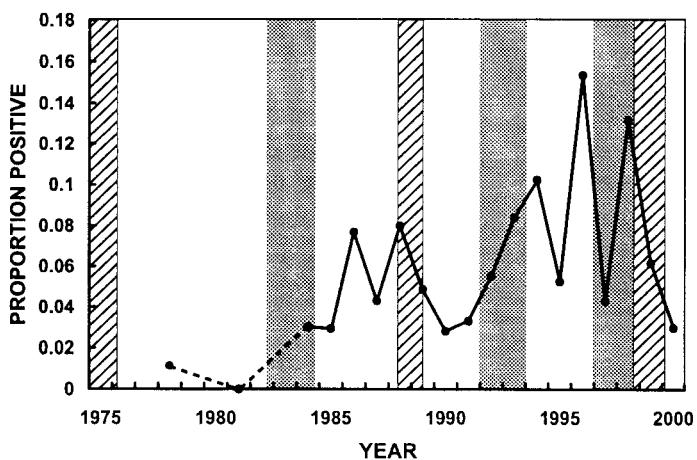
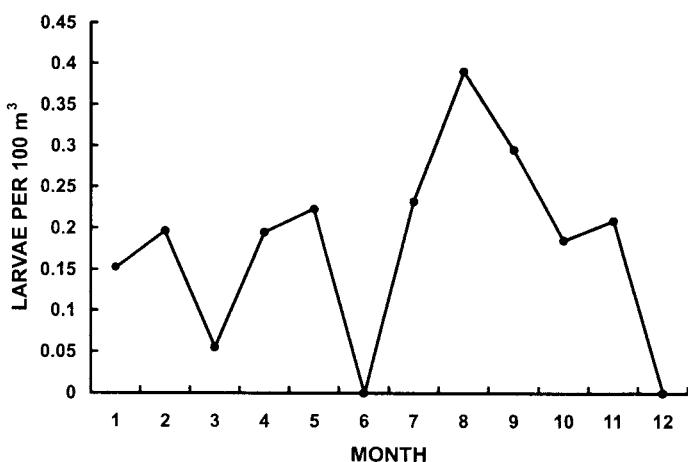
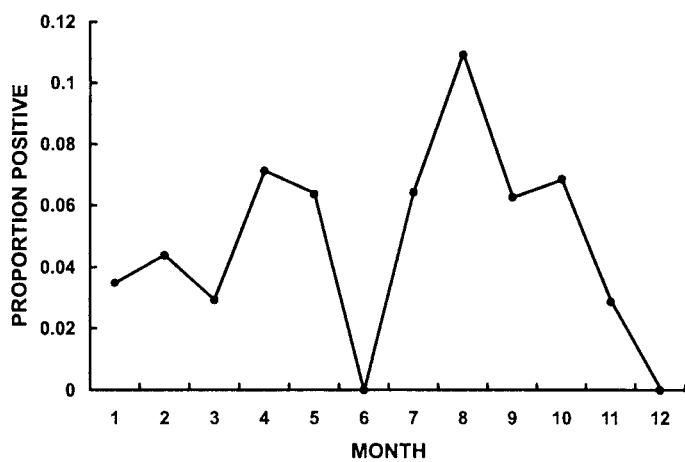
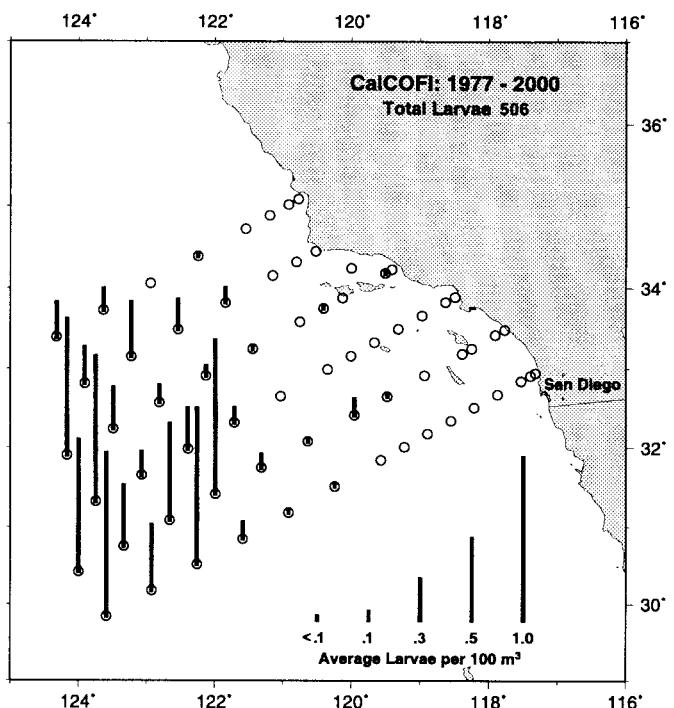
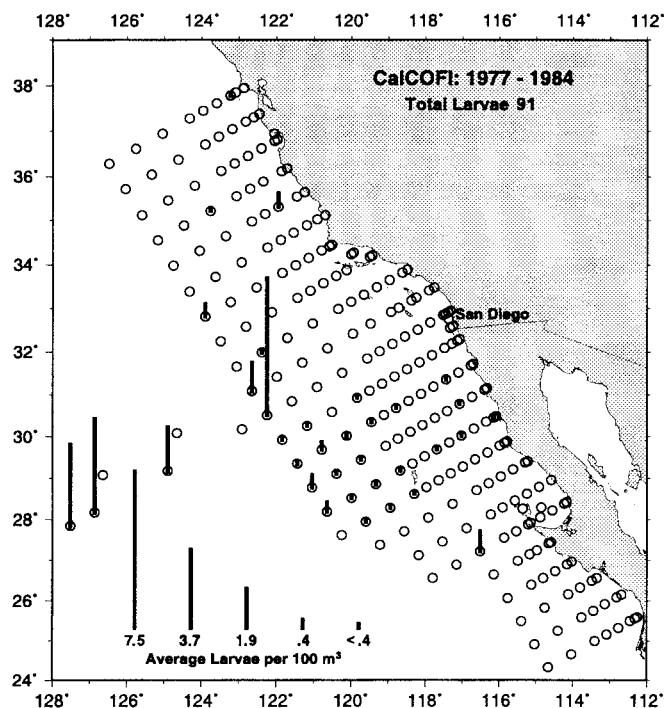
Splitnose rockfish



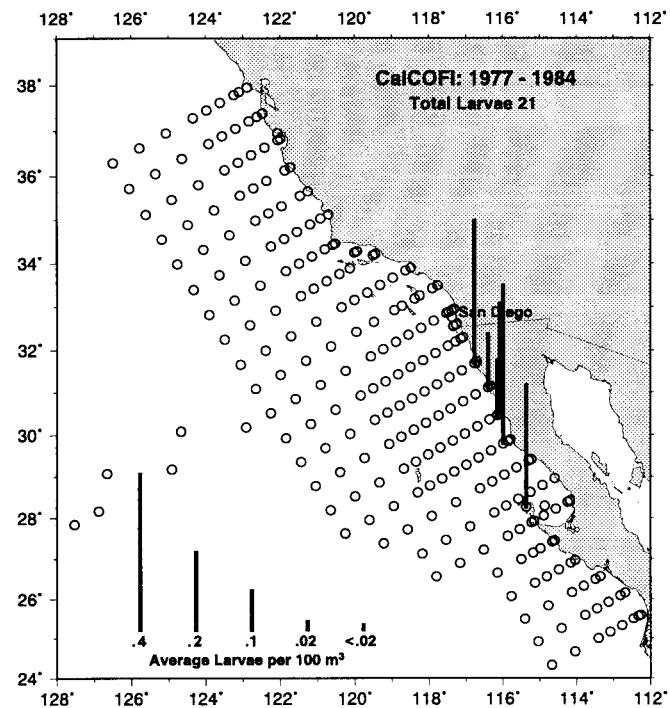
Ceratoscopelus townsendi

Dogtooth lampfish

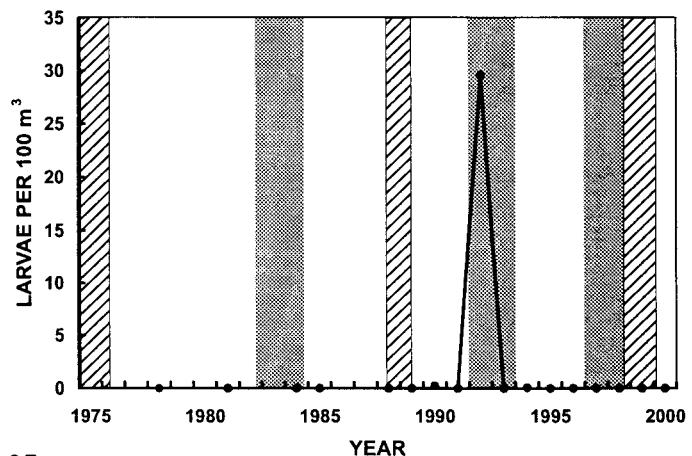
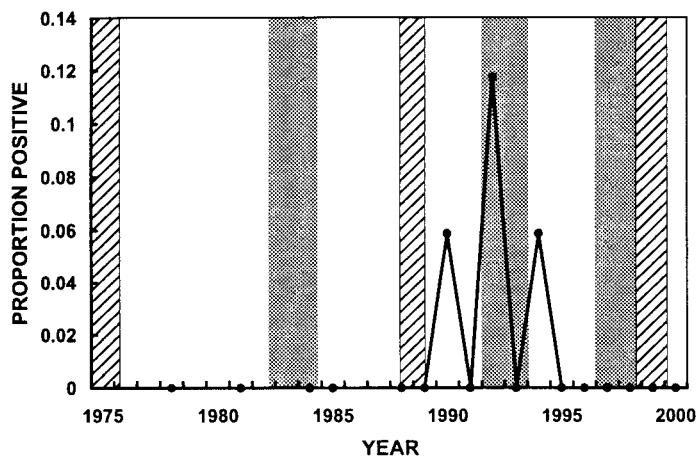
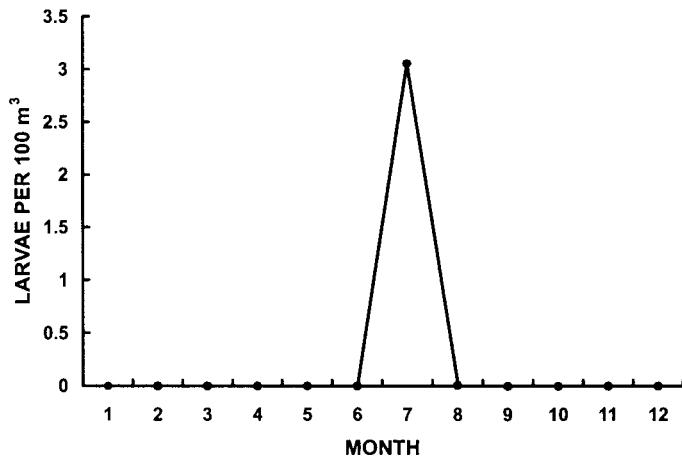
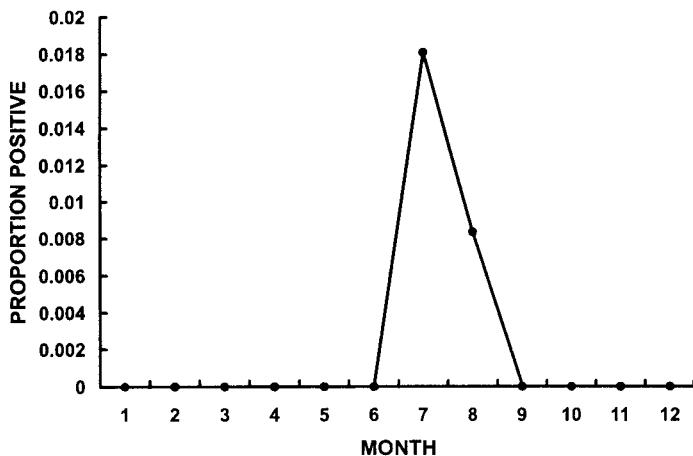
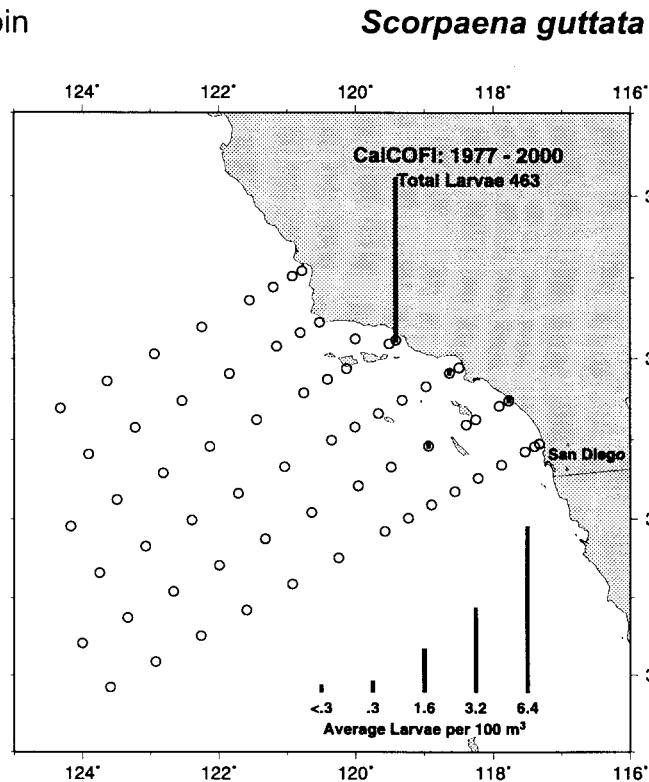
MYCTOPHIDAE



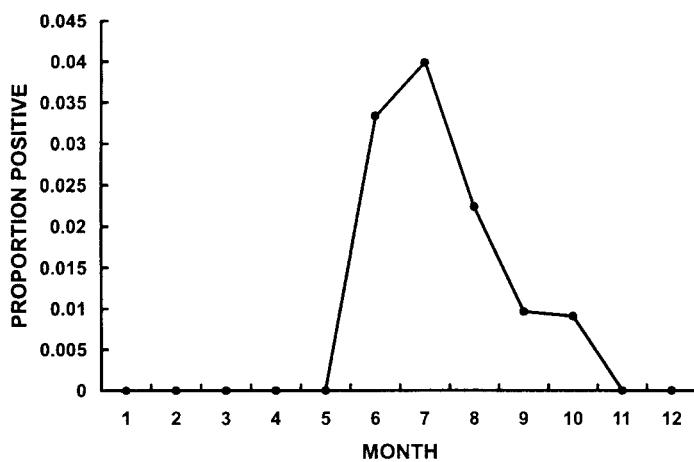
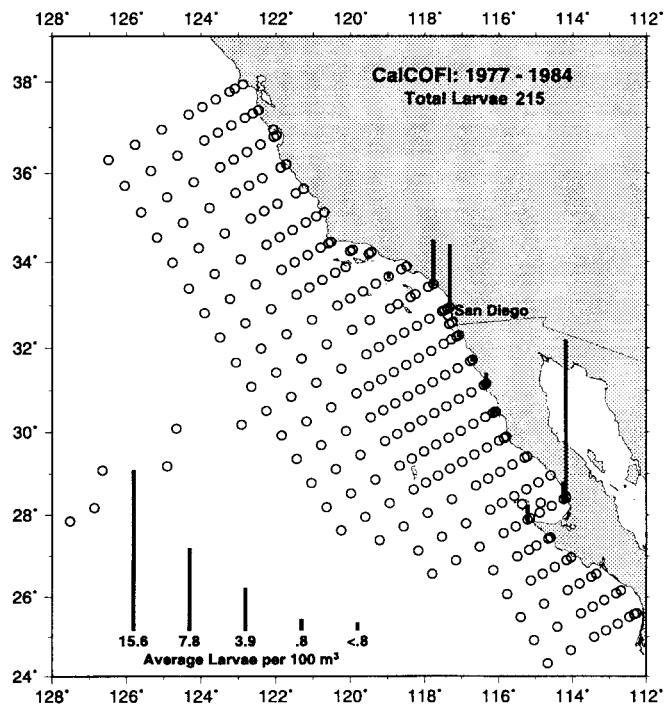
SCORPAENIDAE



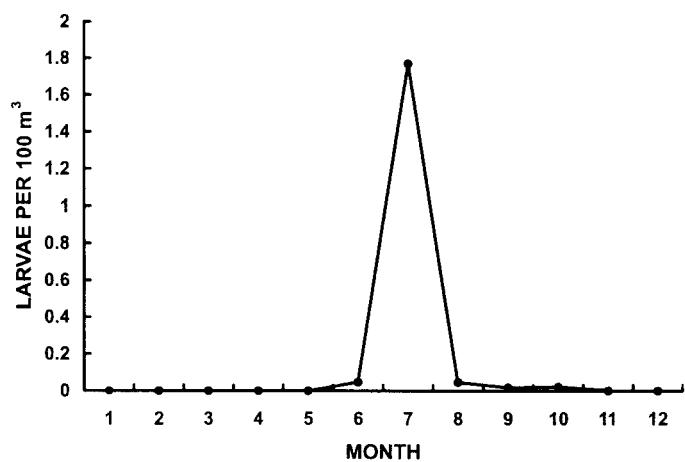
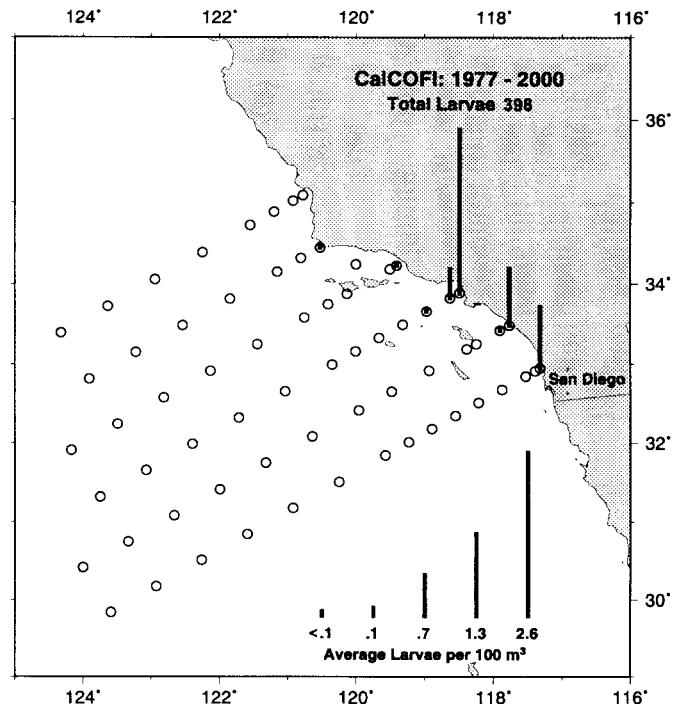
Sculpin



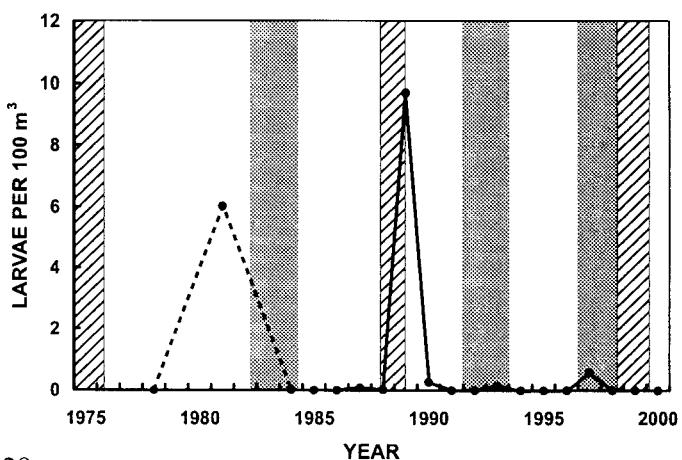
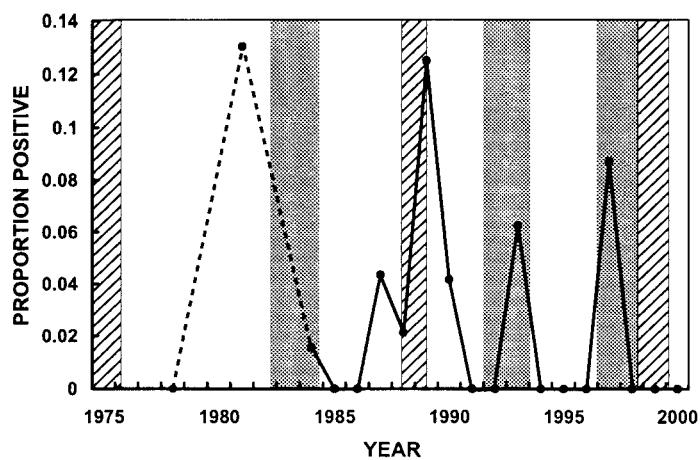
Xenistius californiensis



Salema



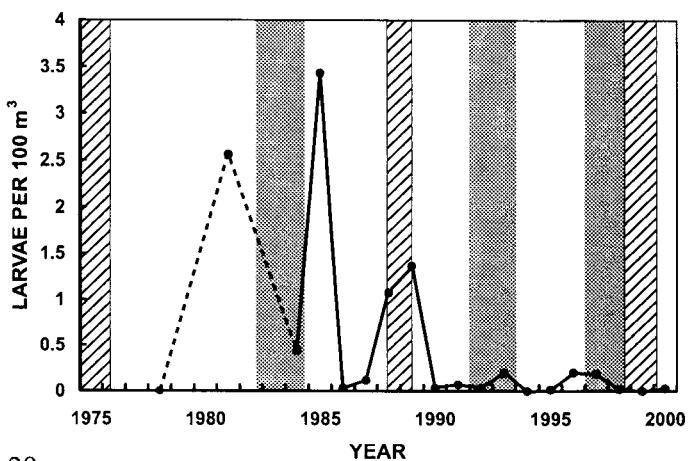
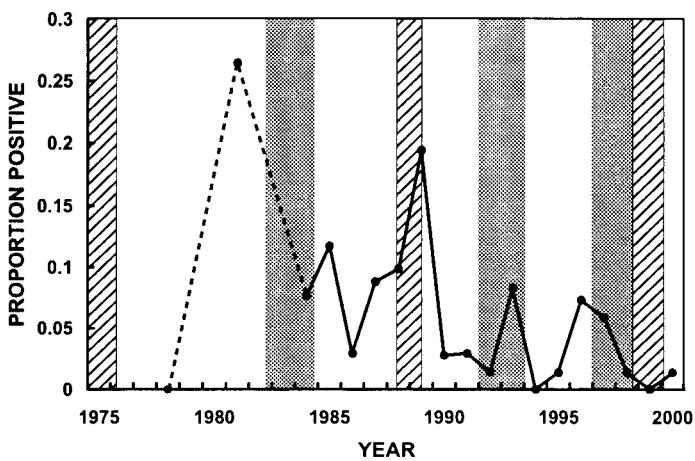
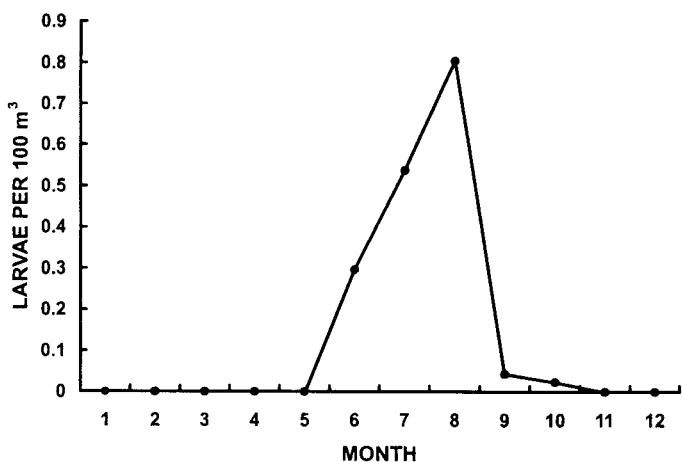
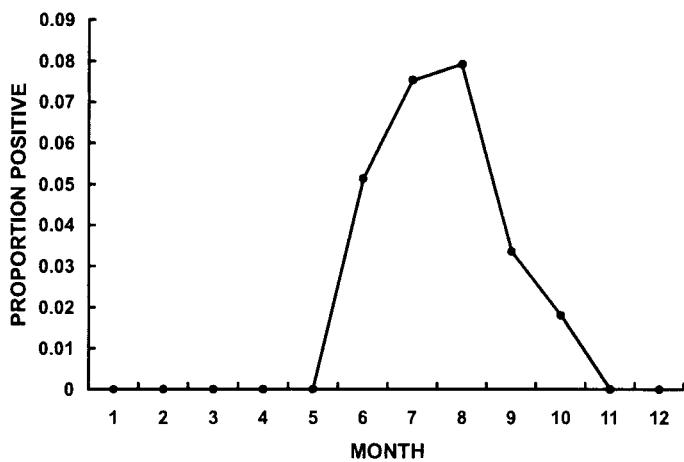
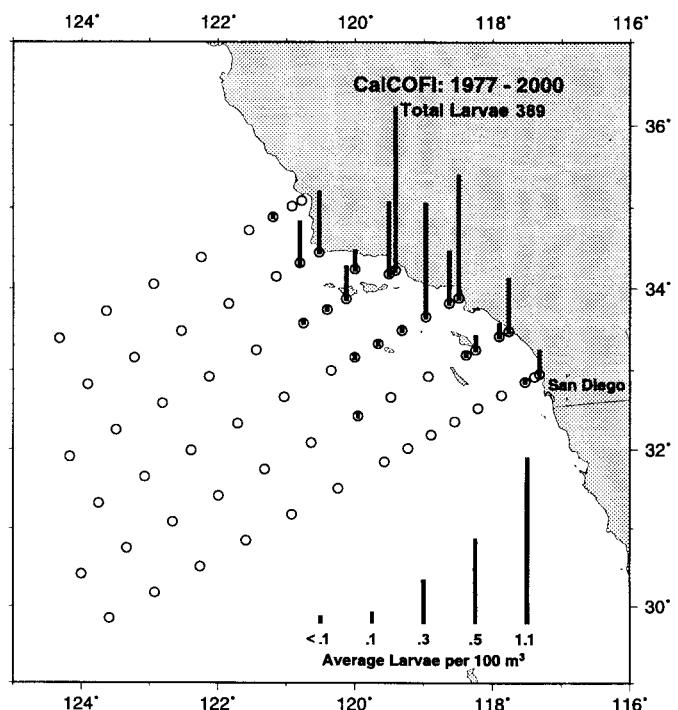
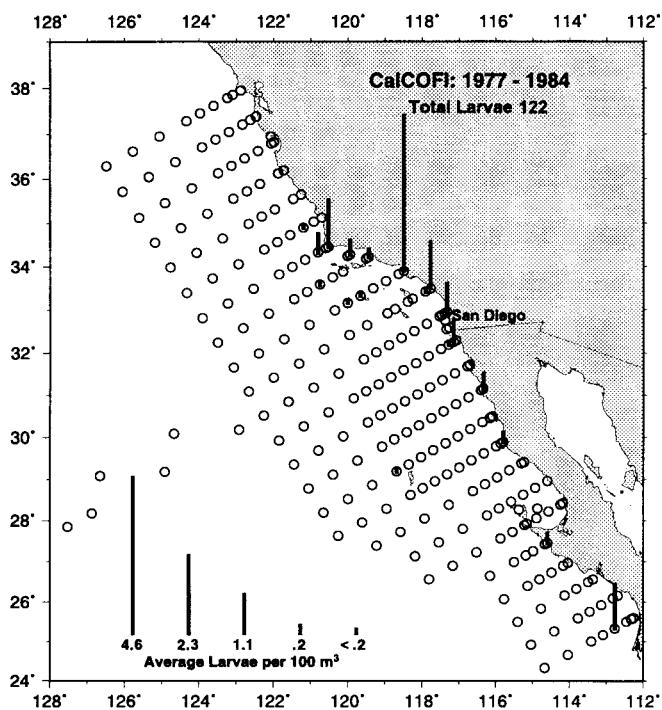
HAEMULIDAE



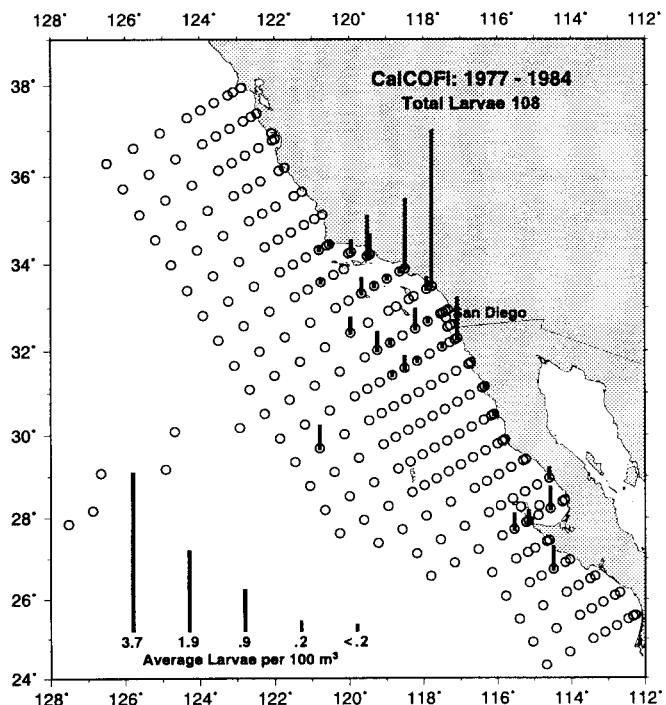
SERRANIDAE

Kelp and sand basses

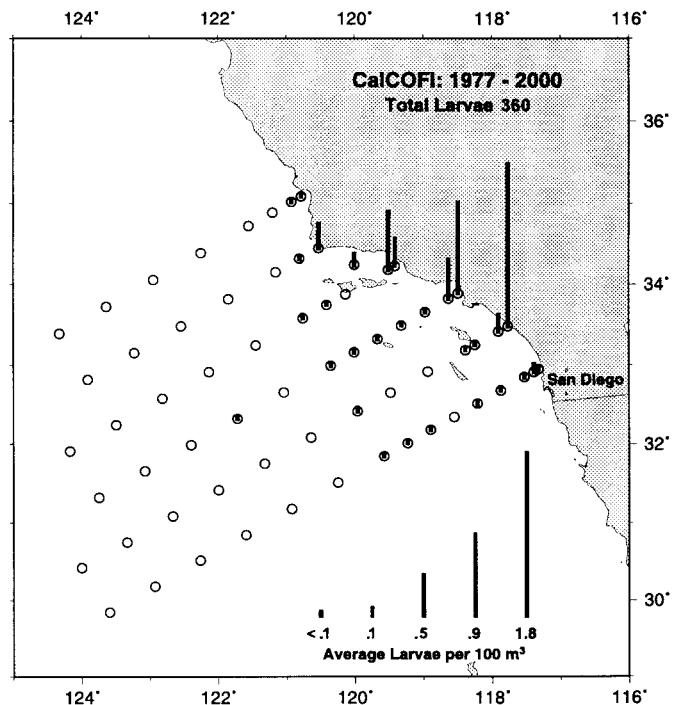
Paralabrax spp.



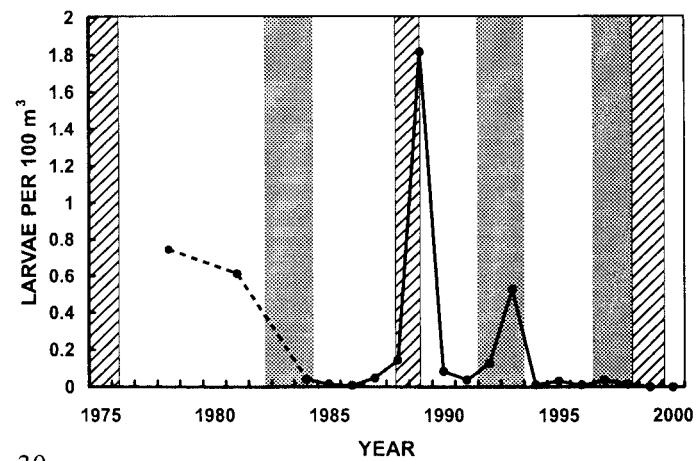
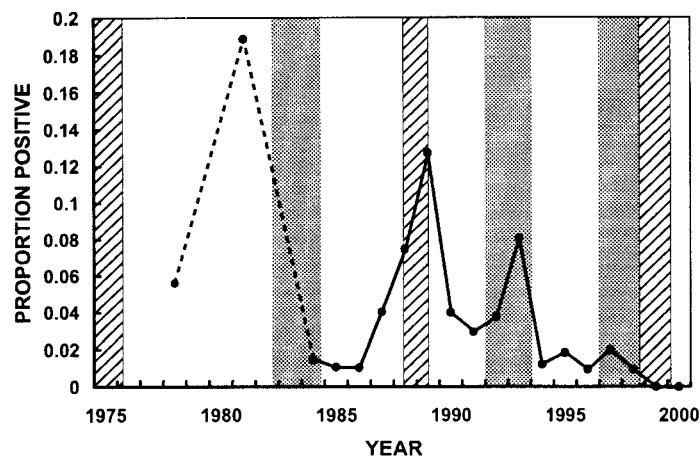
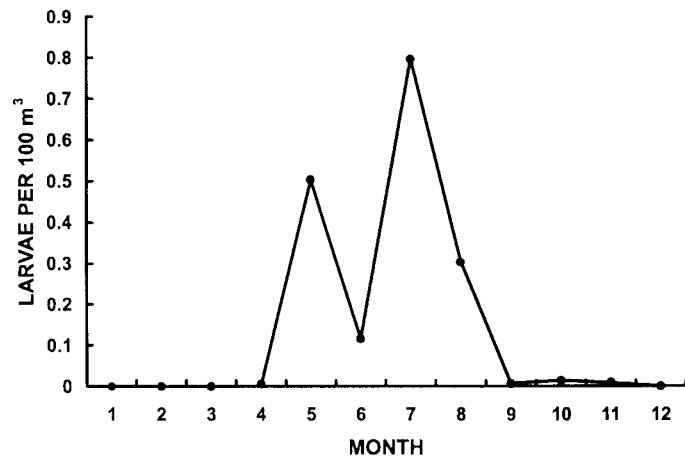
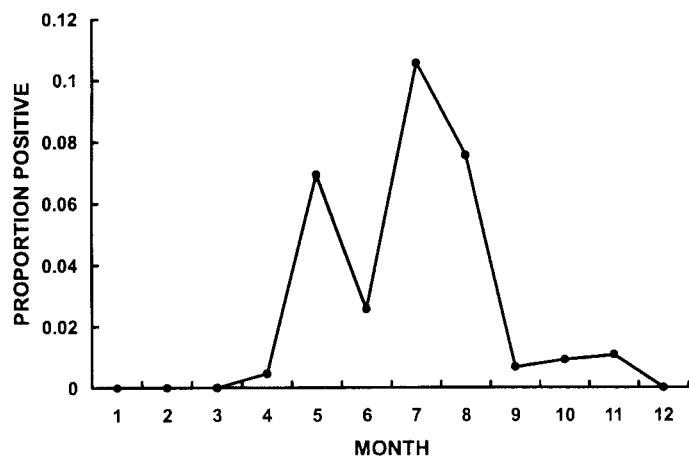
Girella nigricans



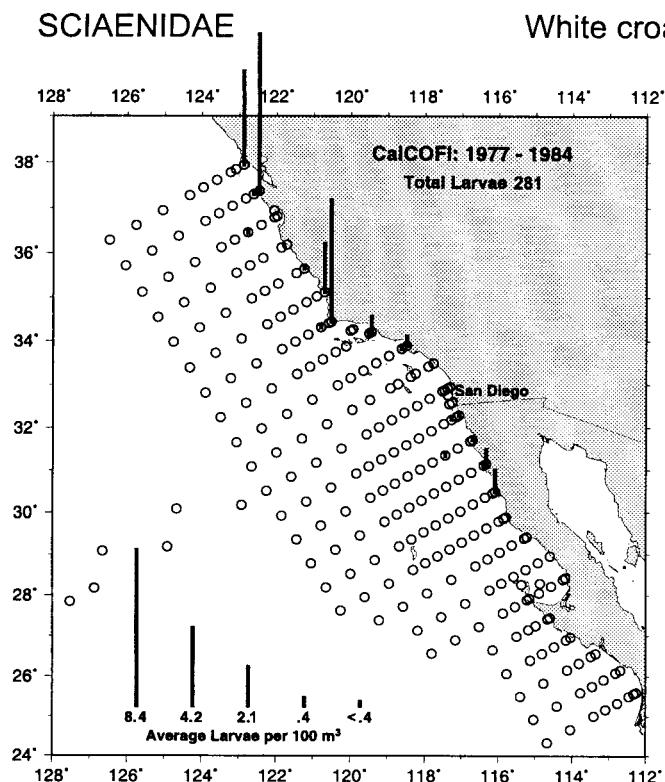
Opaleye



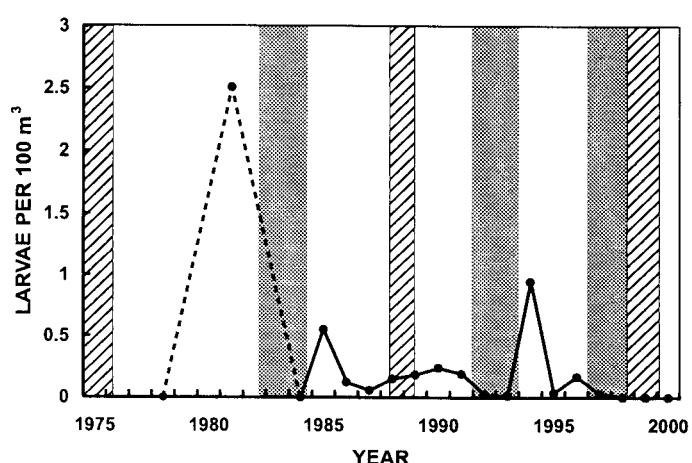
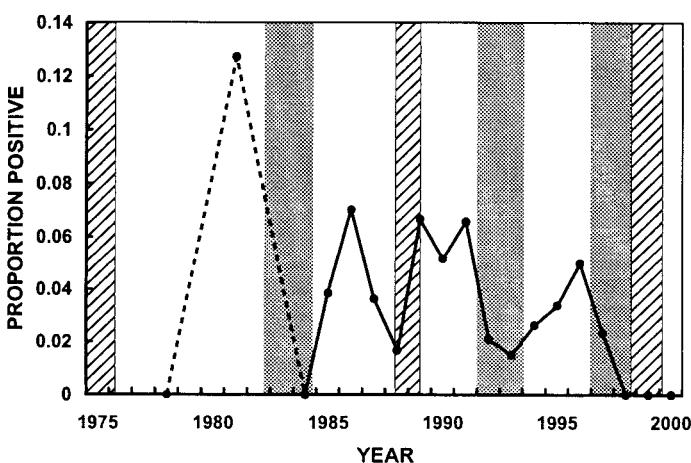
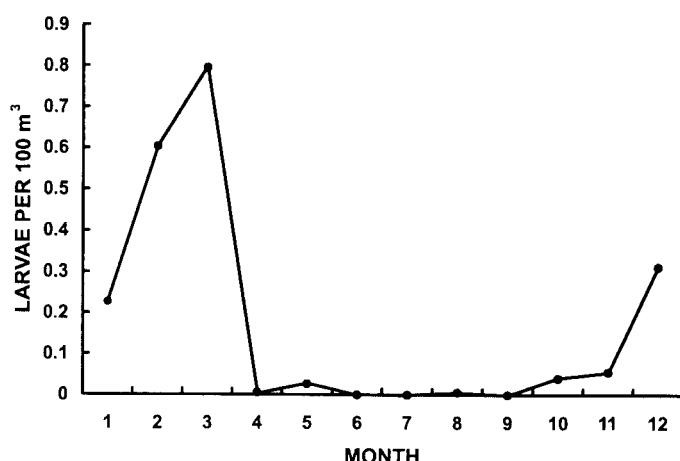
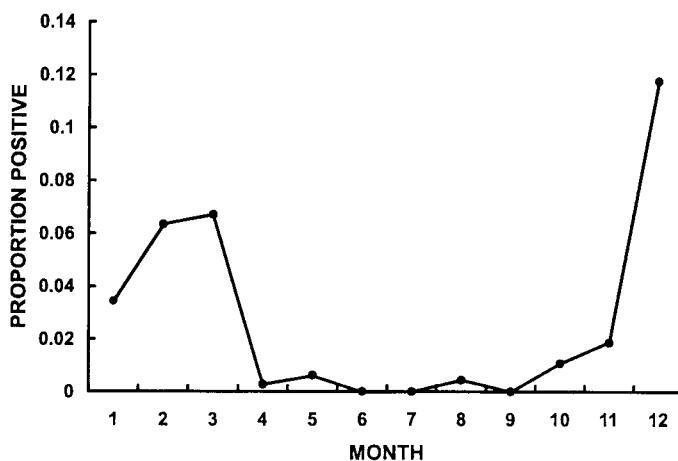
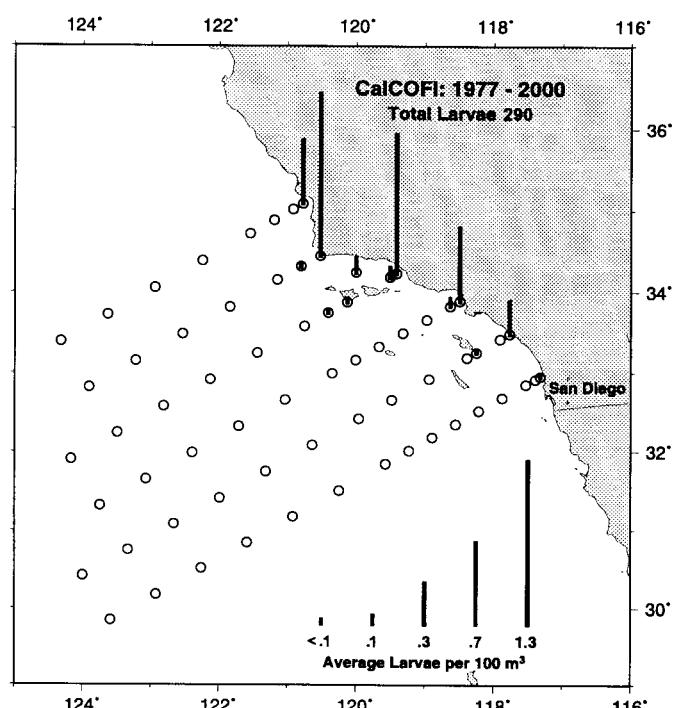
KYPHOSIDAE



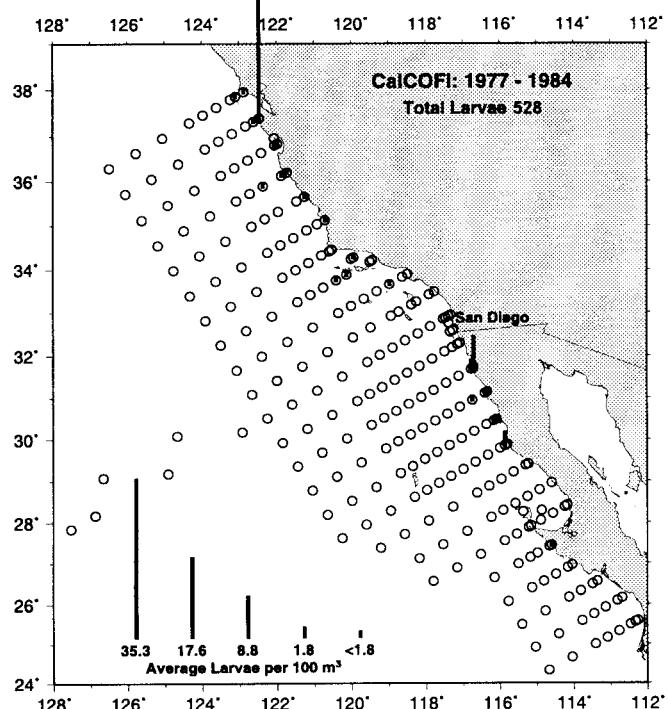
SCIENIDAE



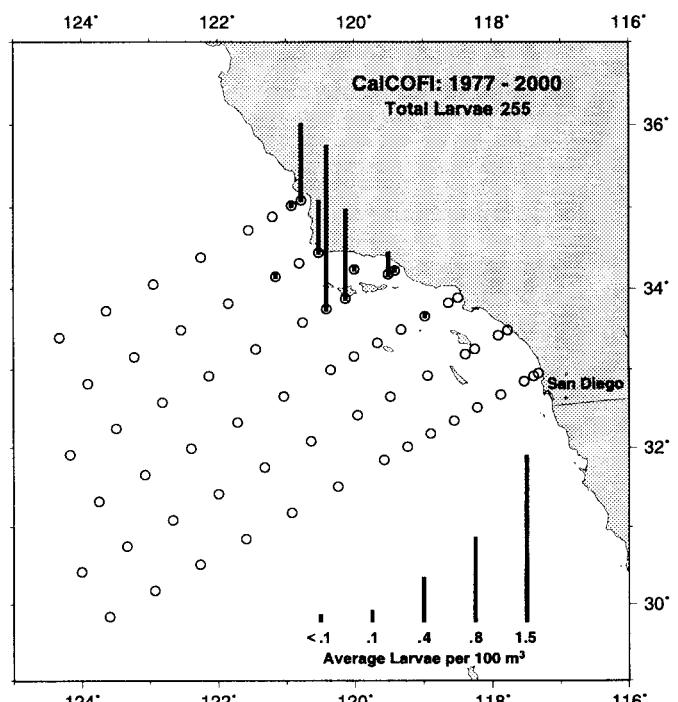
Genyonemus lineatus



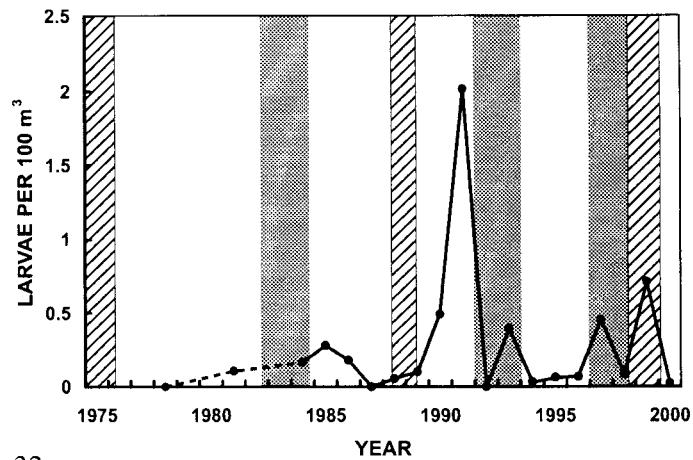
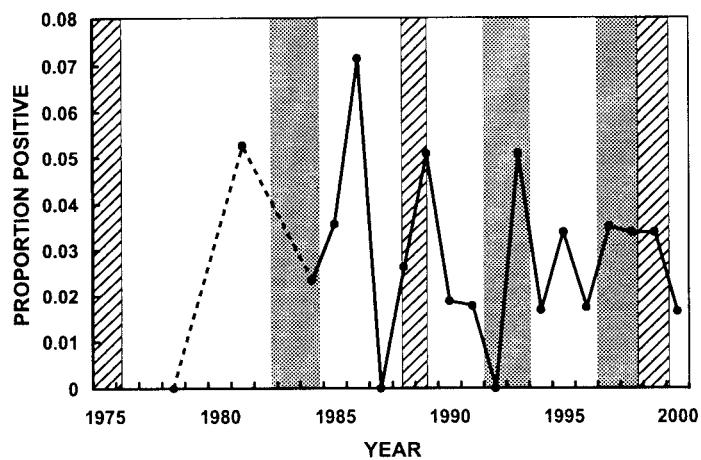
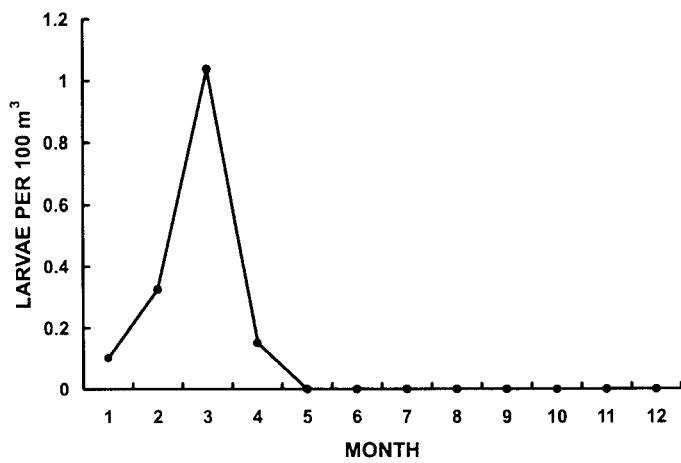
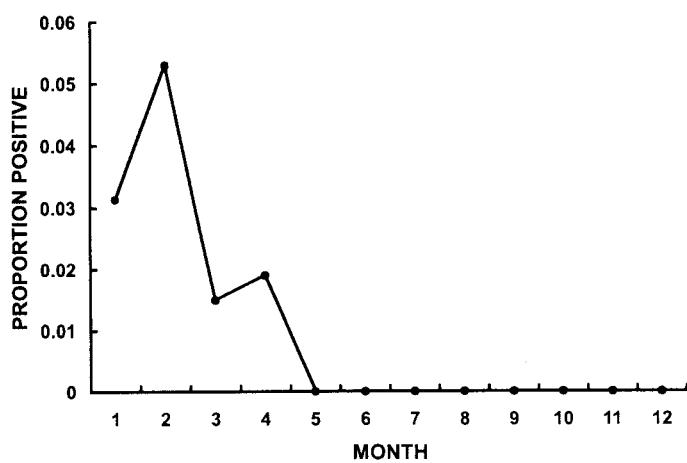
Ophiodon elongatus



Lingcod



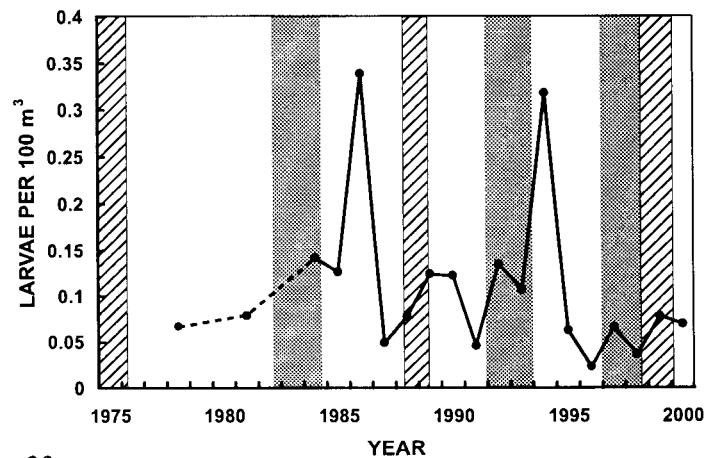
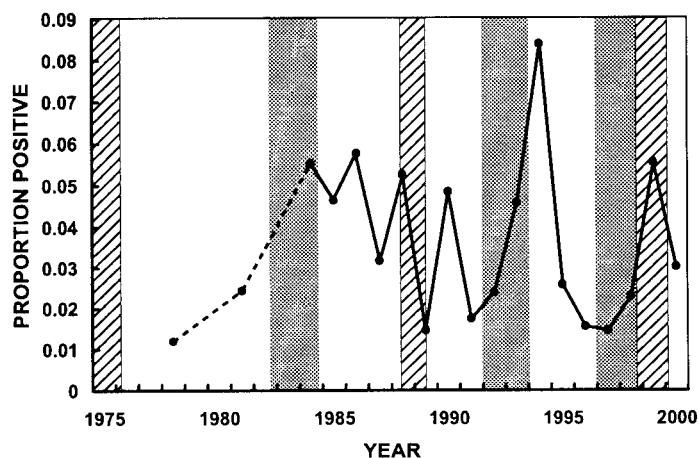
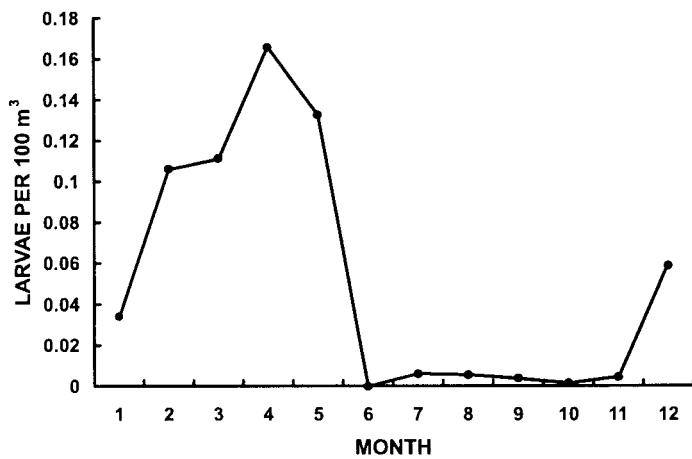
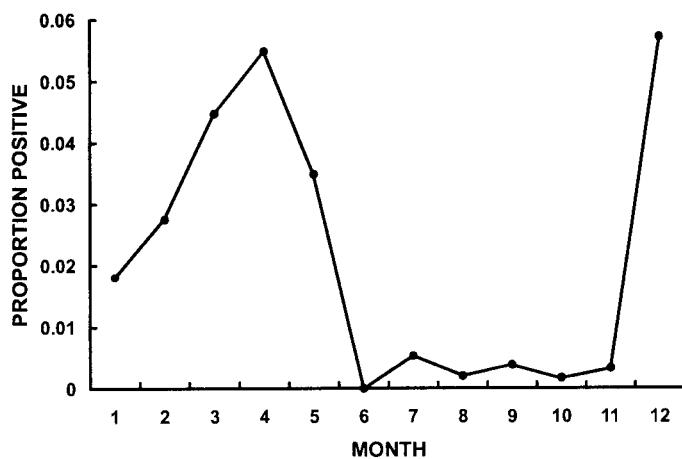
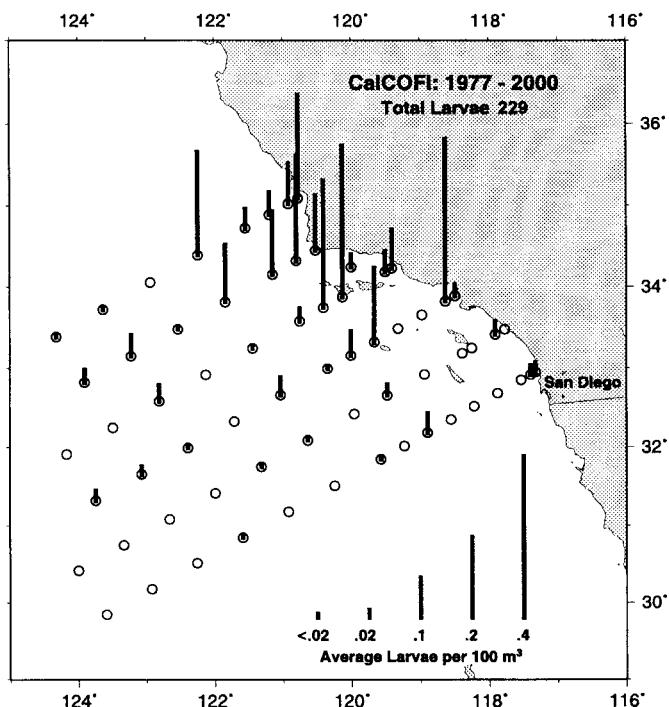
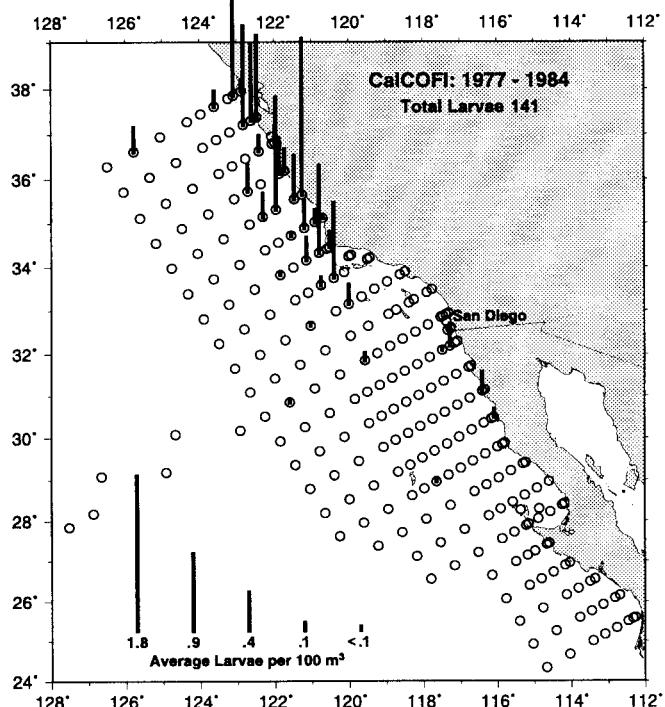
HEXAGRAMMIDAE



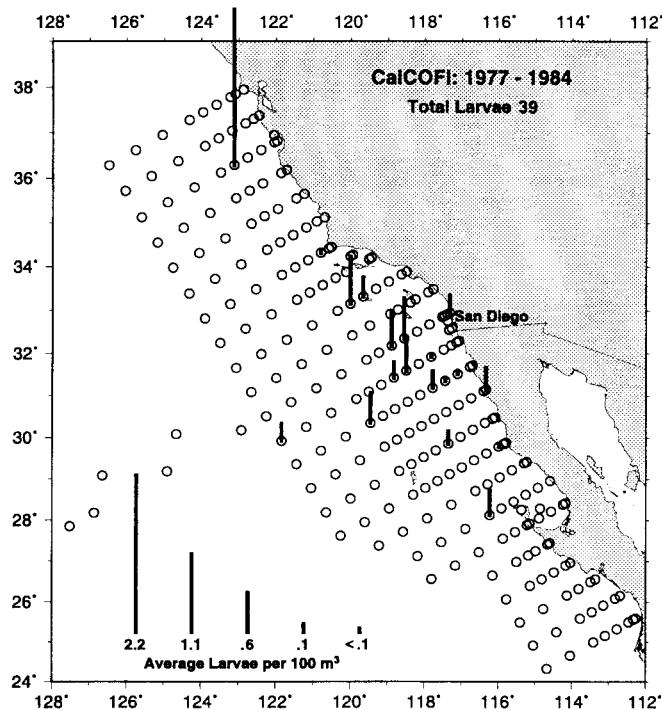
MYCTOPHIDAE

Northern lampfish

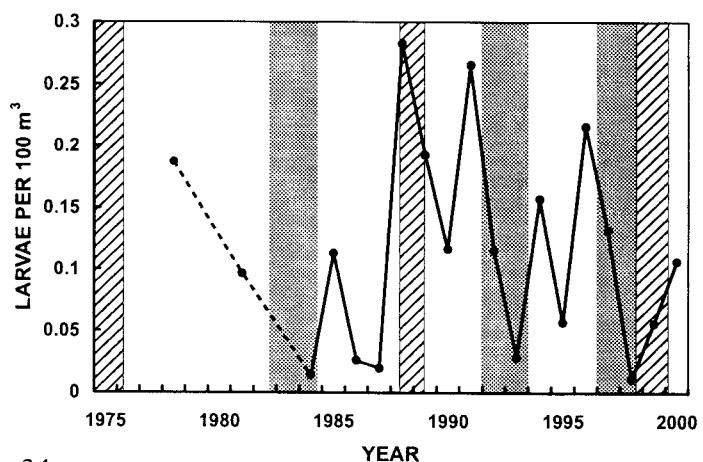
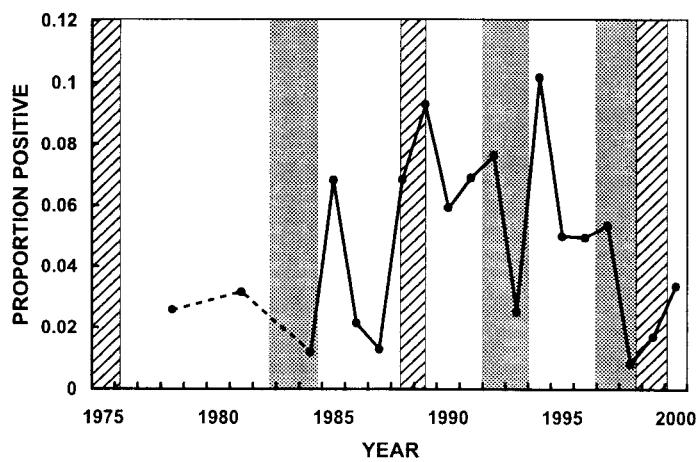
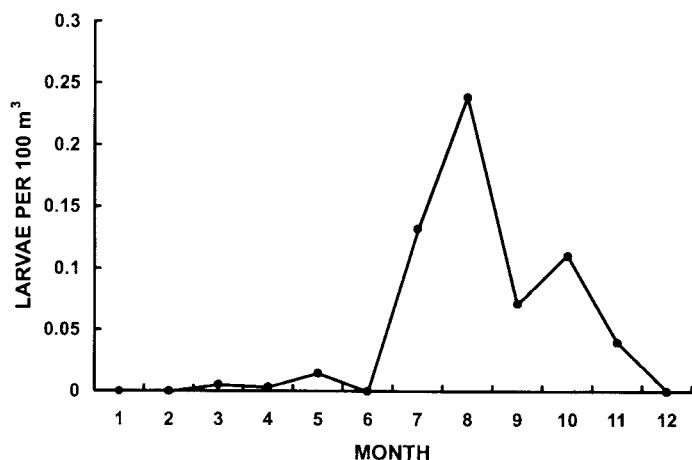
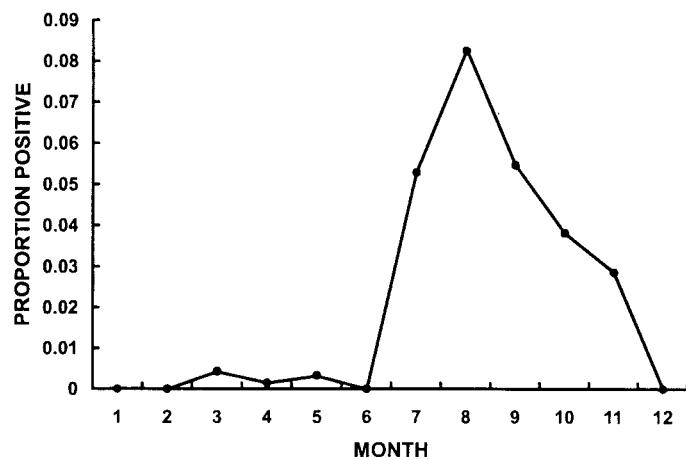
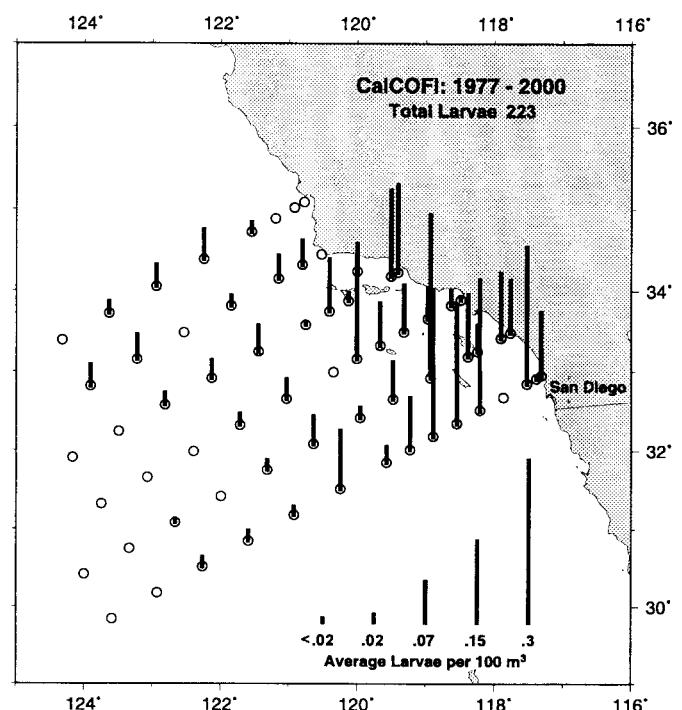
Stenobrachius leucopsarus



Medialuna californiensis



Halfmoon

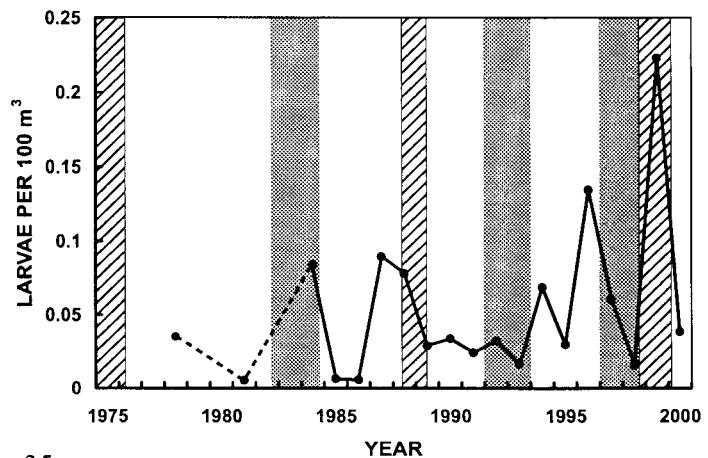
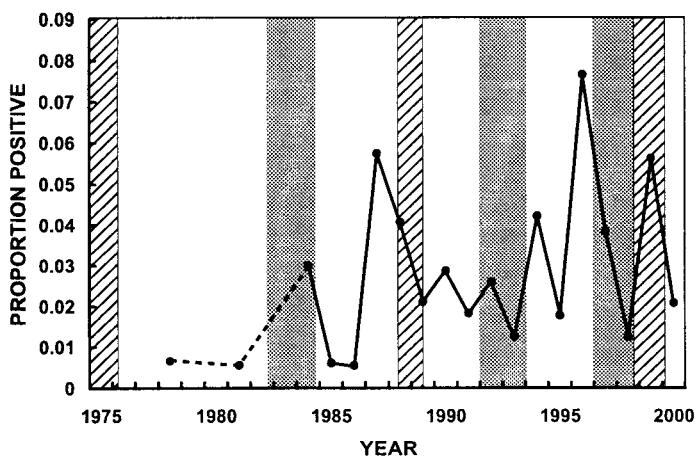
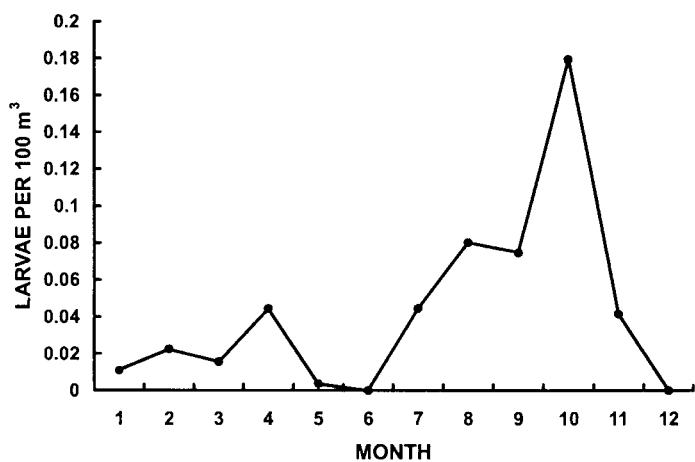
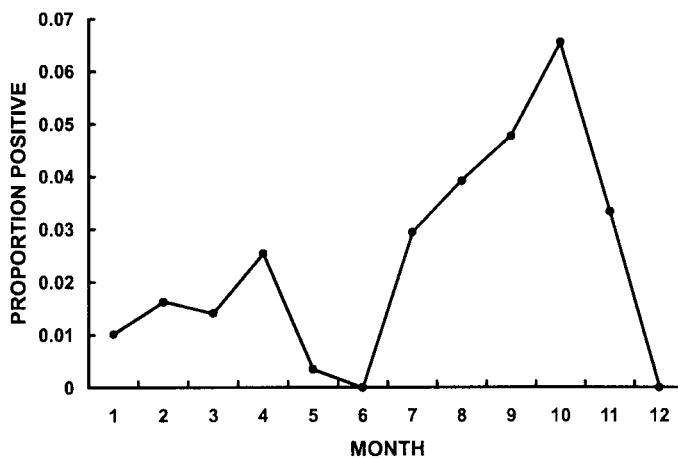
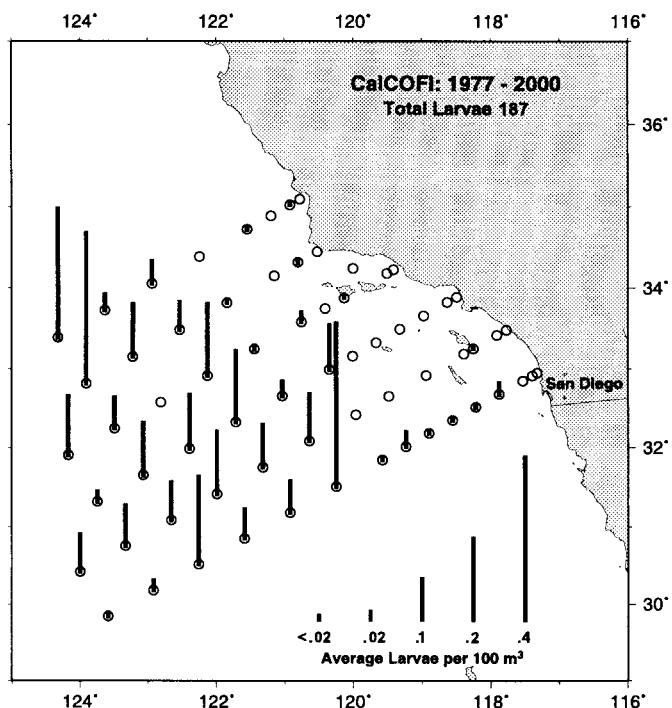
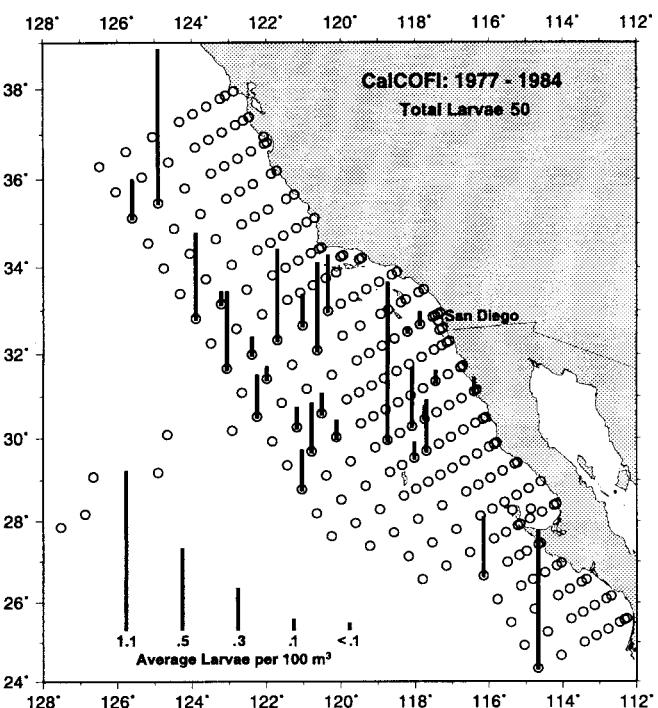


KYPHOSIDAE

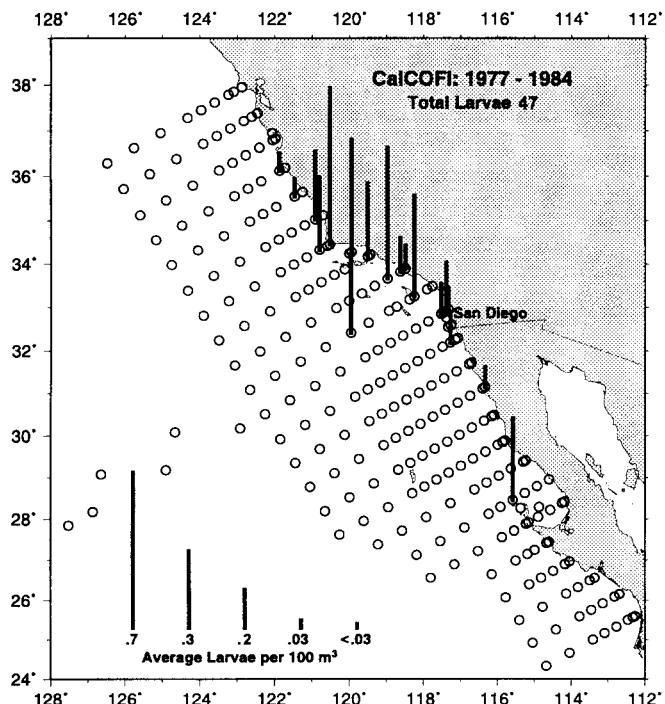
TETRAGONURIDAE

Smalleye squaretail

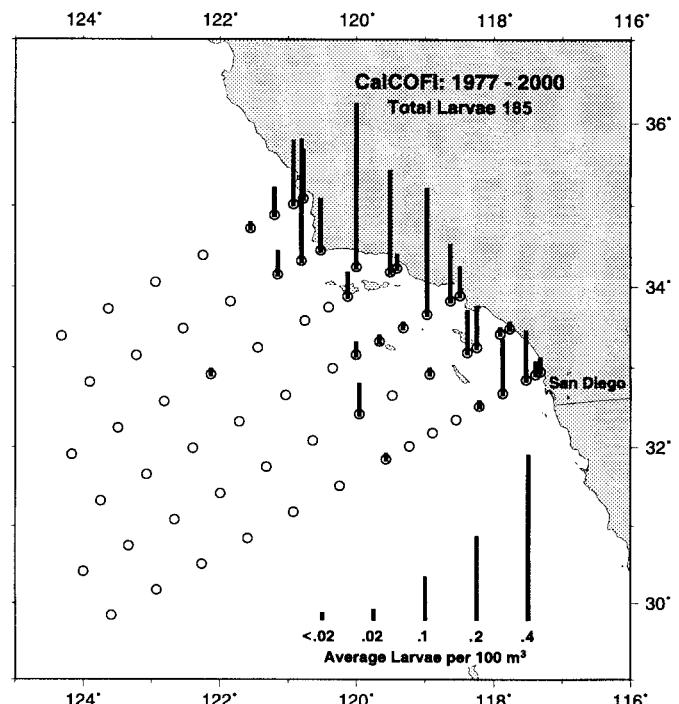
Tetragonurus cuvieri



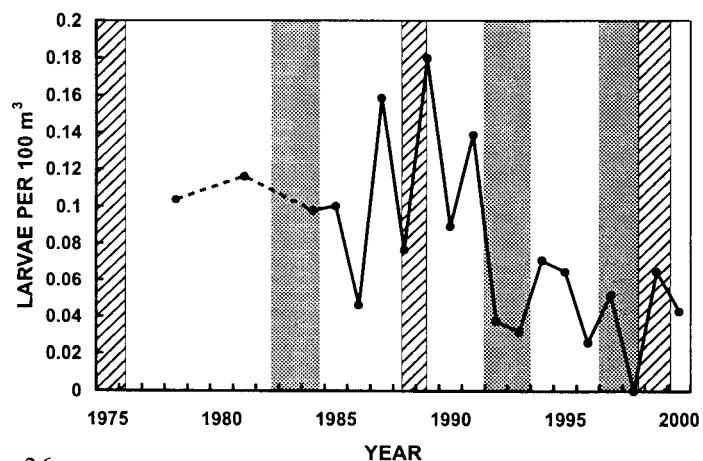
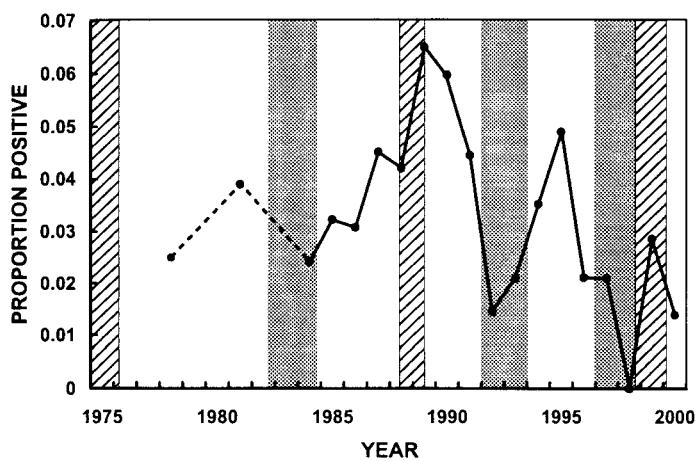
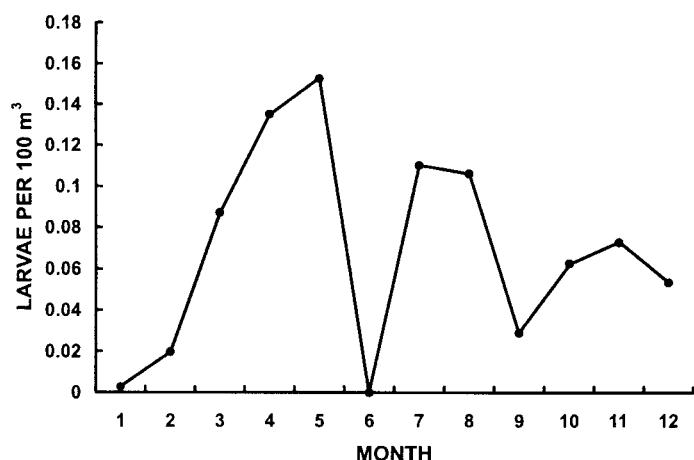
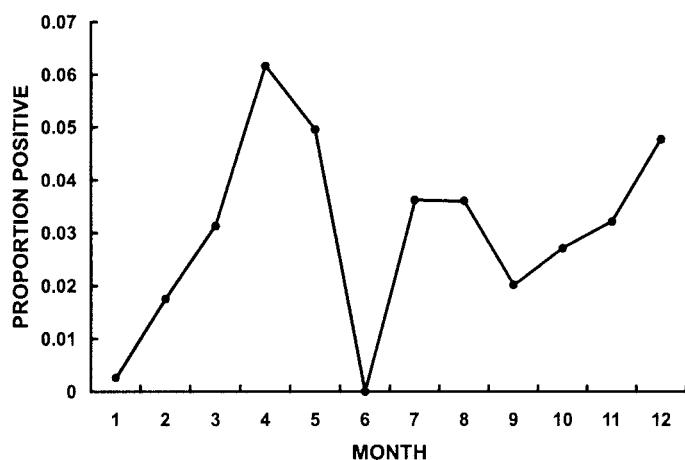
Pleuronichthys coenosus



C-O turbot



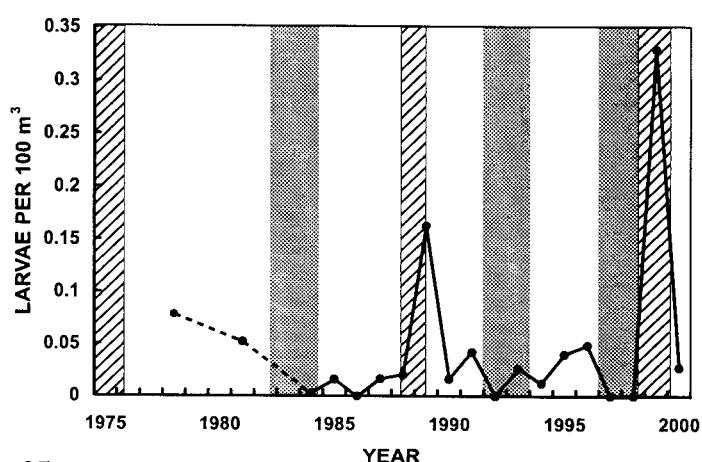
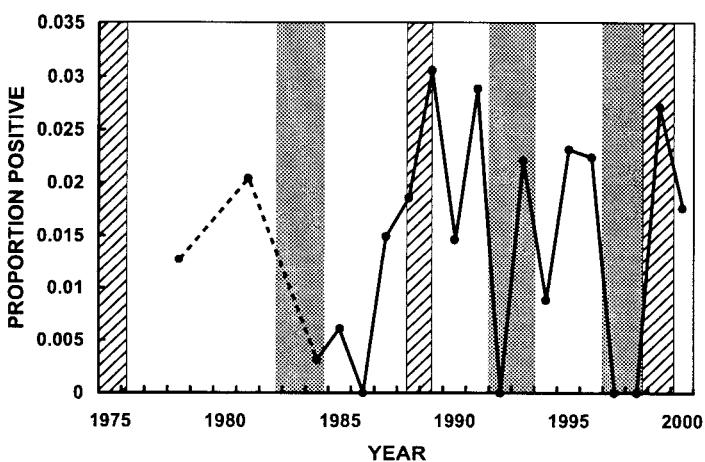
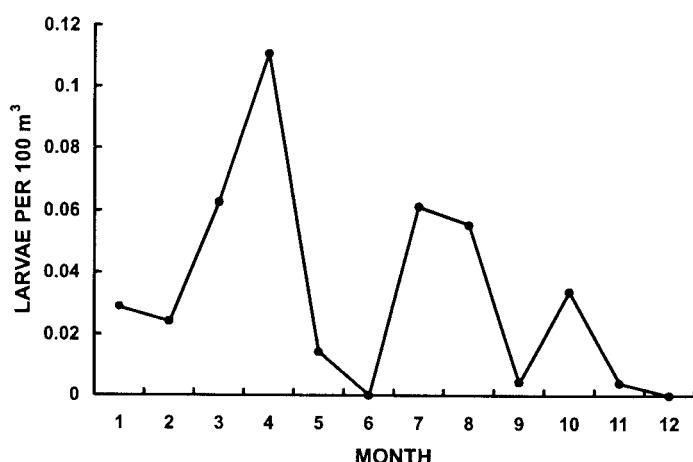
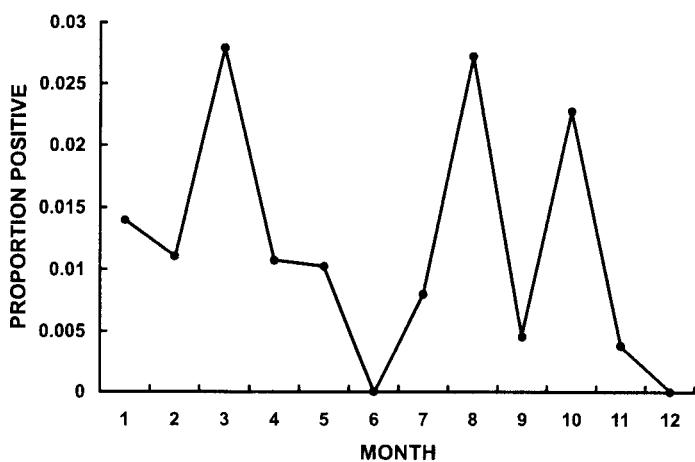
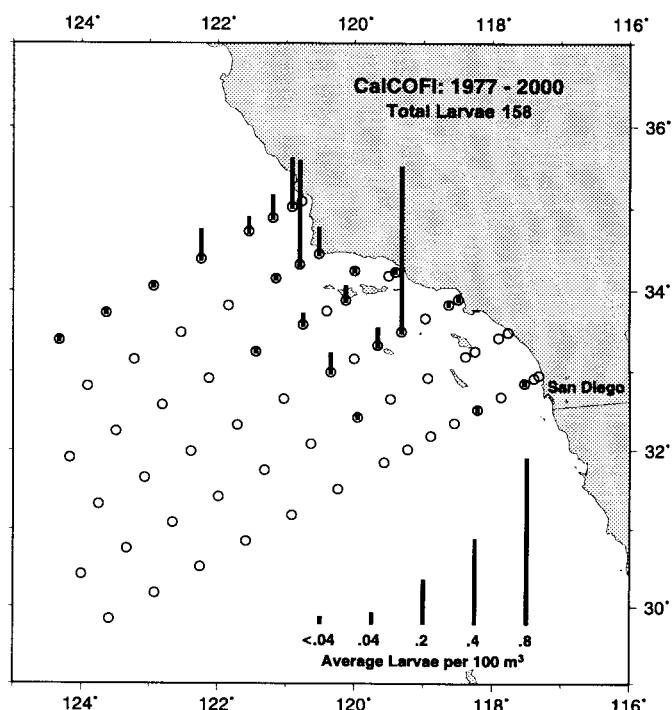
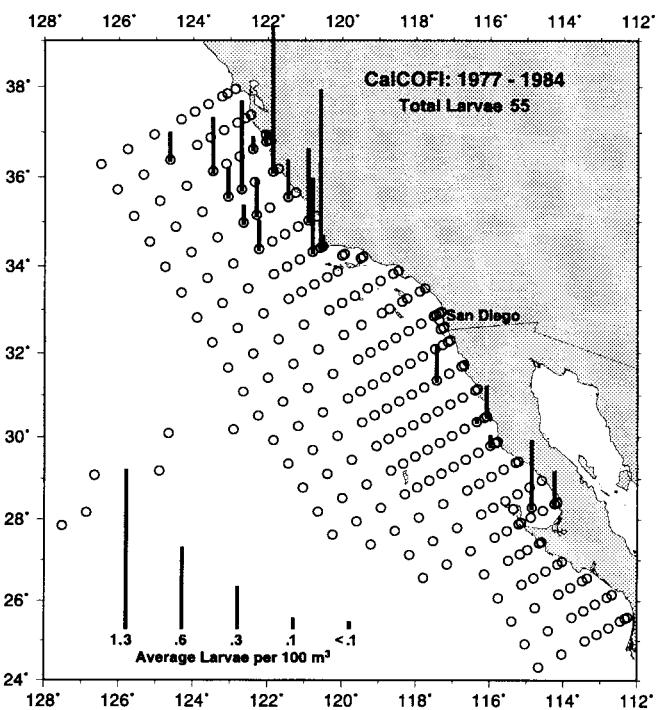
PLEURONECTIDAE



PARALICHTHYIDAE

Speckled sanddab

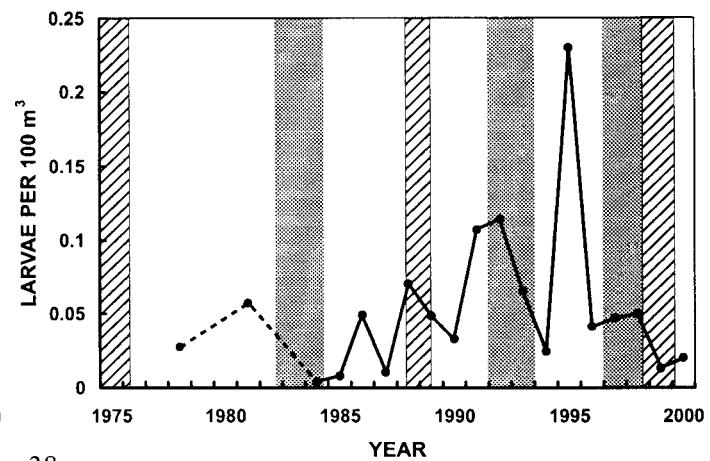
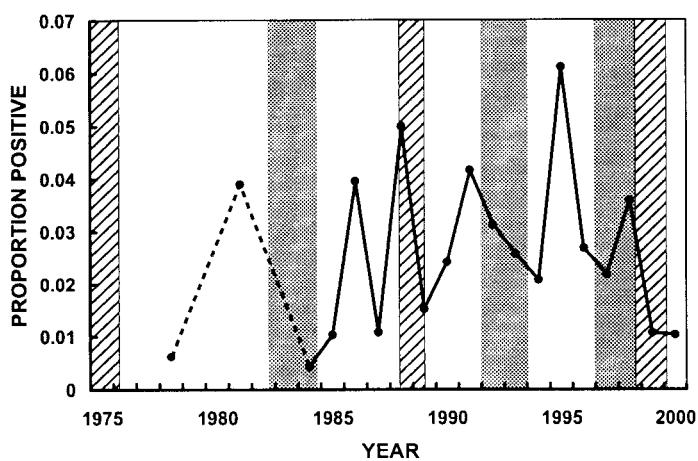
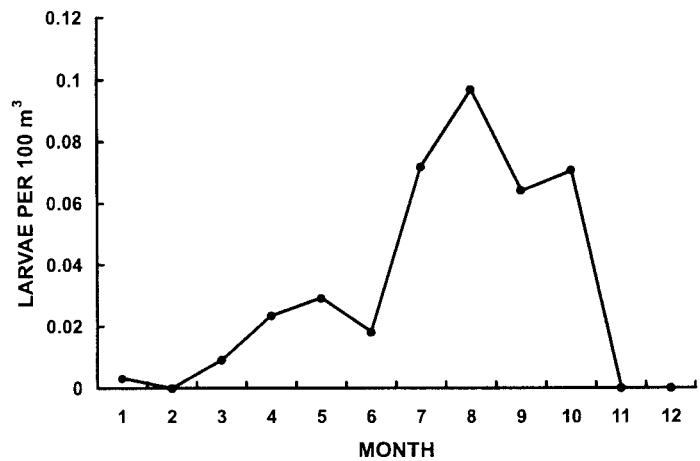
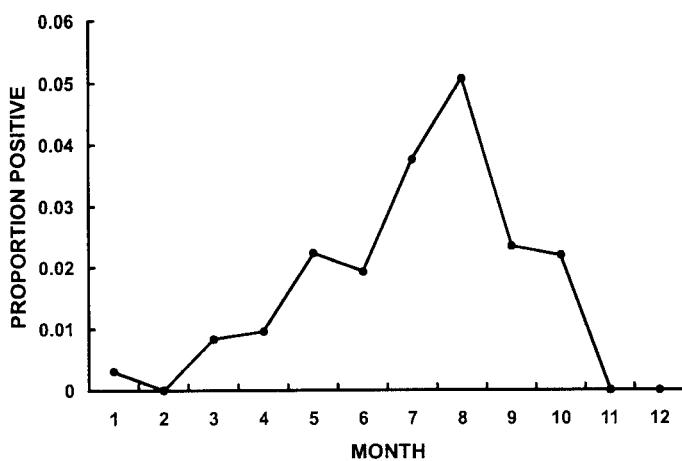
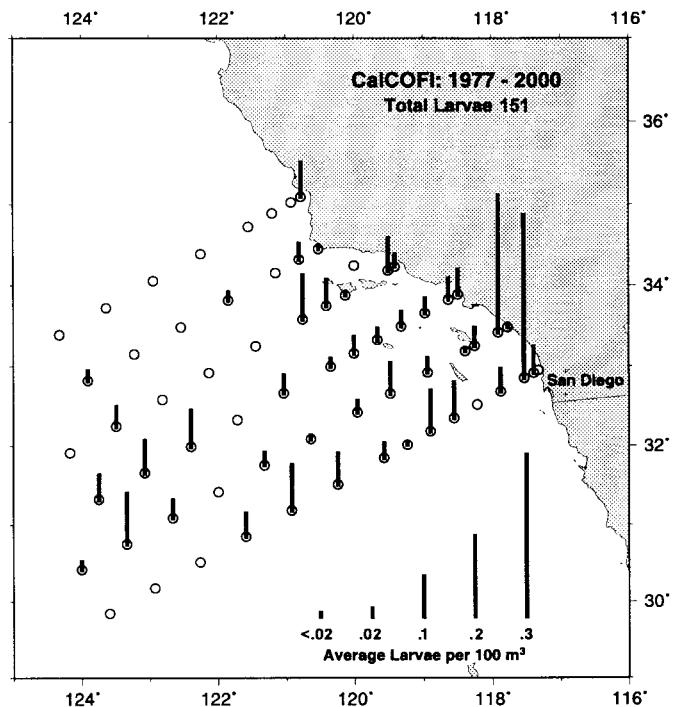
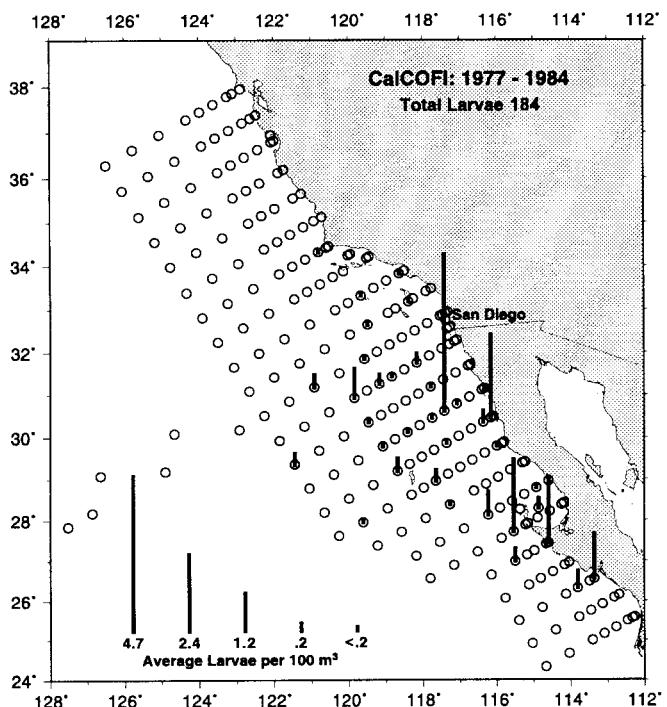
Citharichthys stigmaeus



Triphoturus mexicanus

Mexican lampfish

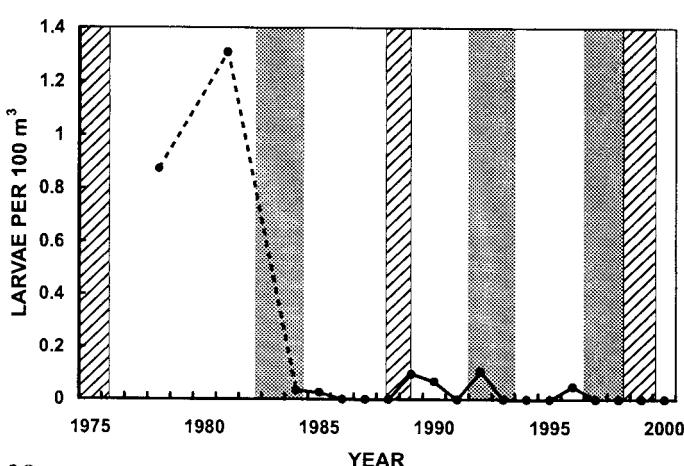
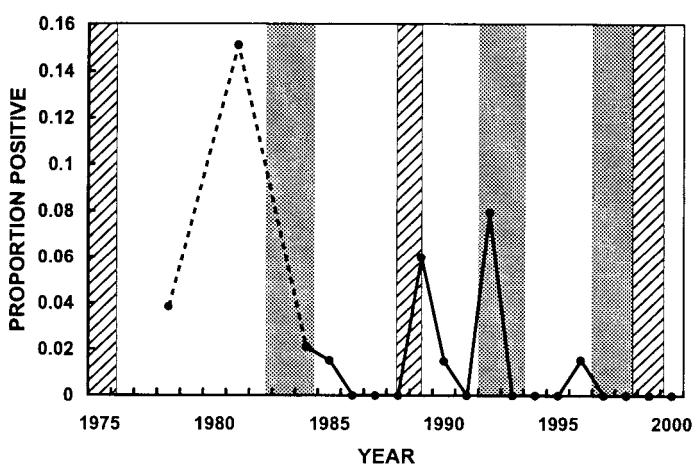
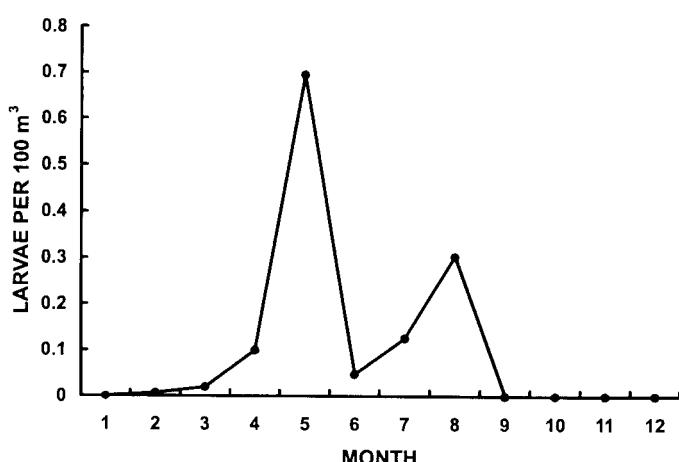
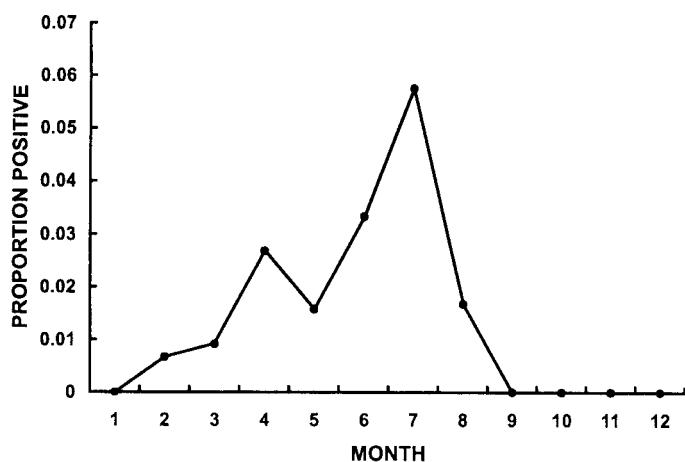
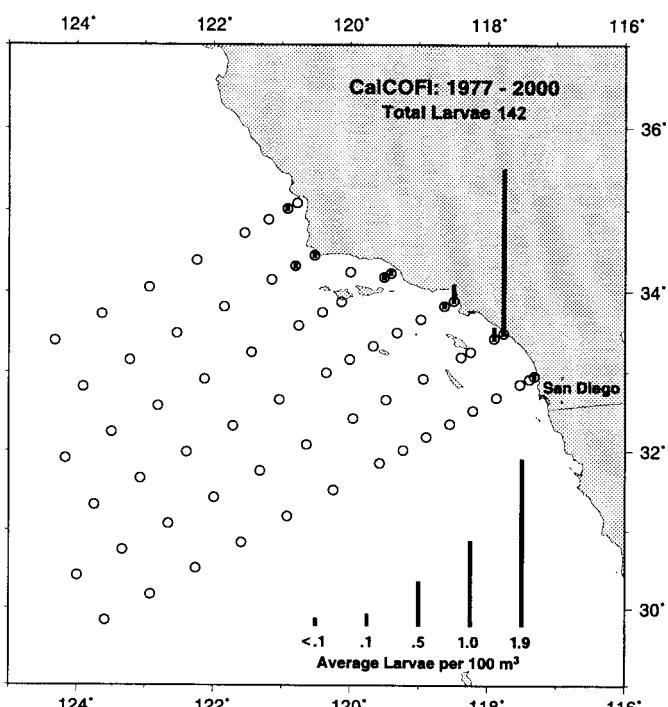
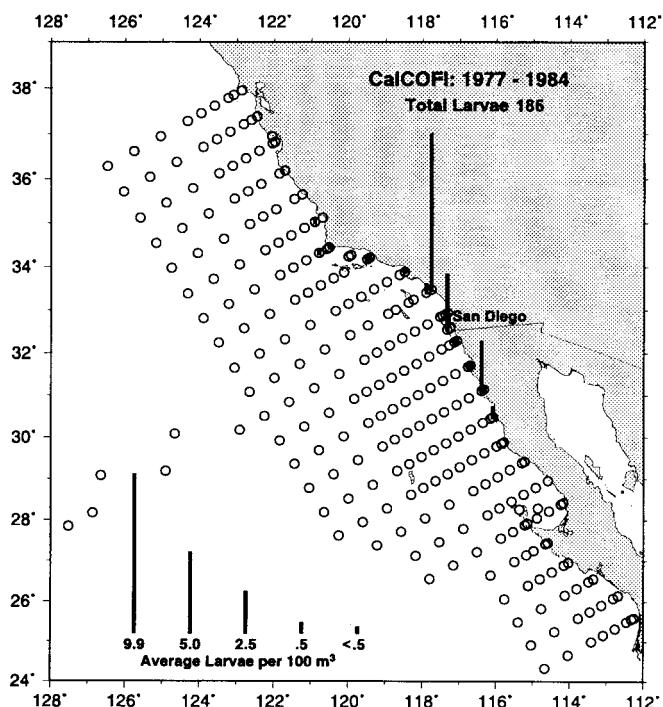
MYCTOPHIDAE



SCIENIDAE

Queenfish

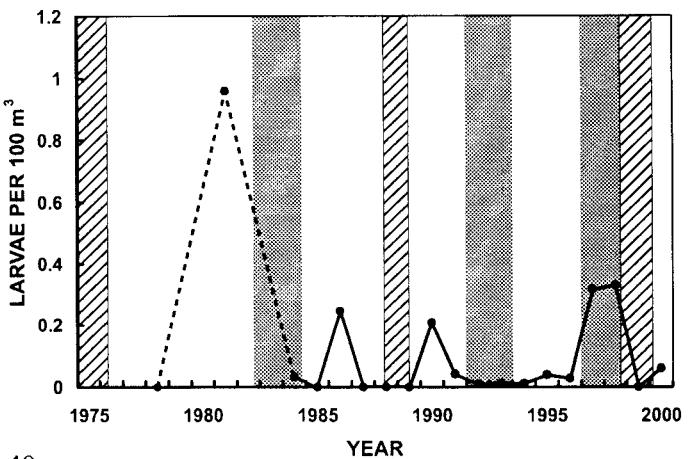
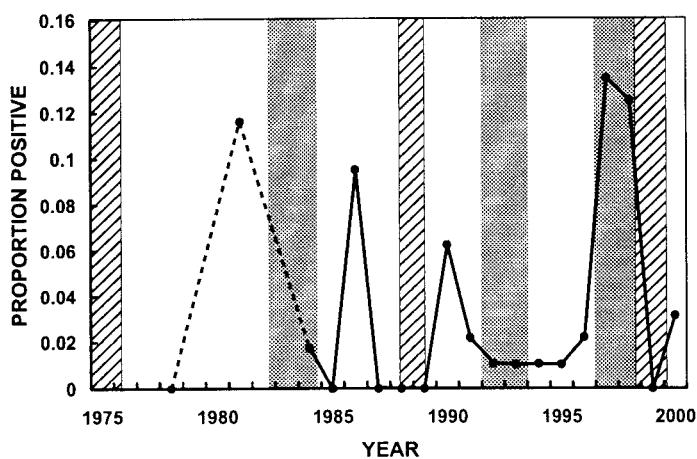
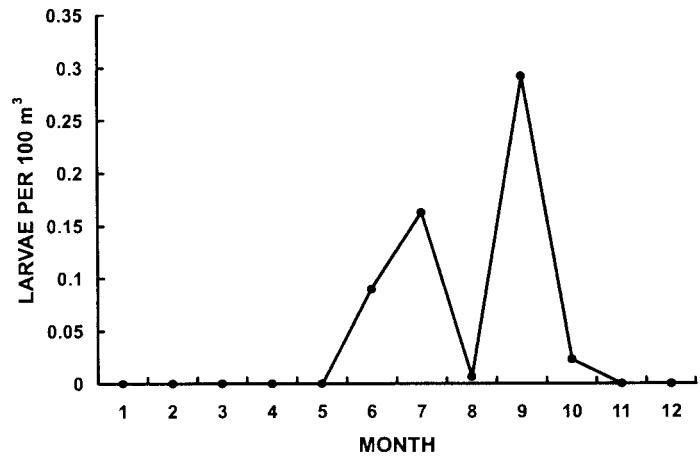
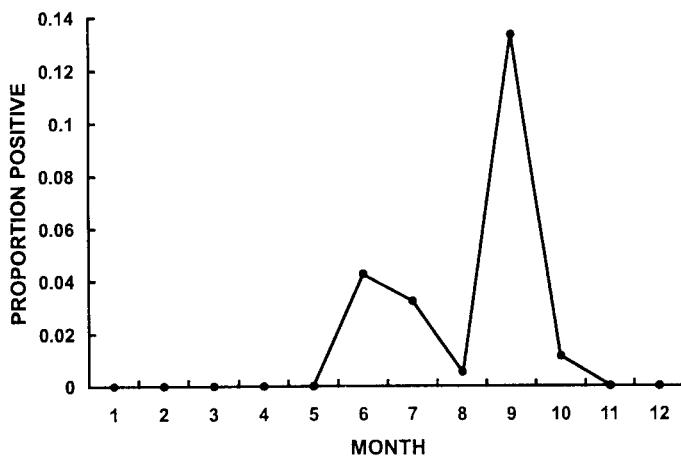
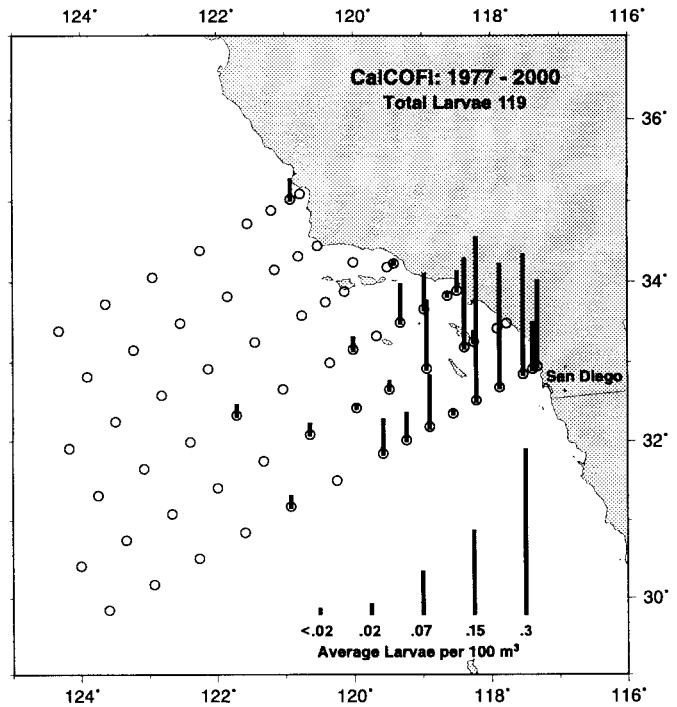
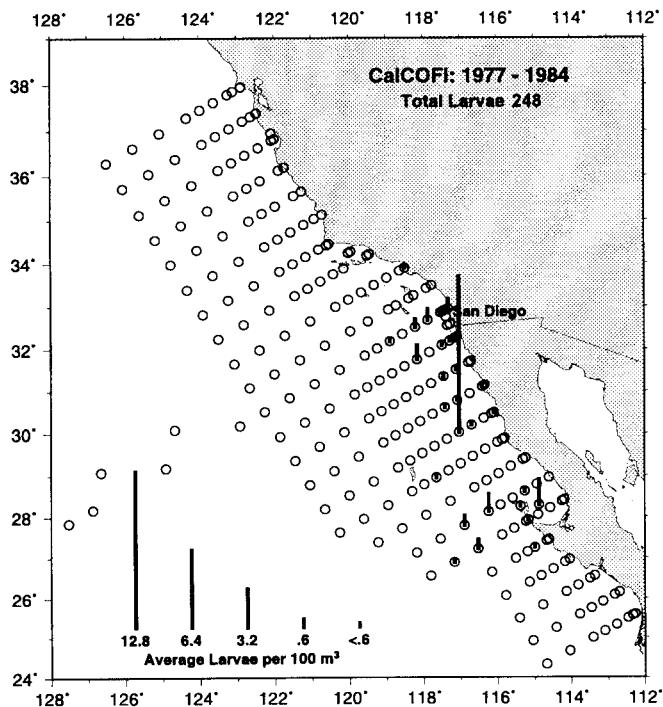
Seriphus politus



Cheilopogon heterurus

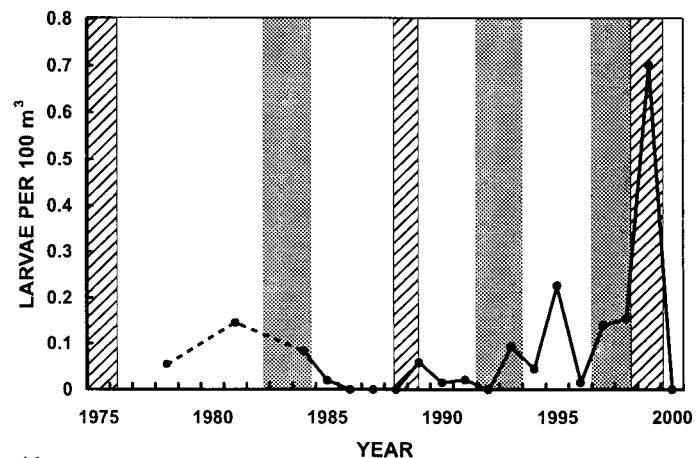
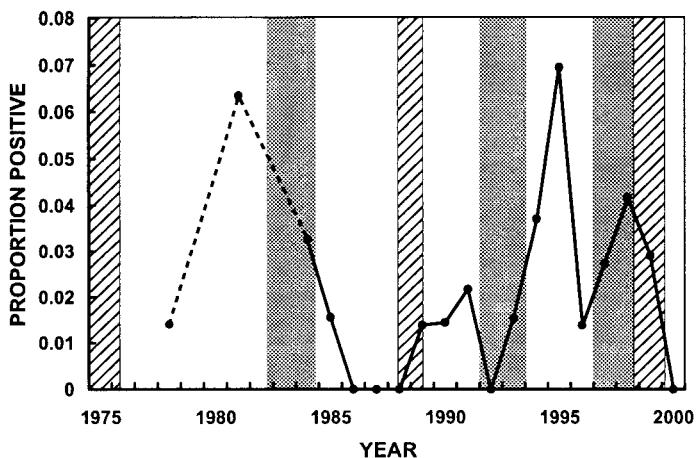
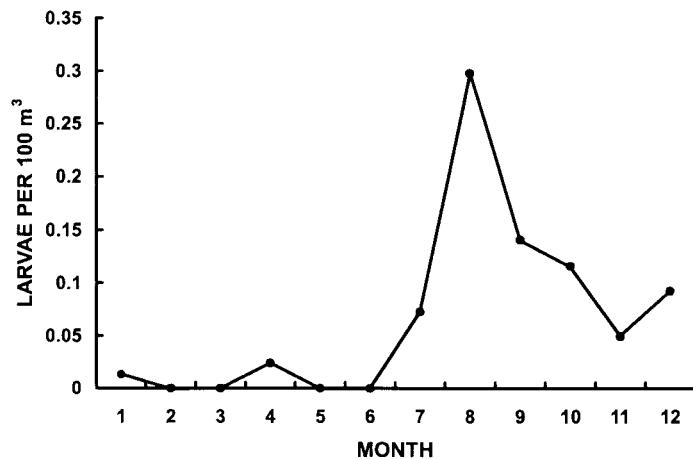
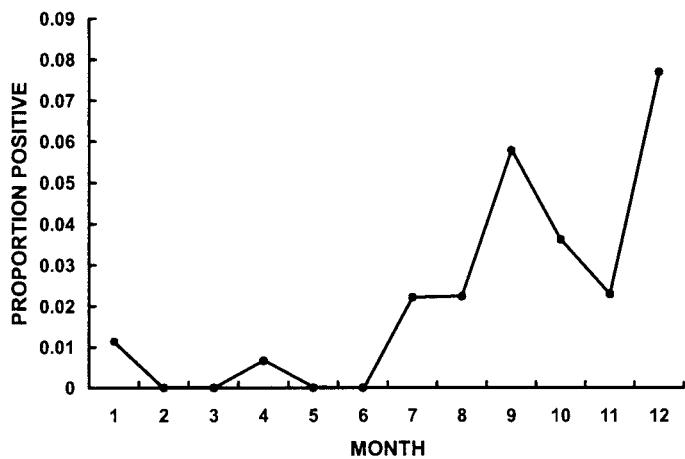
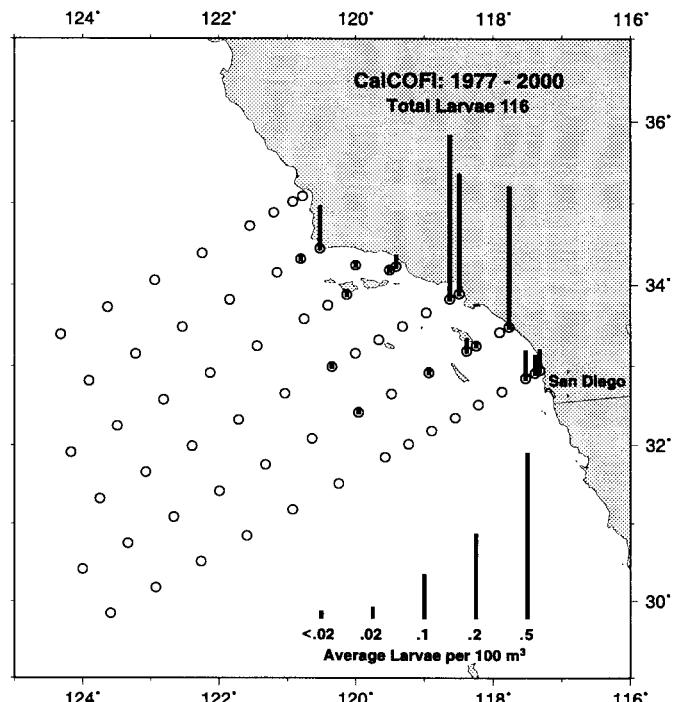
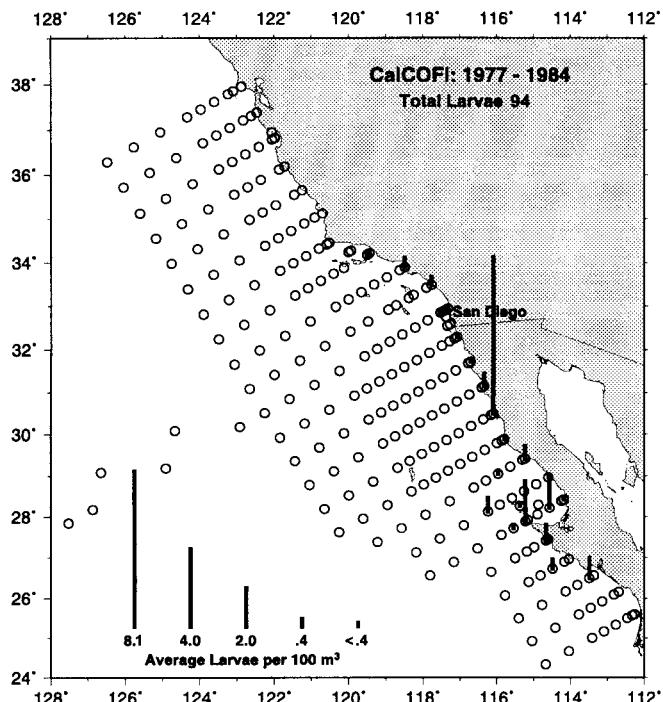
Blotchwing flyingfish

EXOCOETIDAE

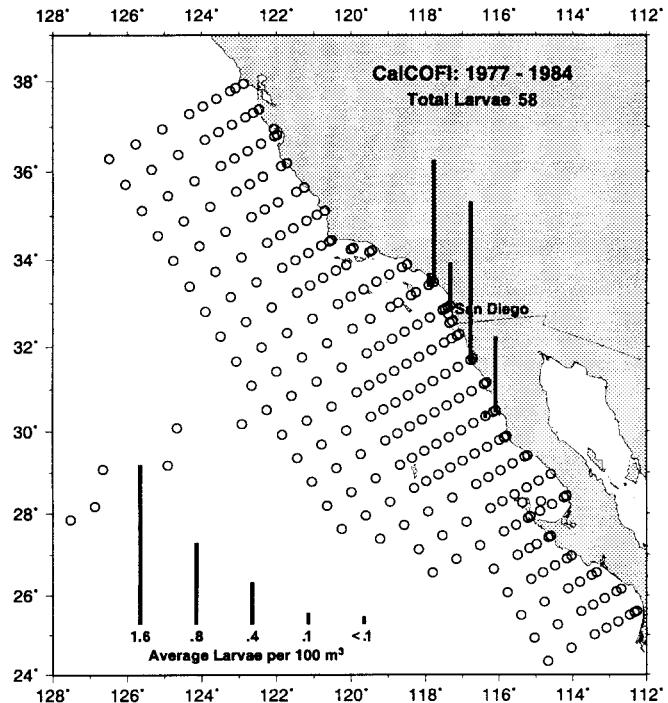


BLENNIIDAE

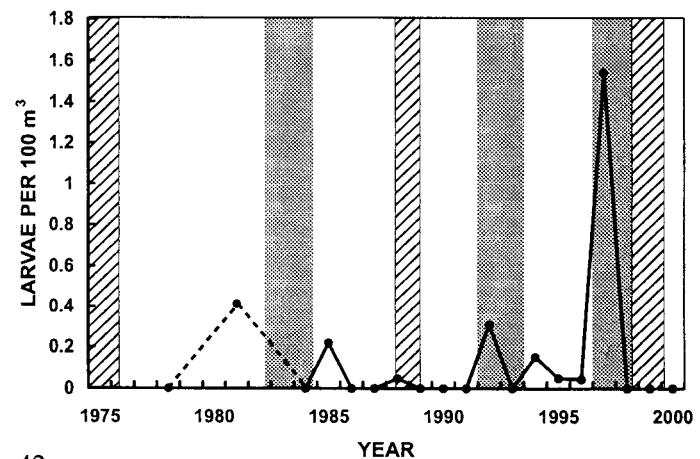
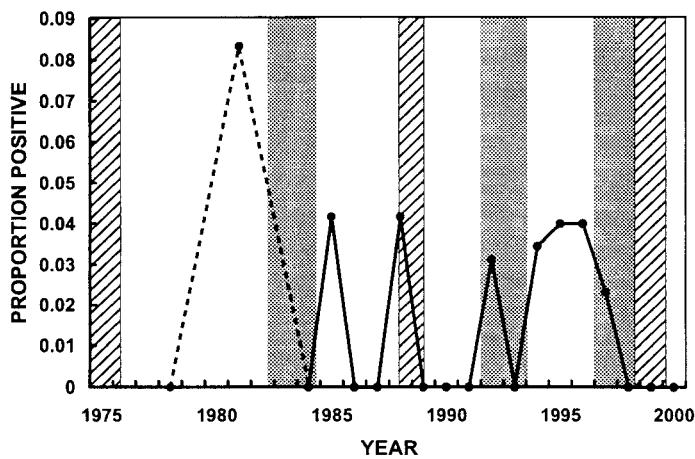
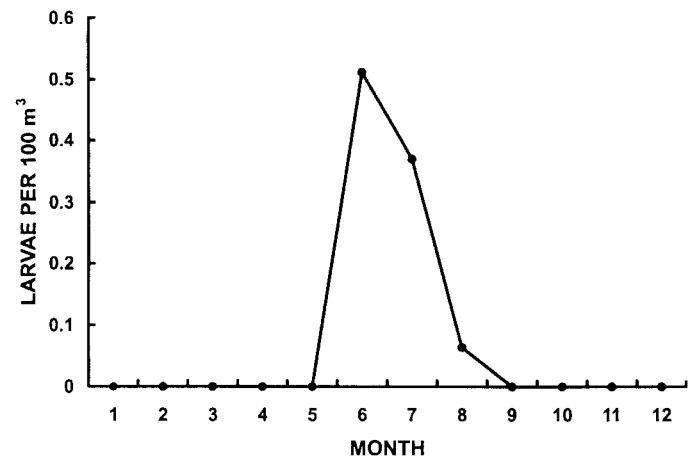
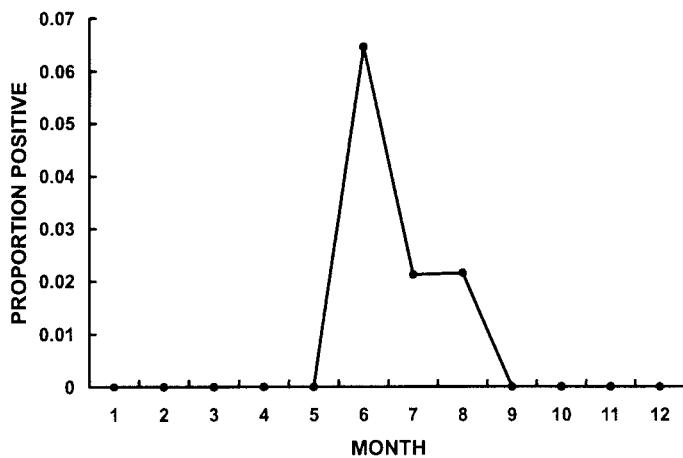
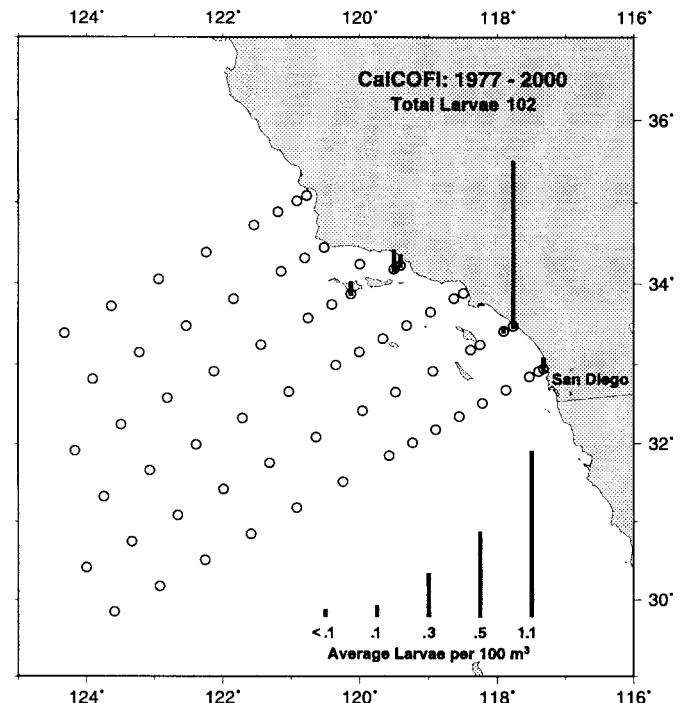
Bay blenny

Hypsoblennius gentilis

Hypsypops rubicundus



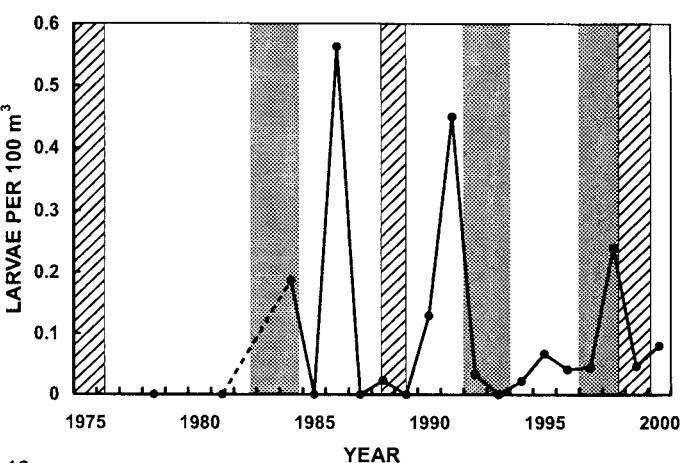
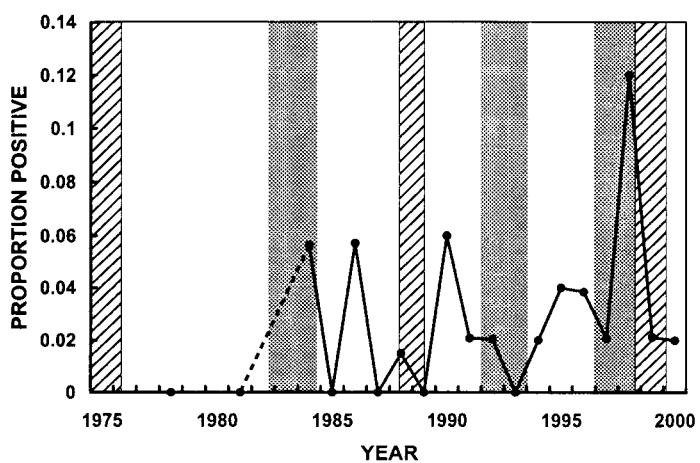
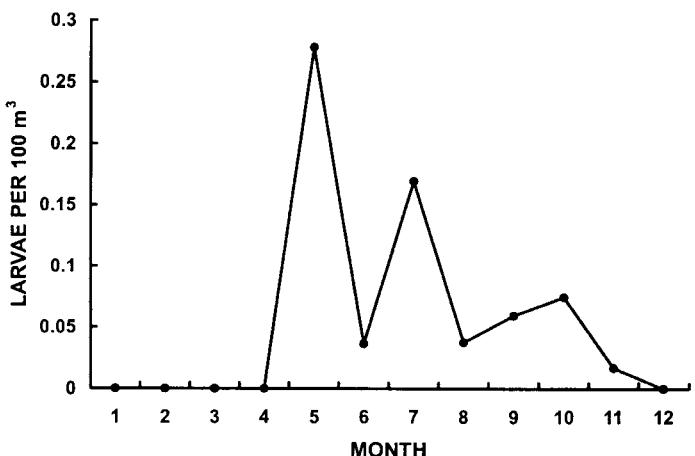
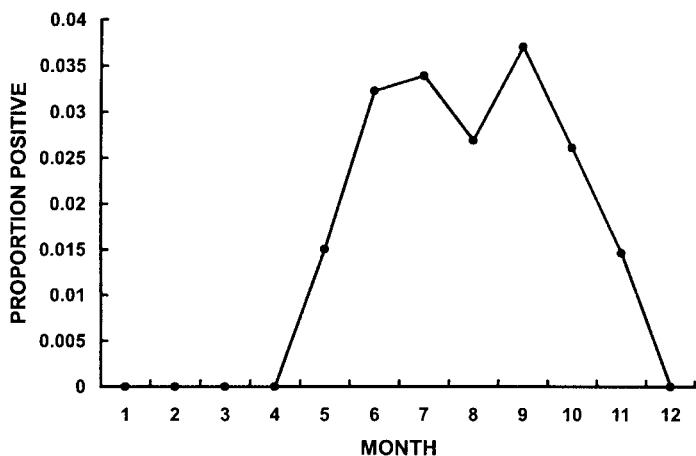
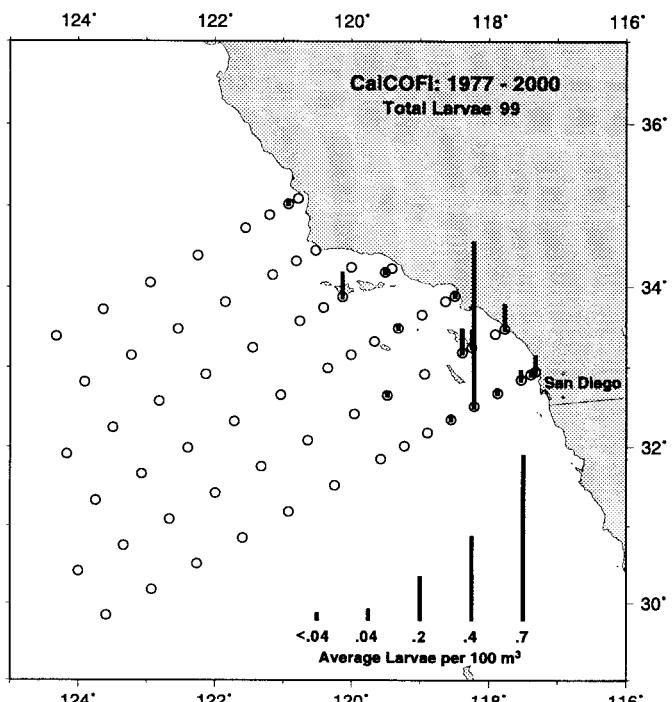
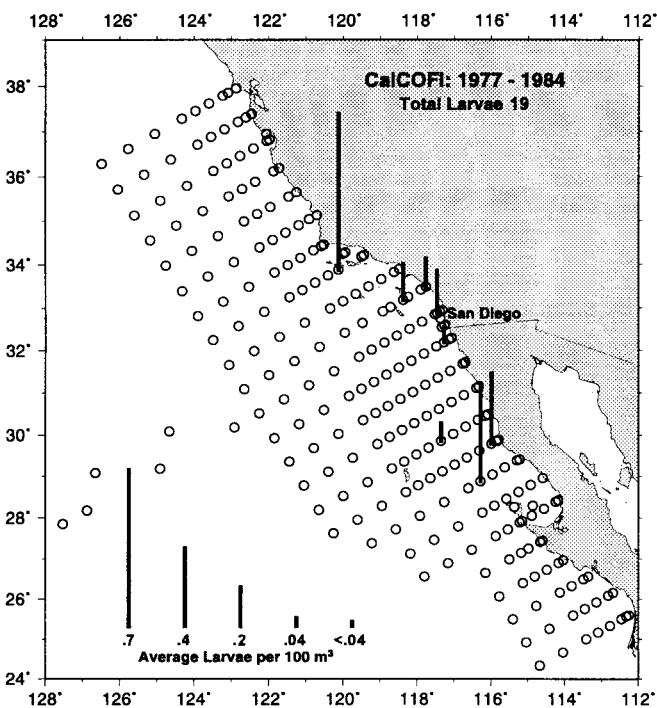
Garibaldi



EXOCOETIDAE

California flyingfish

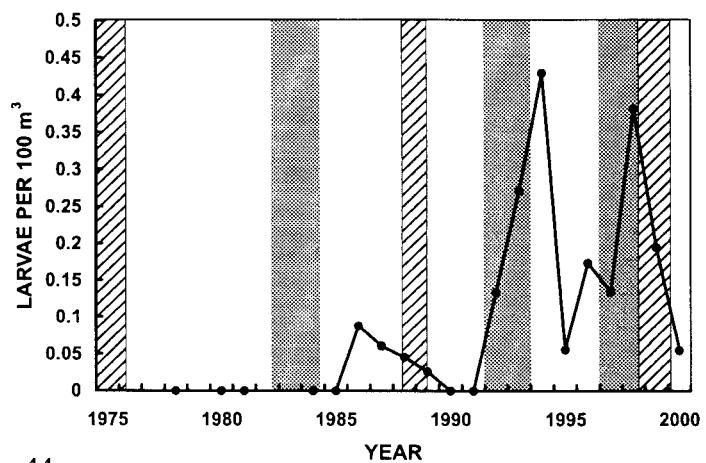
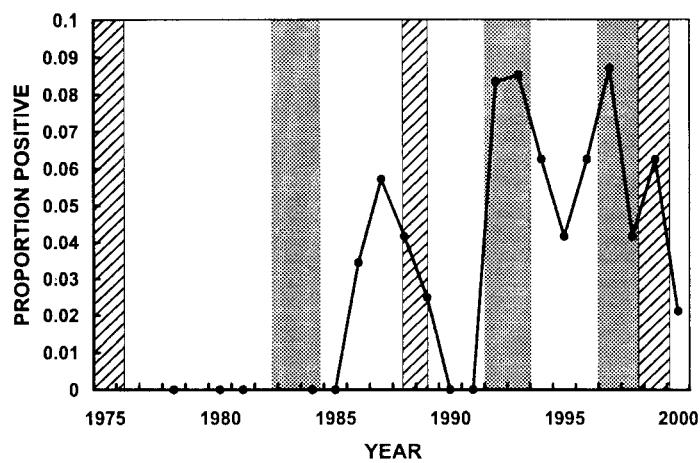
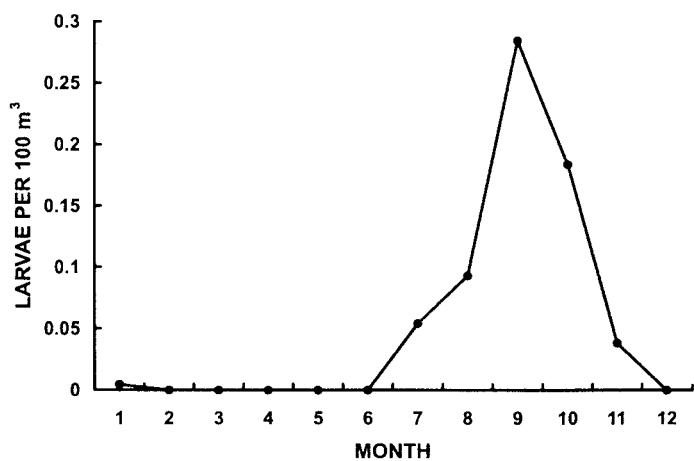
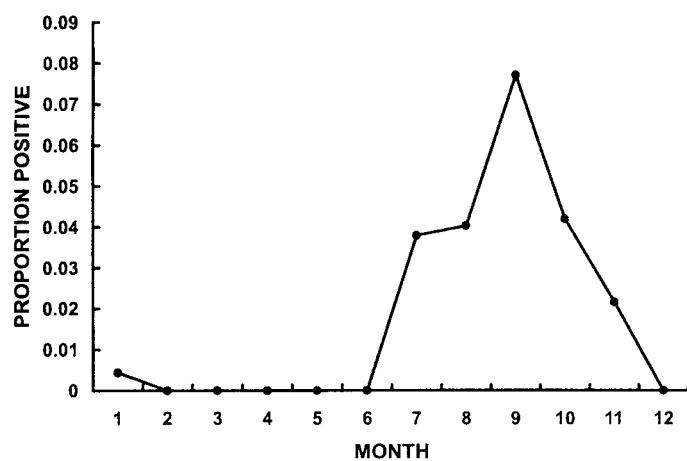
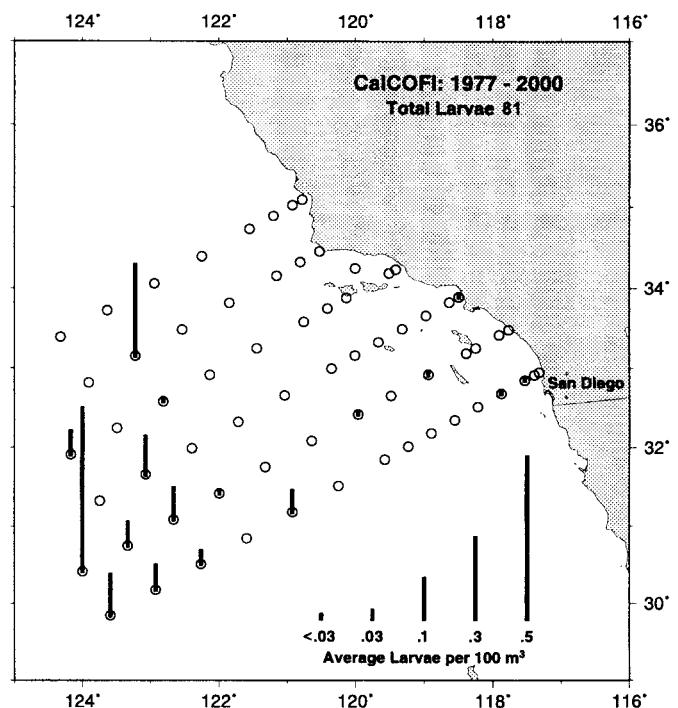
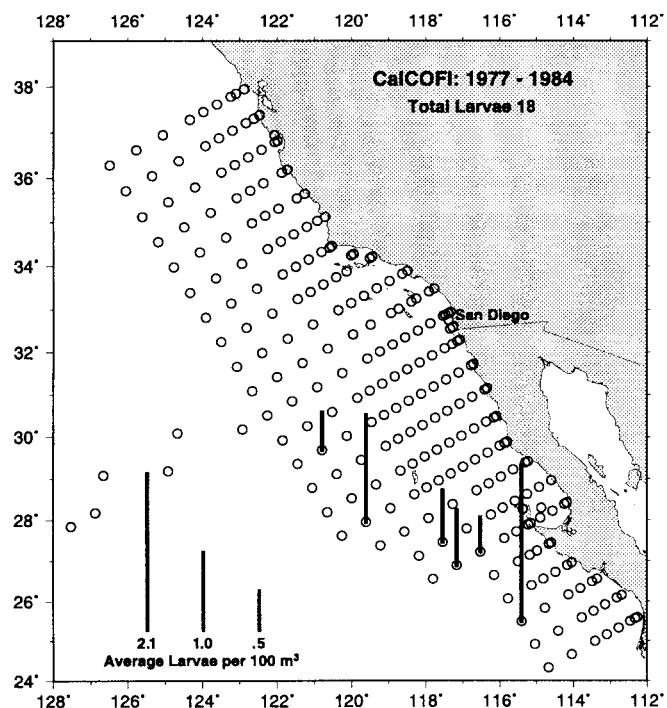
Cheilopogon pinnatibarbus



*Lampadena urophao*s

Sunbeam lampfish

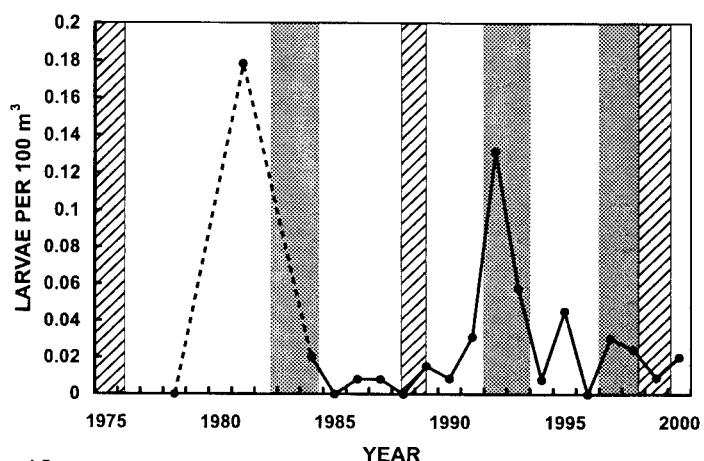
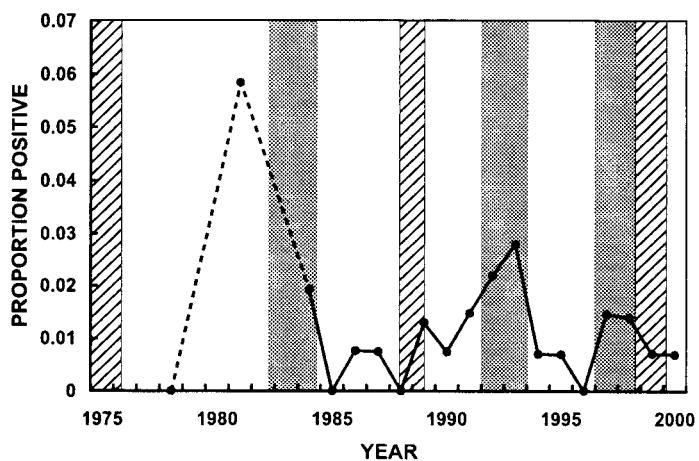
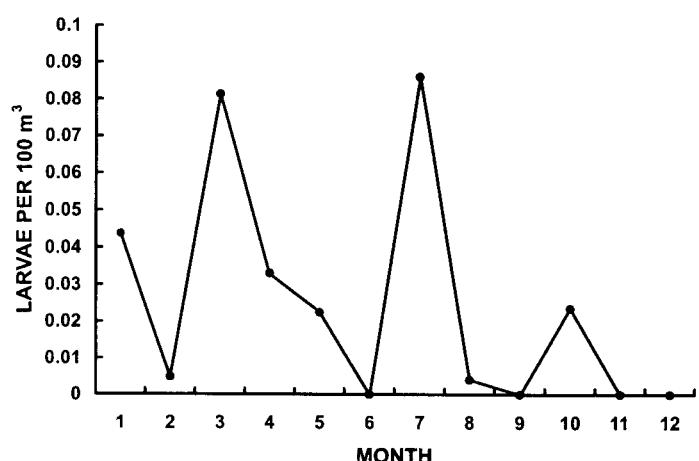
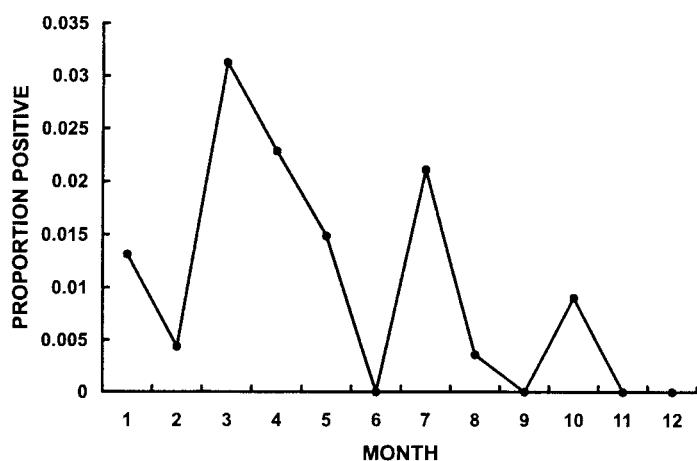
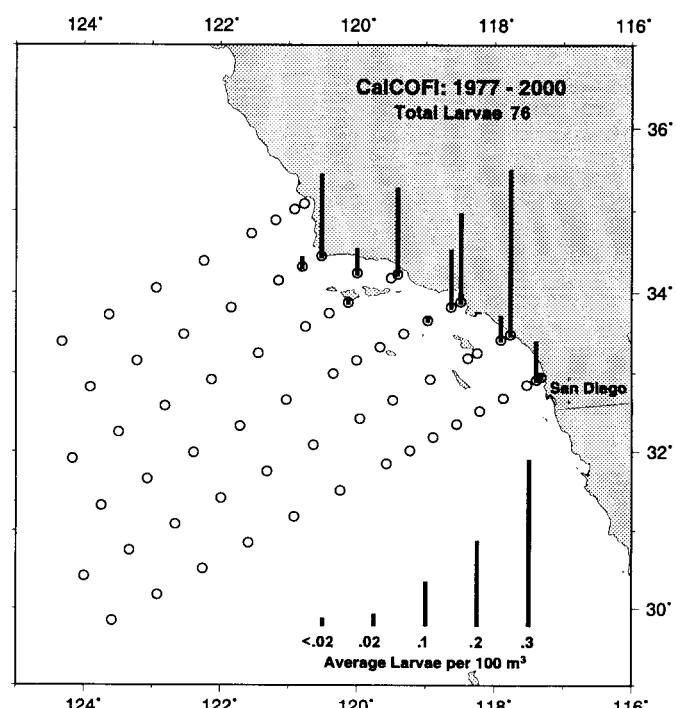
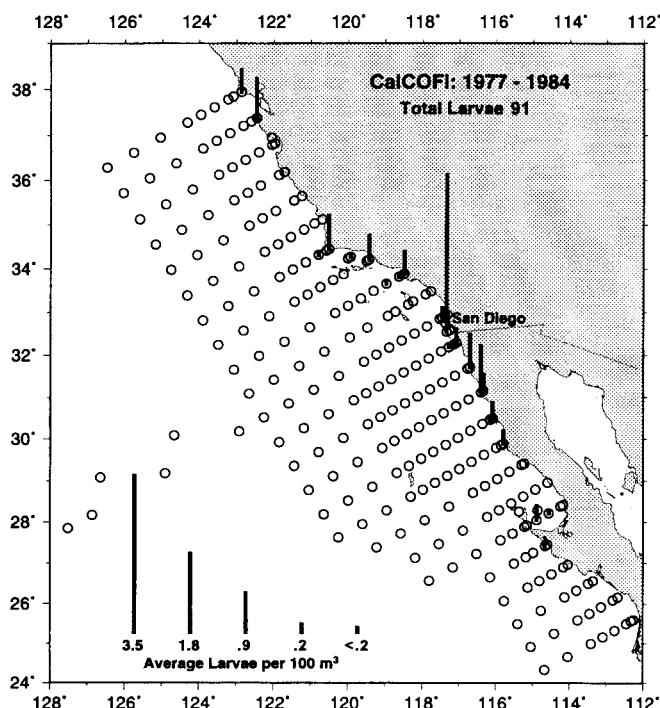
MYCTOPHIDAE



PARALICHTHYIDAE

California halibut

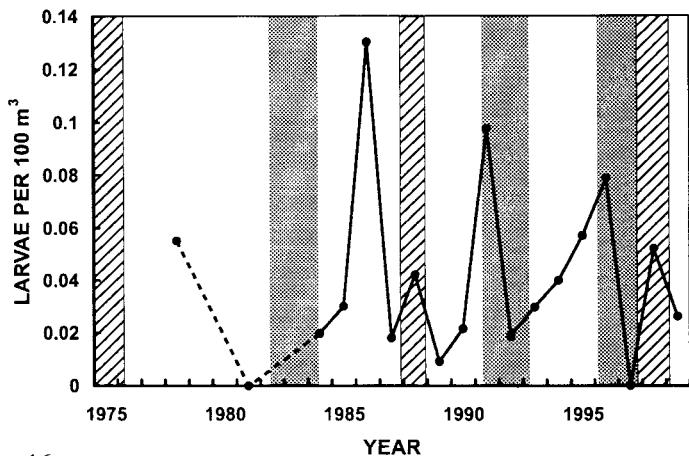
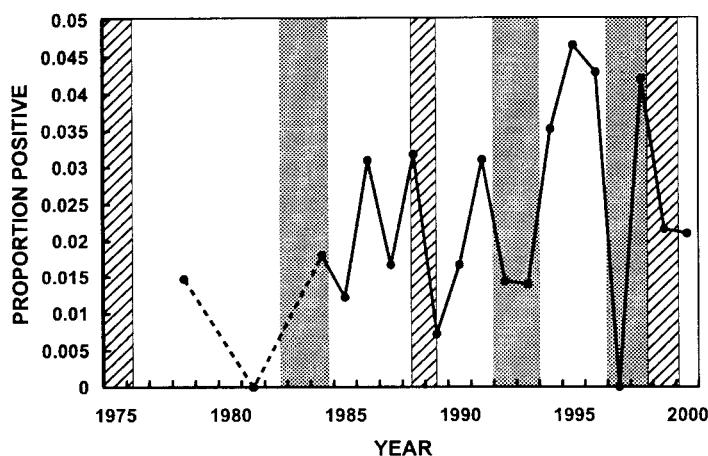
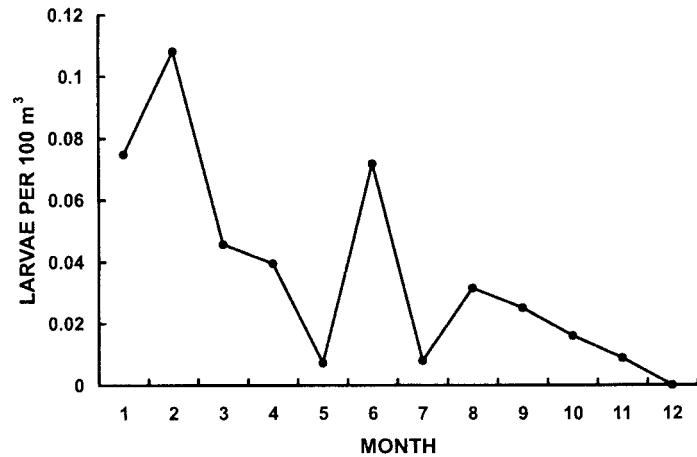
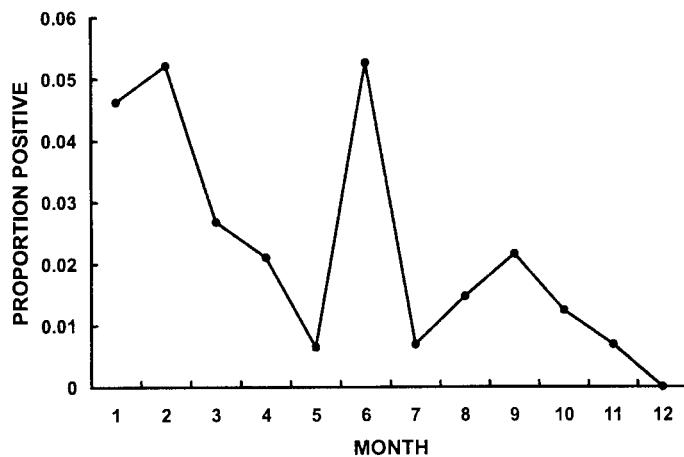
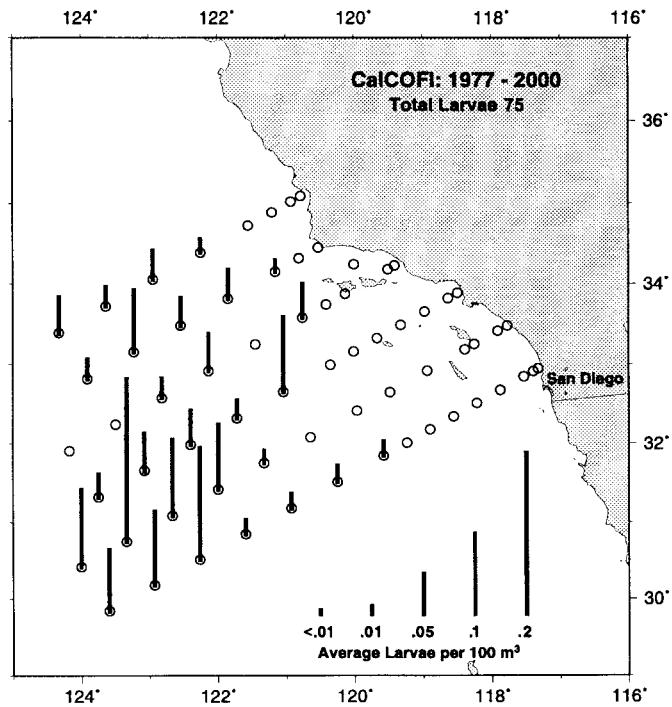
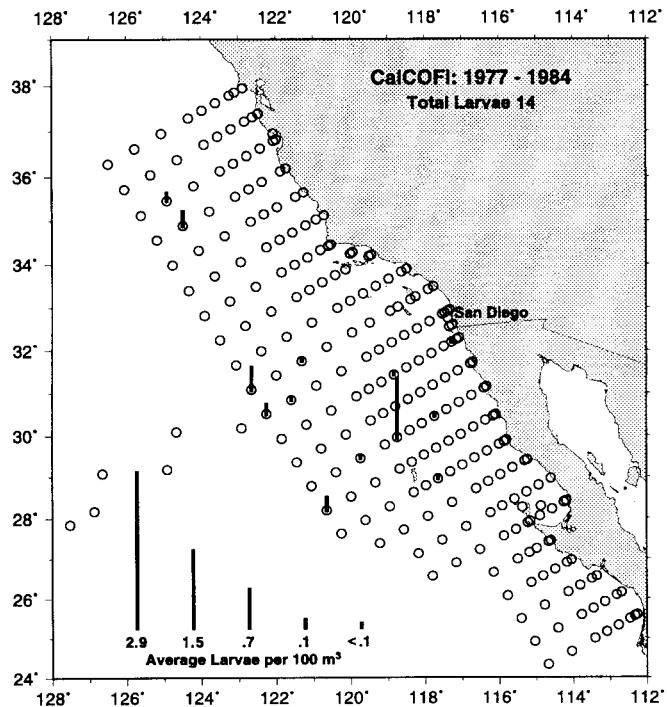
Paralichthys californicus



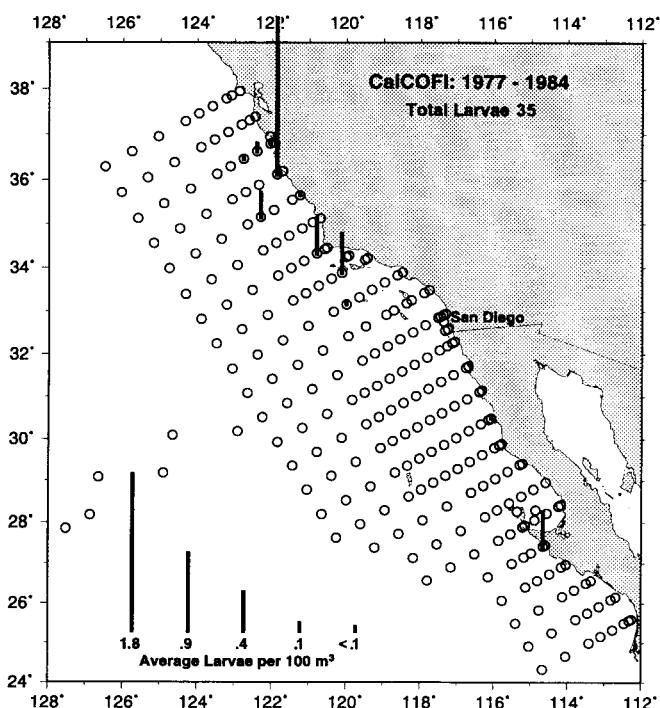
Cyclothona signata

Showy bristlemouth

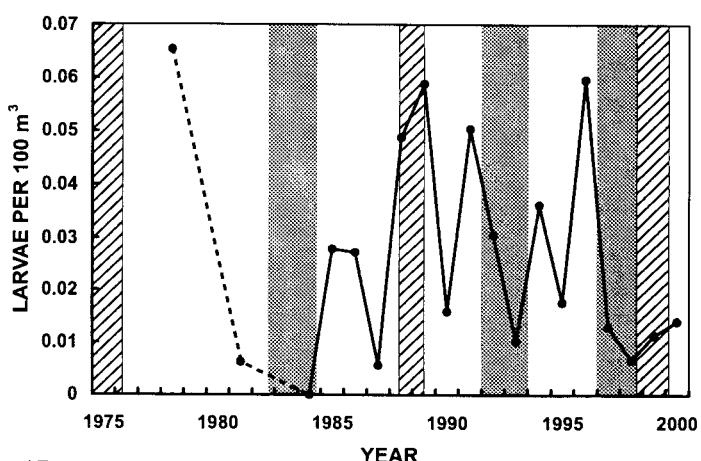
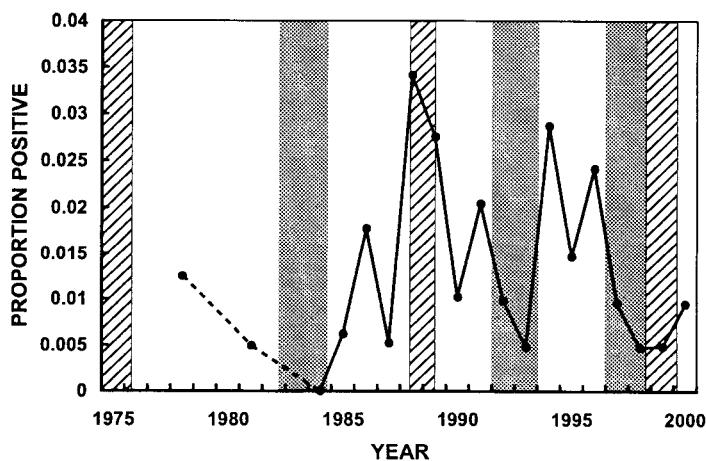
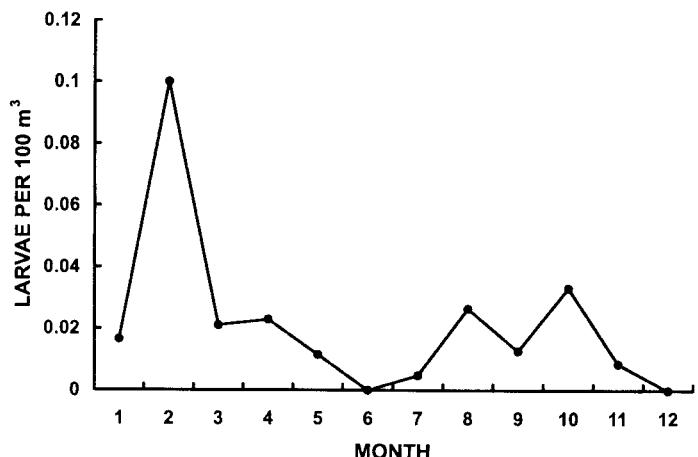
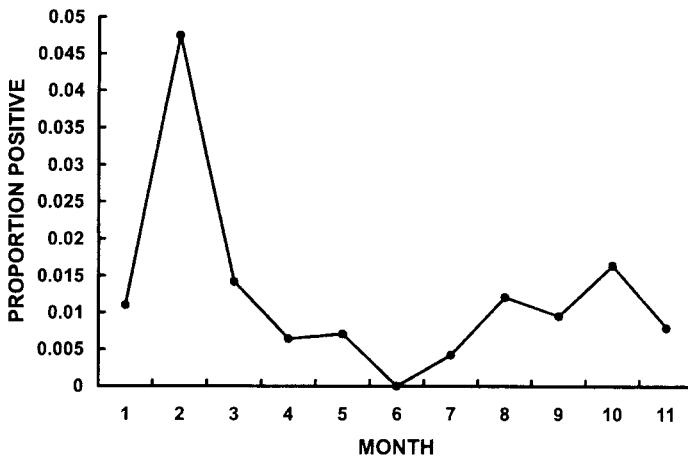
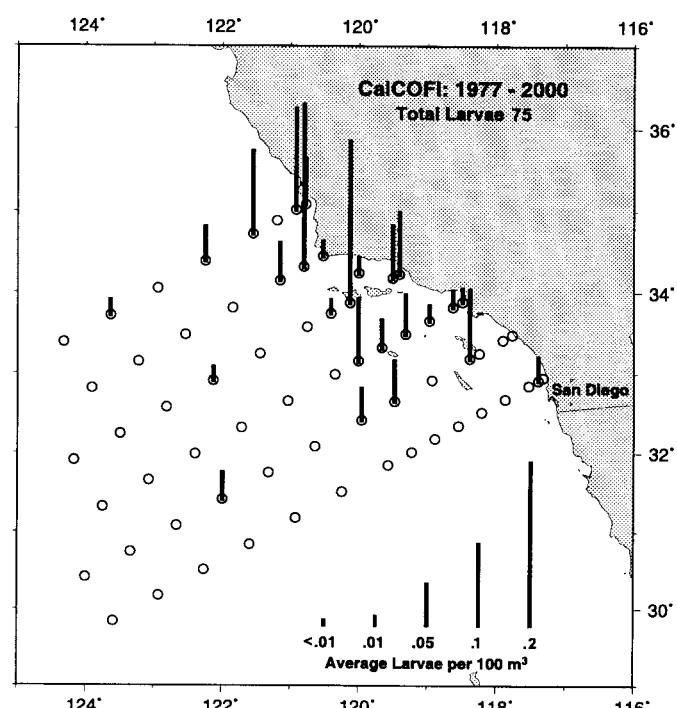
GONOSTOMATIDAE



PARALICHTHYIDAE



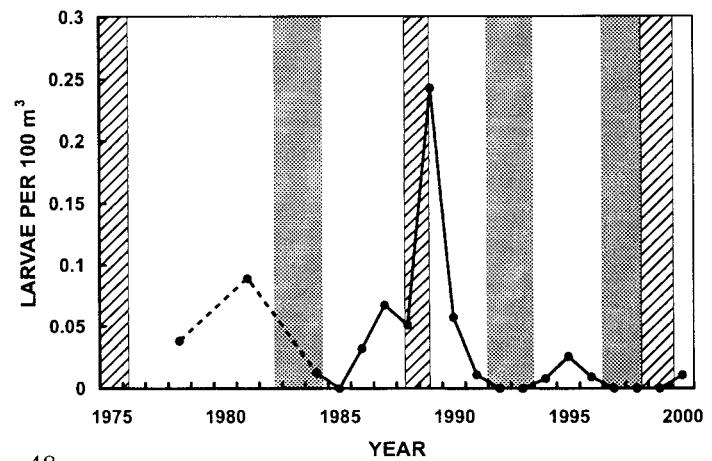
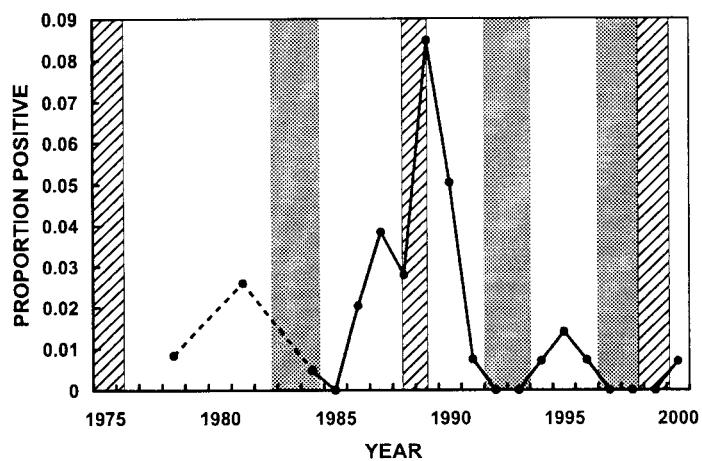
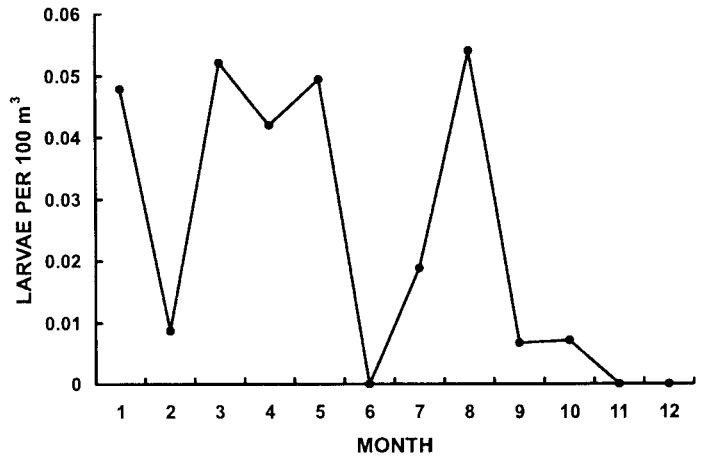
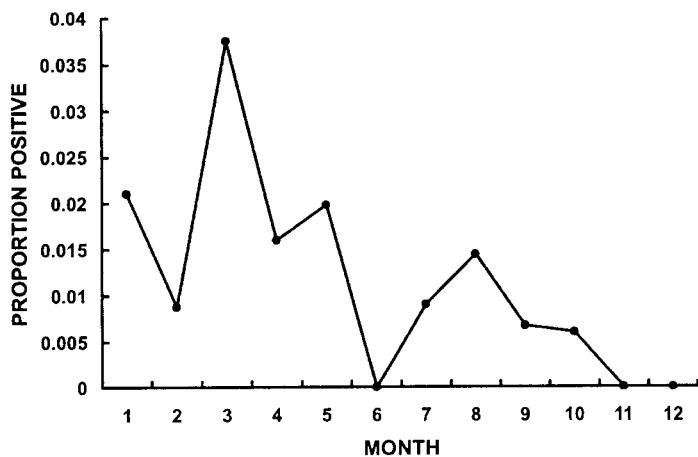
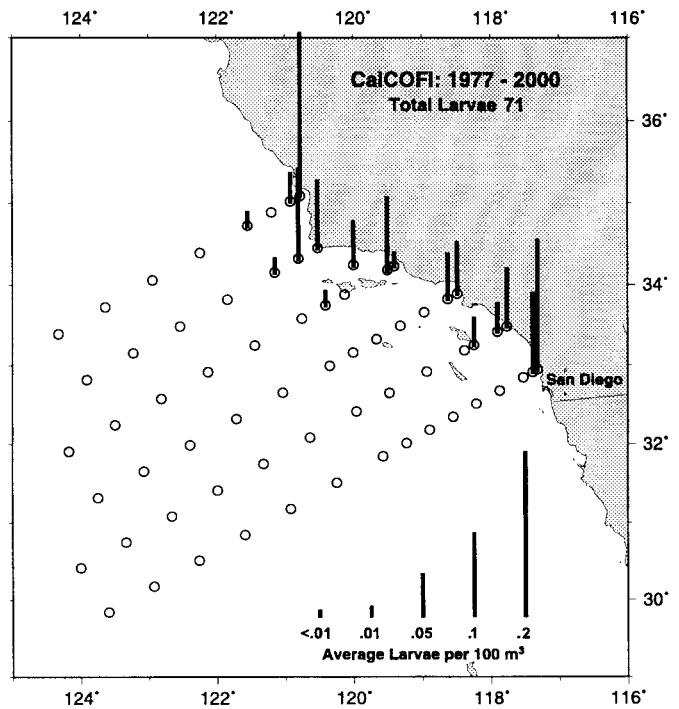
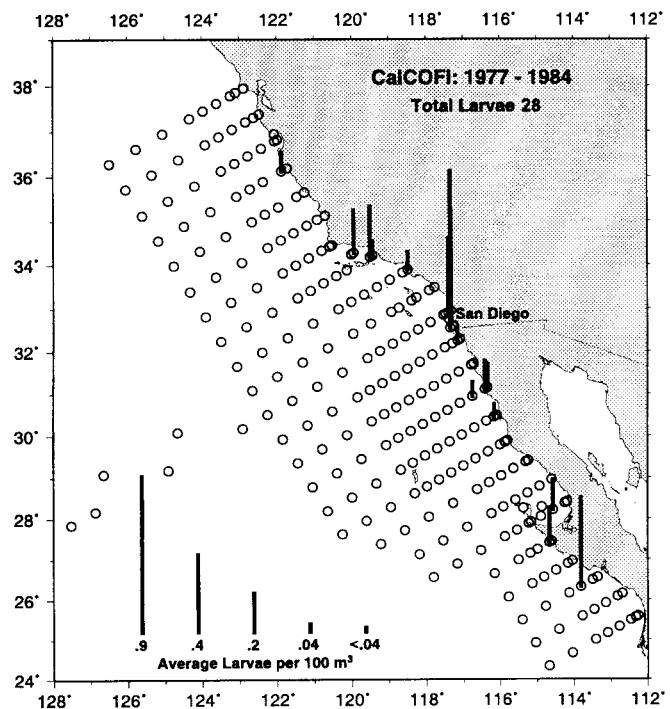
Pacific sanddab



Pleuronichthys verticalis

Hornyhead turbot

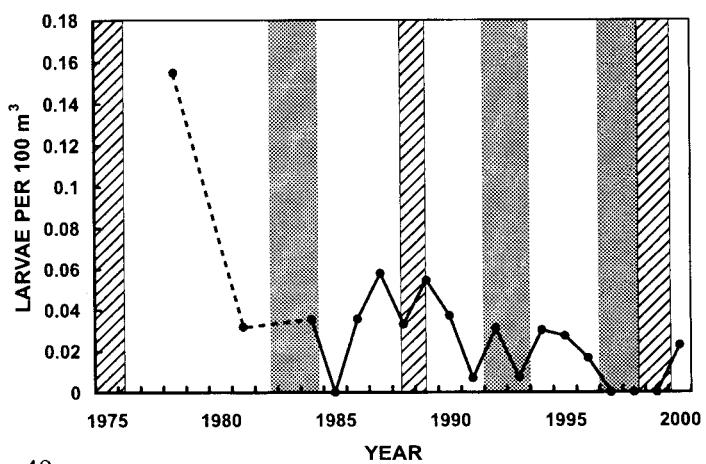
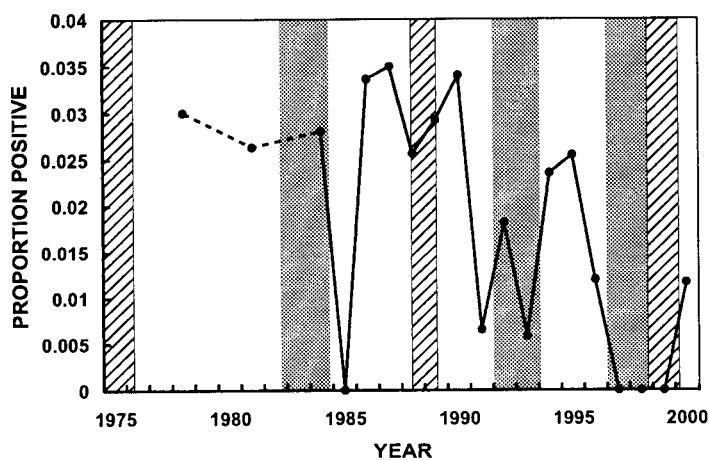
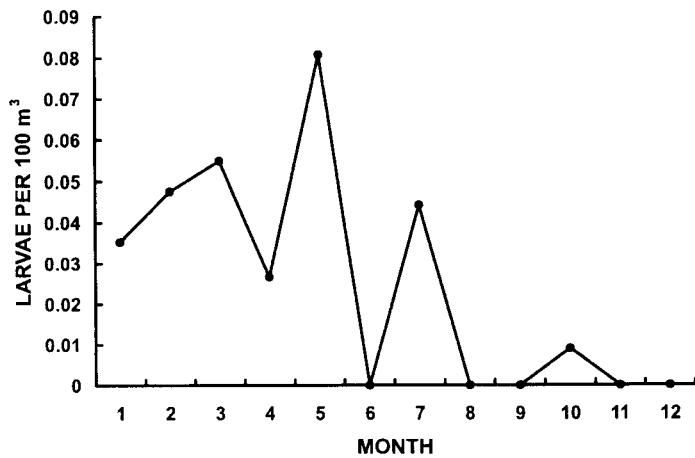
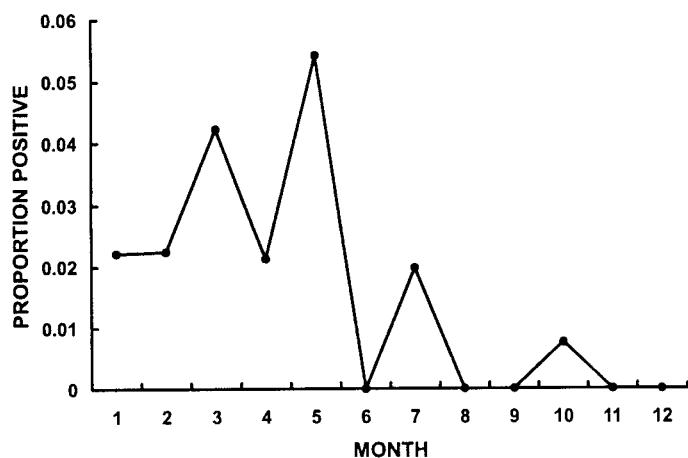
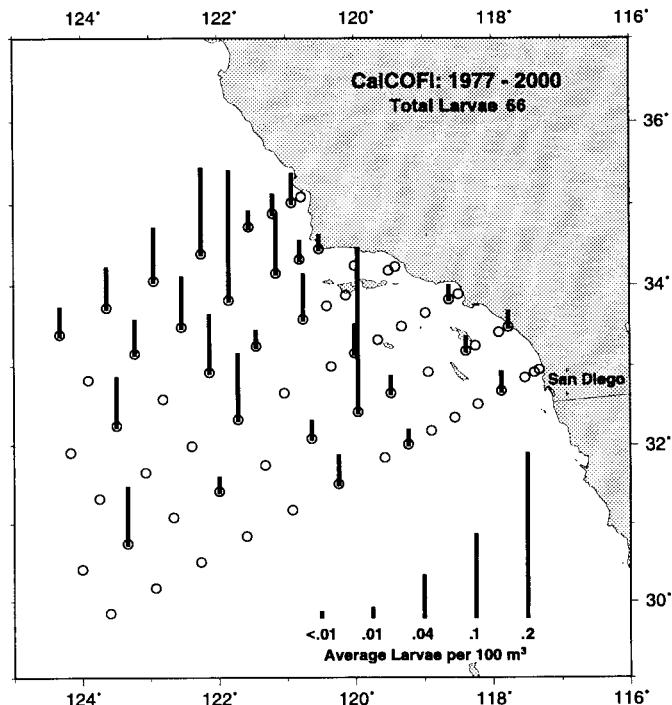
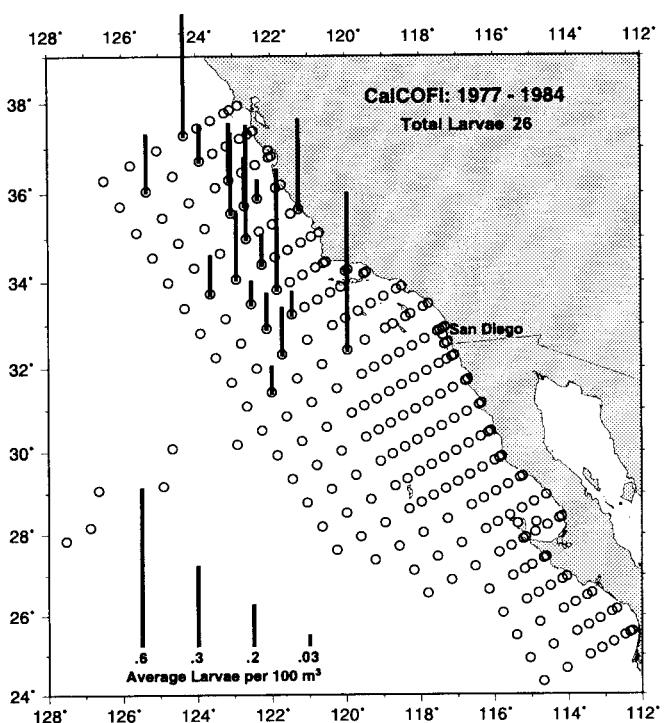
PLEURONECTIDAE



CENTROLOPHIDAE

Medusafish

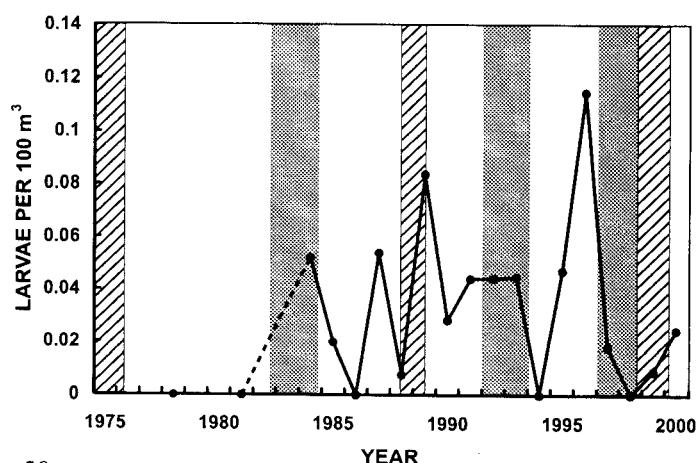
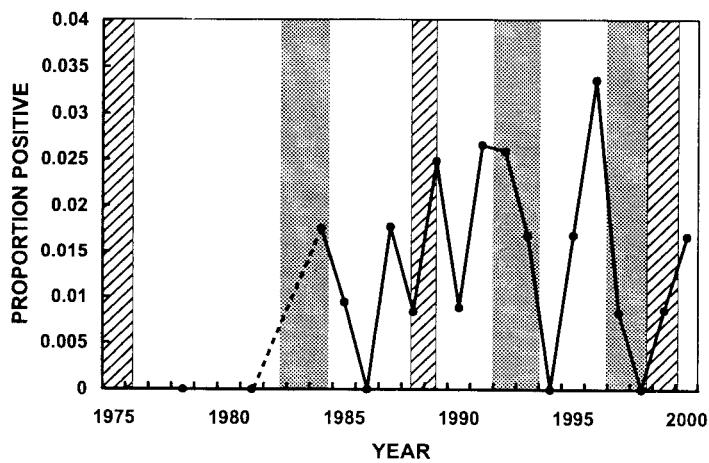
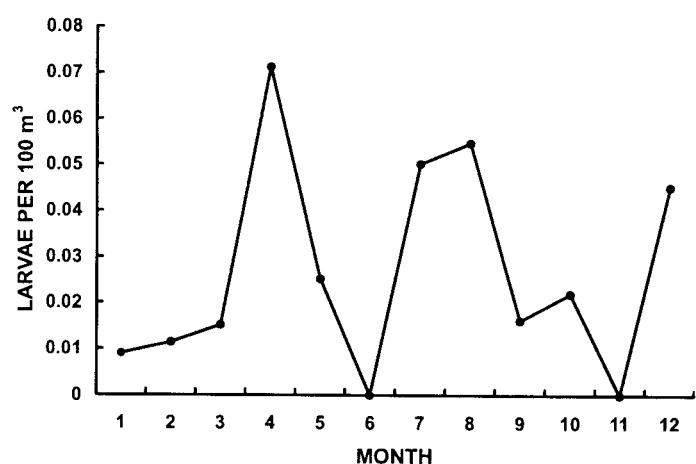
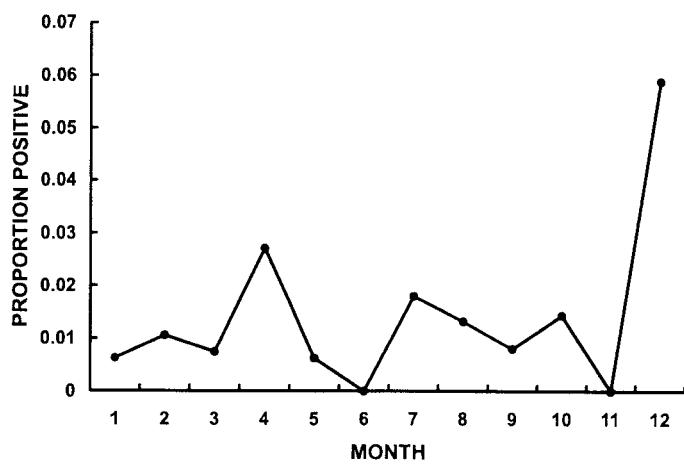
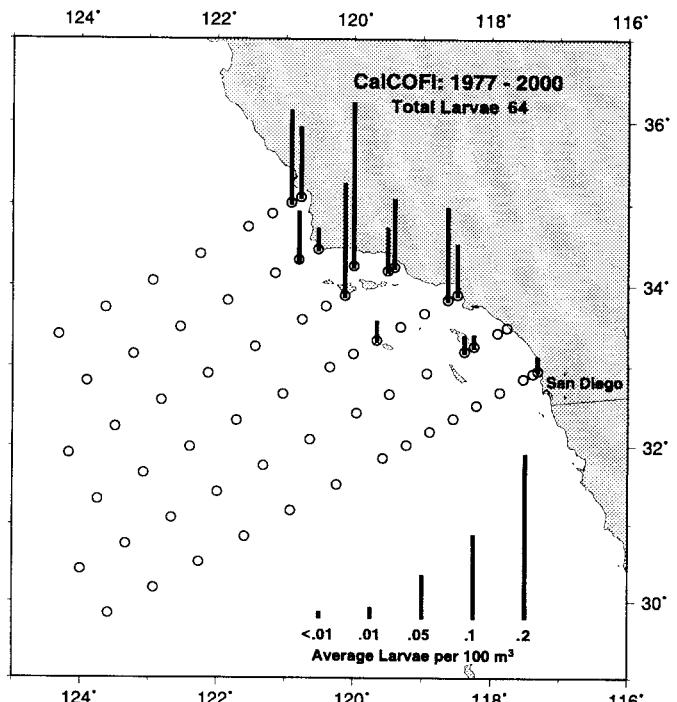
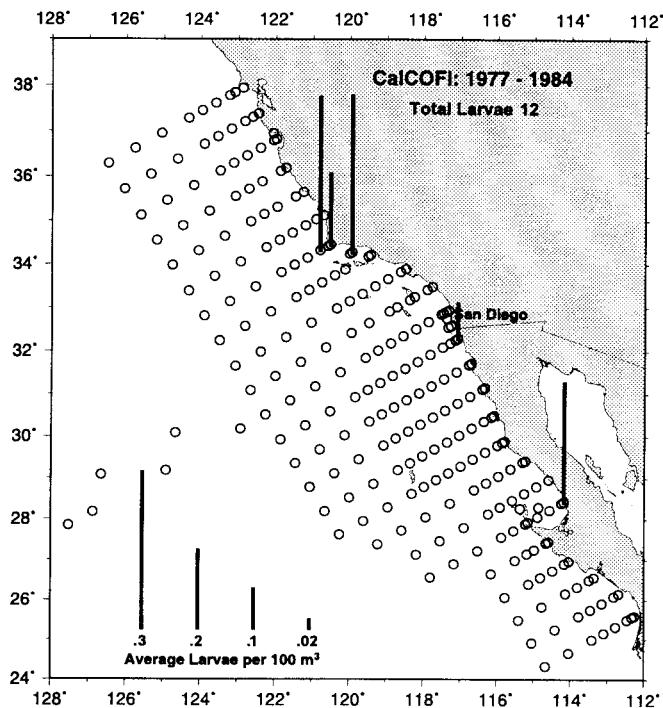
Icichthys lockingtoni



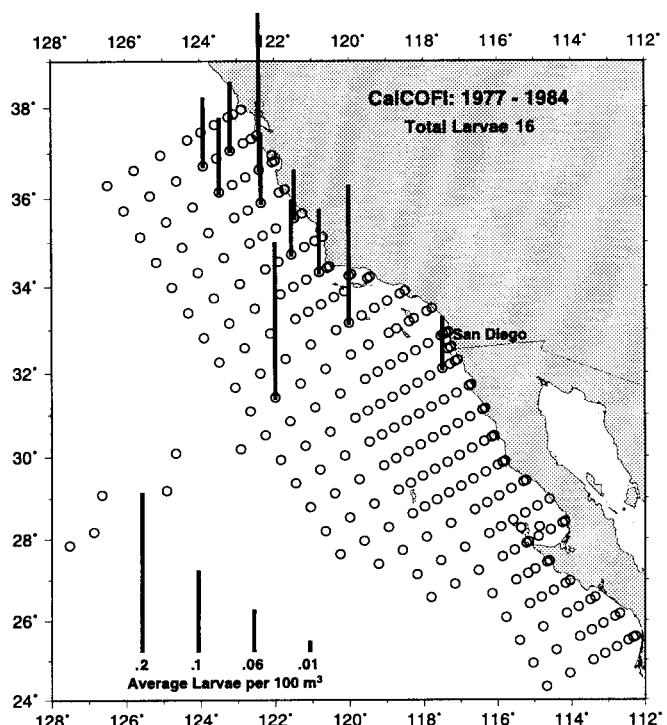
Neoclinus blanchardi

Sarcastic fringehead

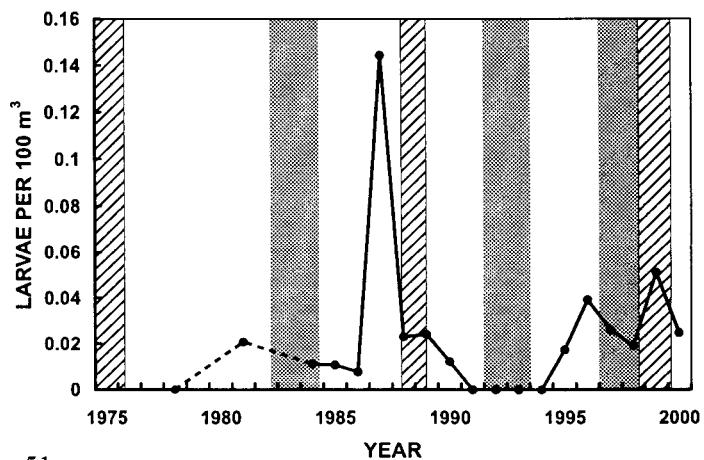
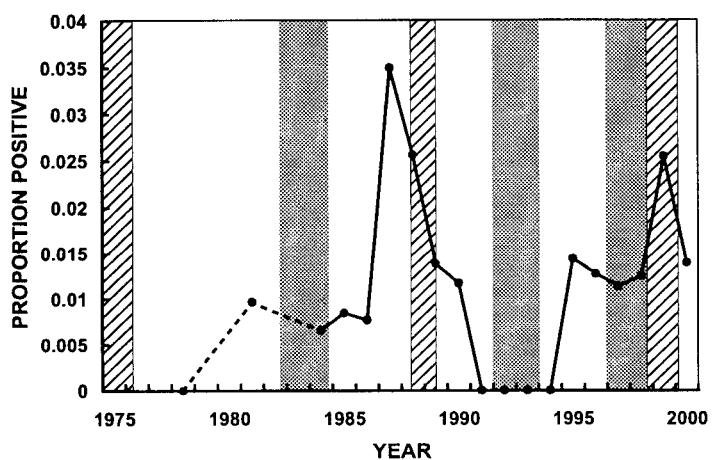
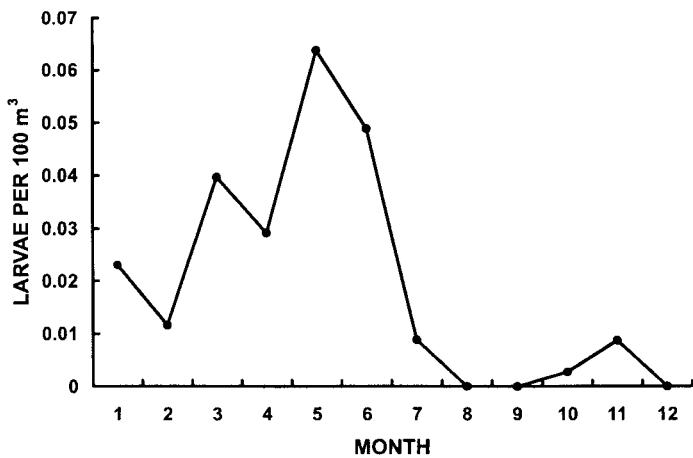
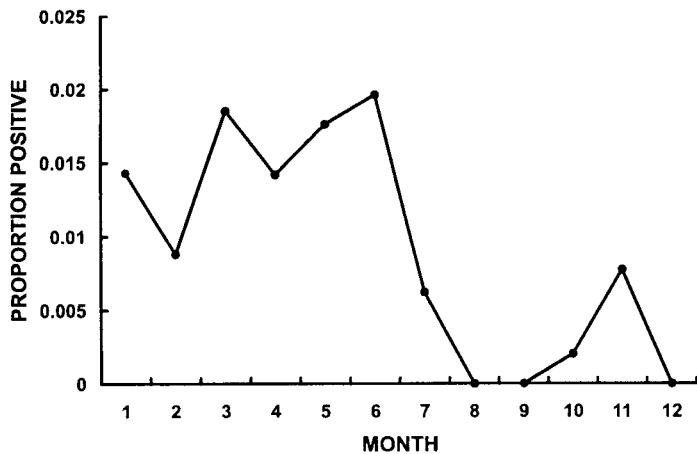
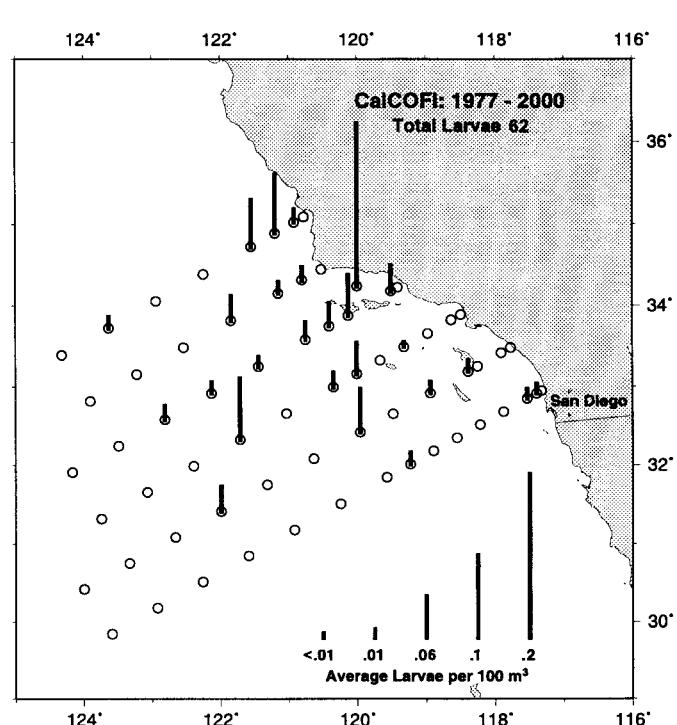
CHAENOPSIDAE



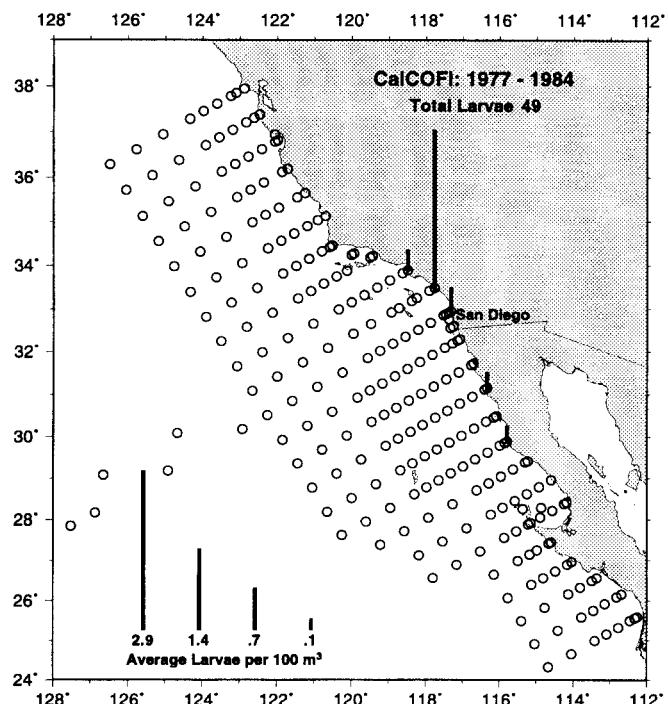
HEXAGRAMMIDAE



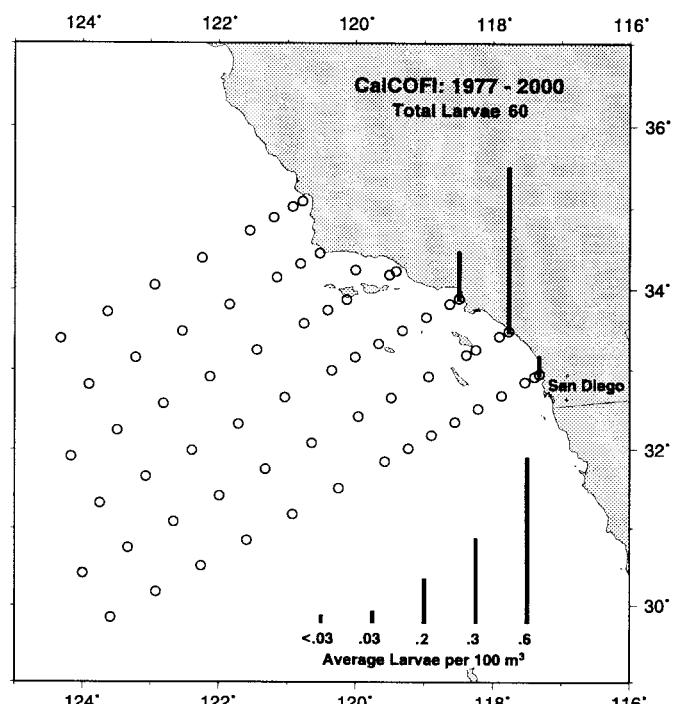
Painted greenling



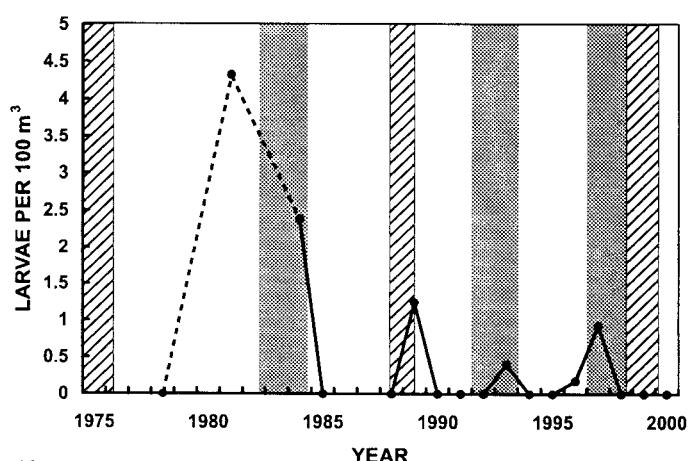
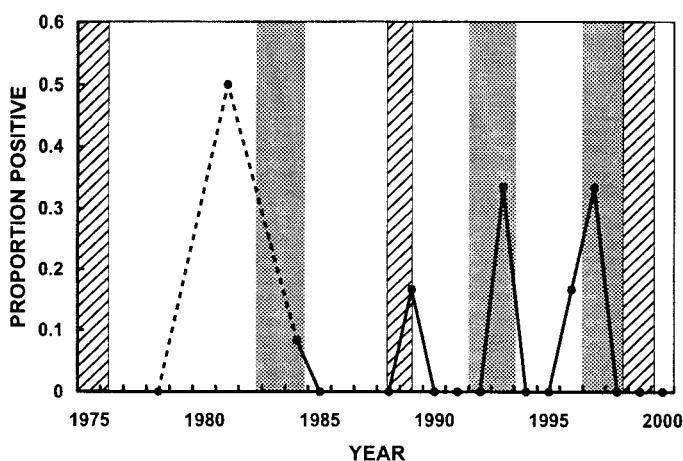
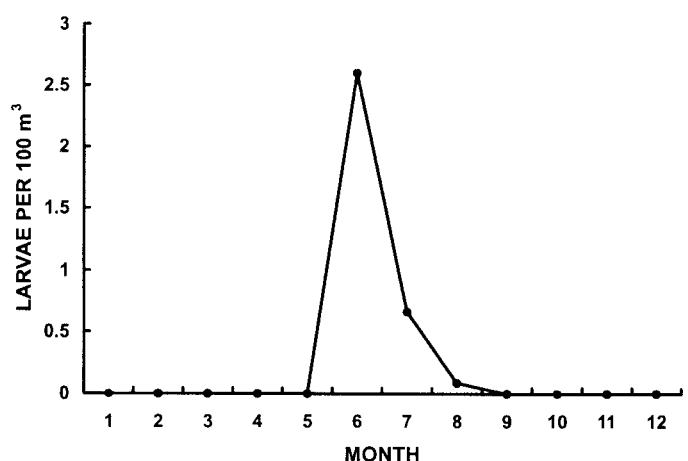
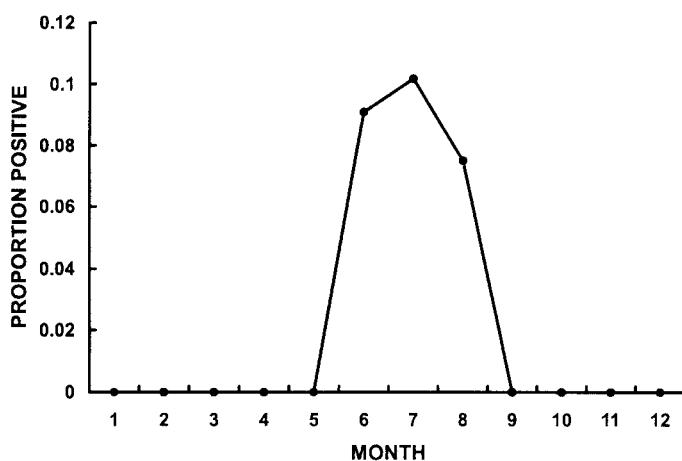
Anisotremus davidsoni



Sargo

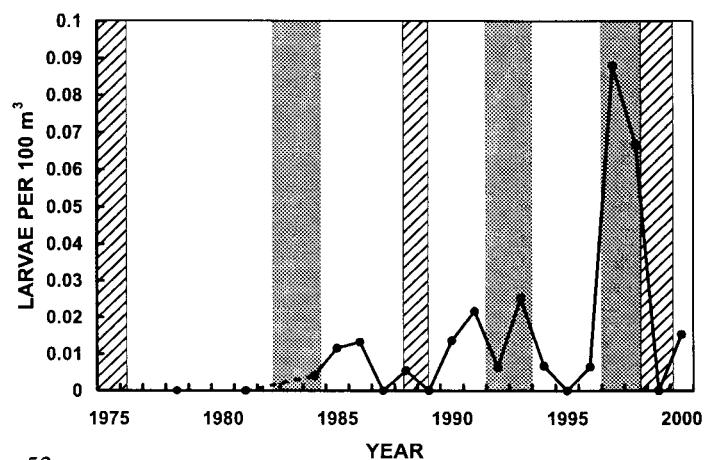
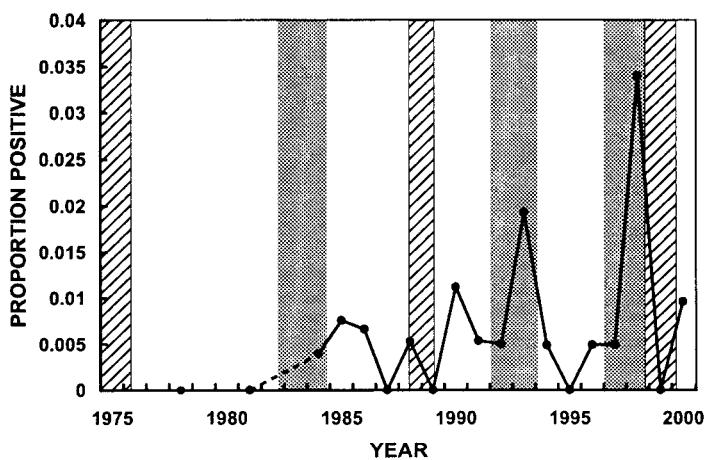
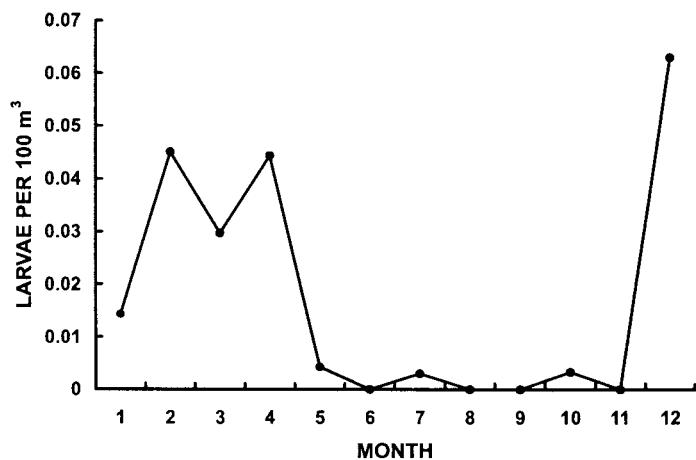
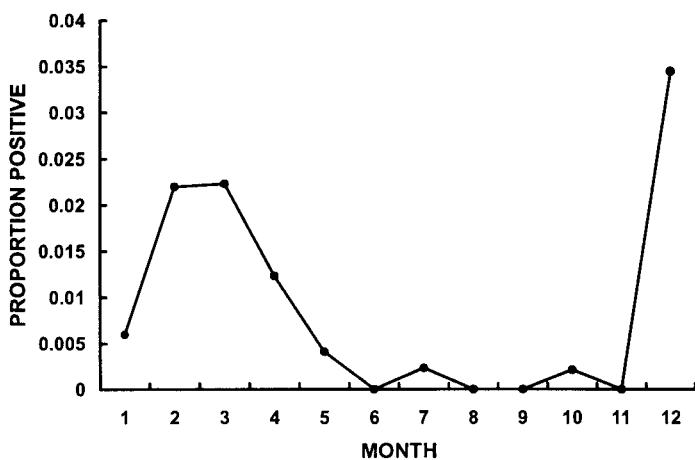
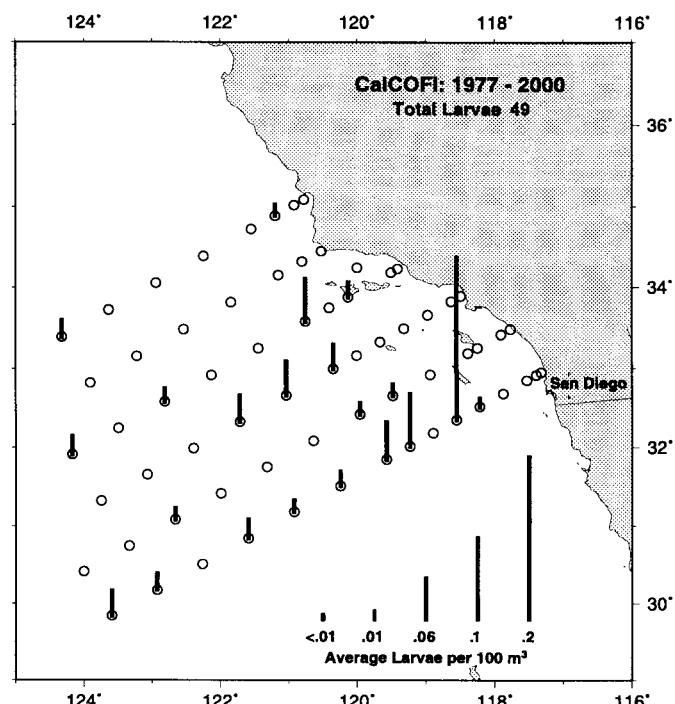
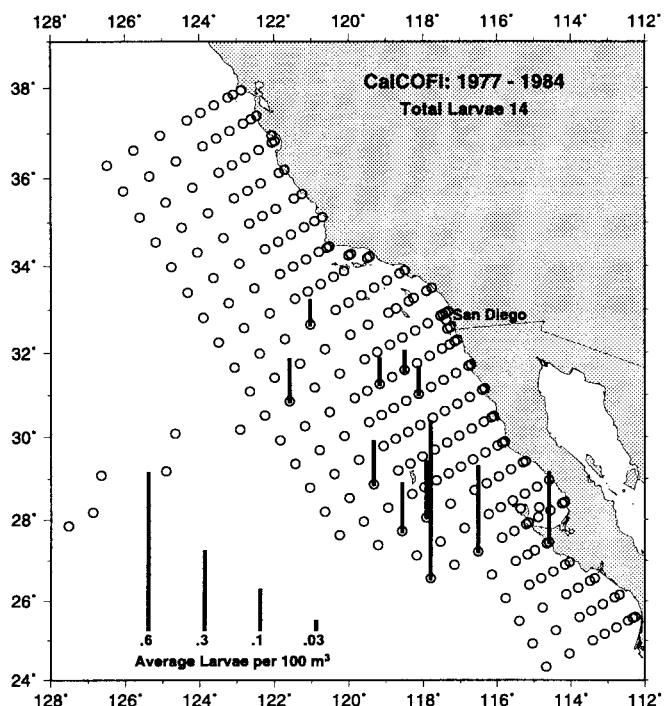


HAEMULIDAE

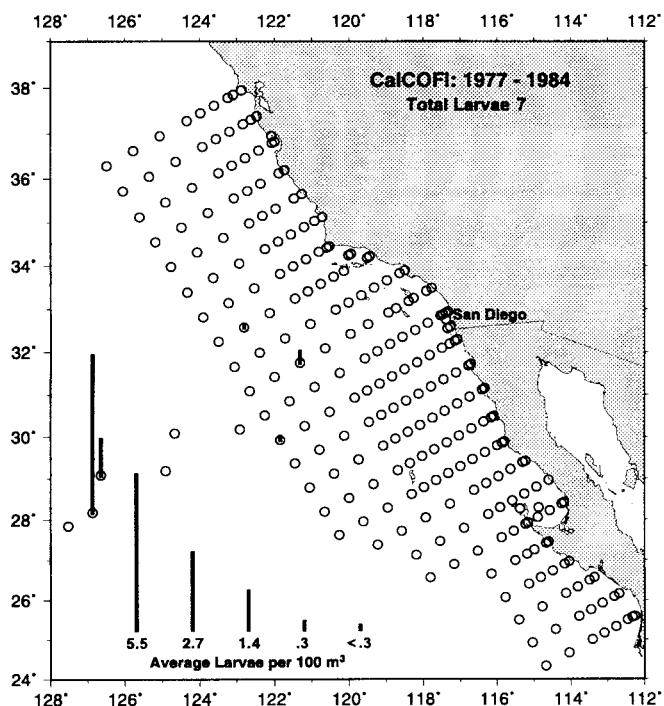


STOMIIDAE

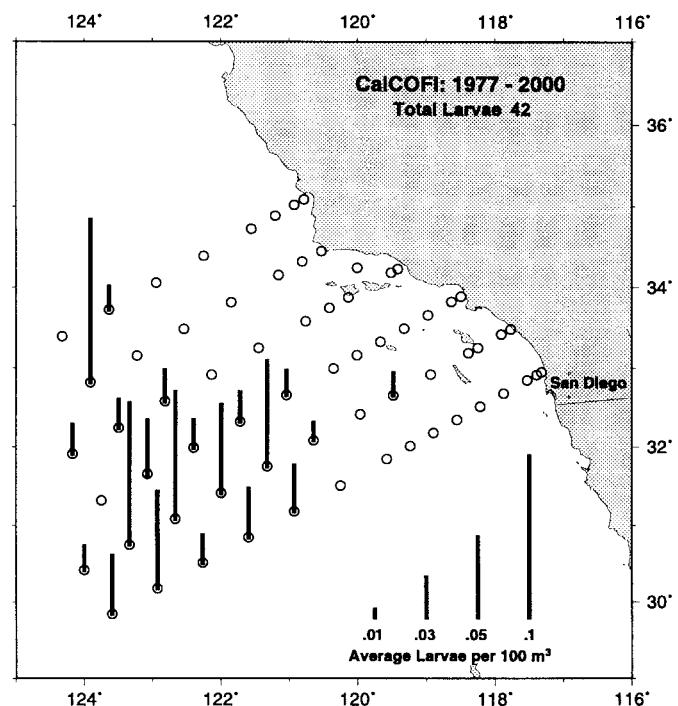
Blackbelly dragonfish

Stomias atriventer

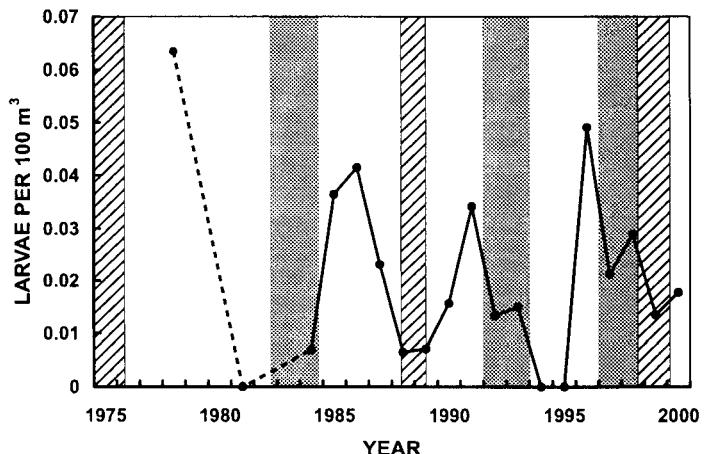
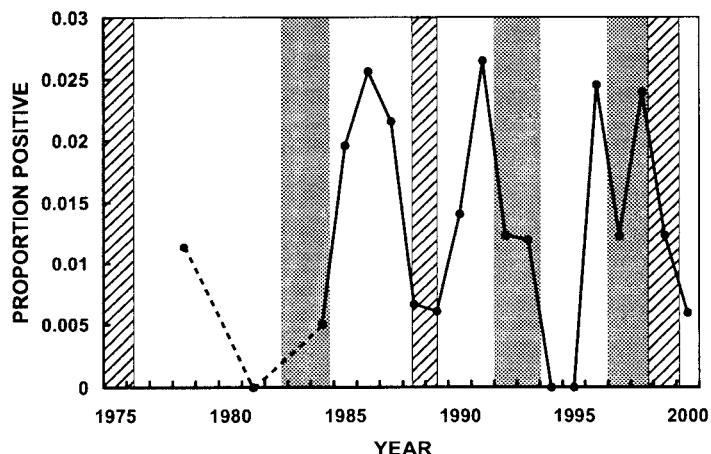
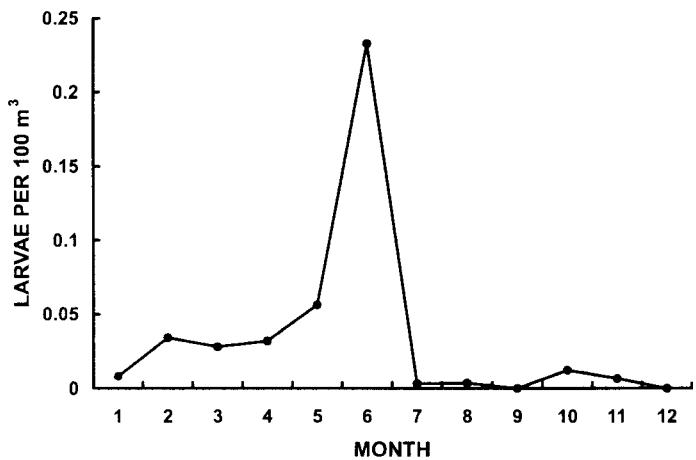
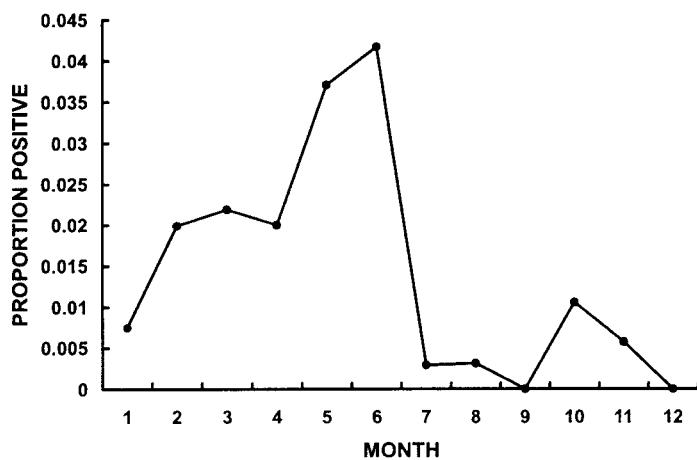
Aristostomias scintillans



Shining loosejaw



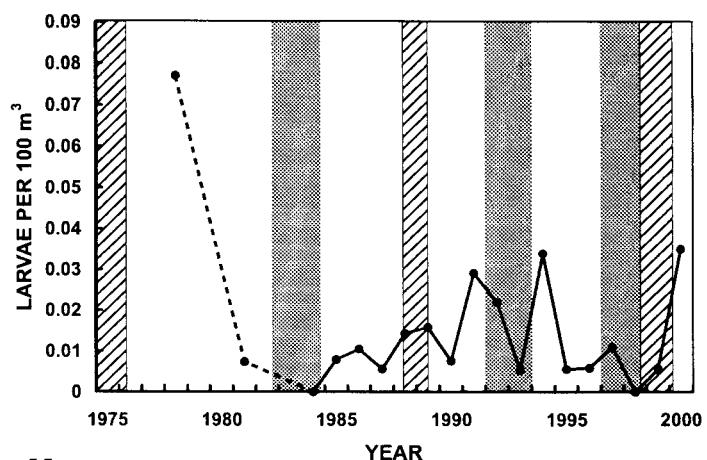
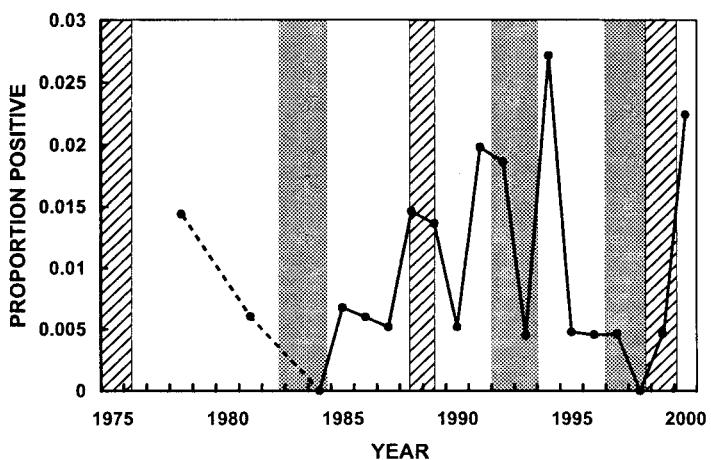
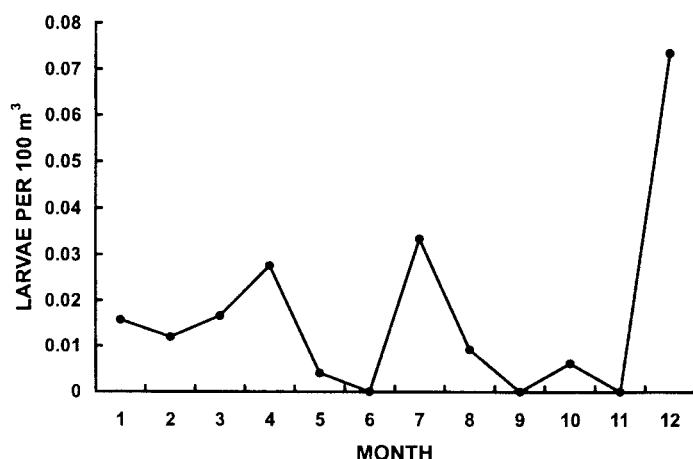
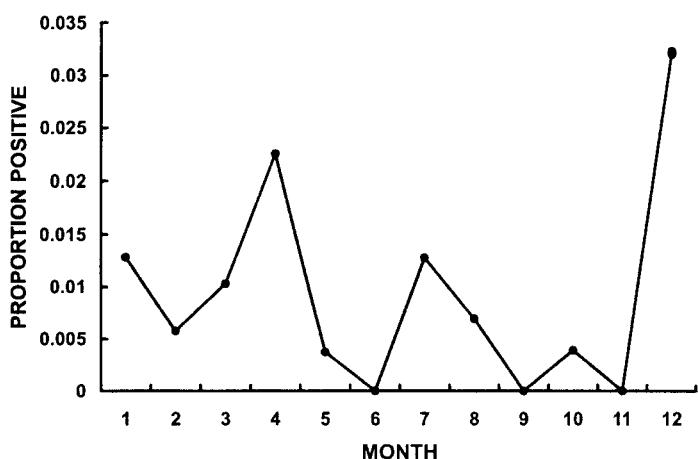
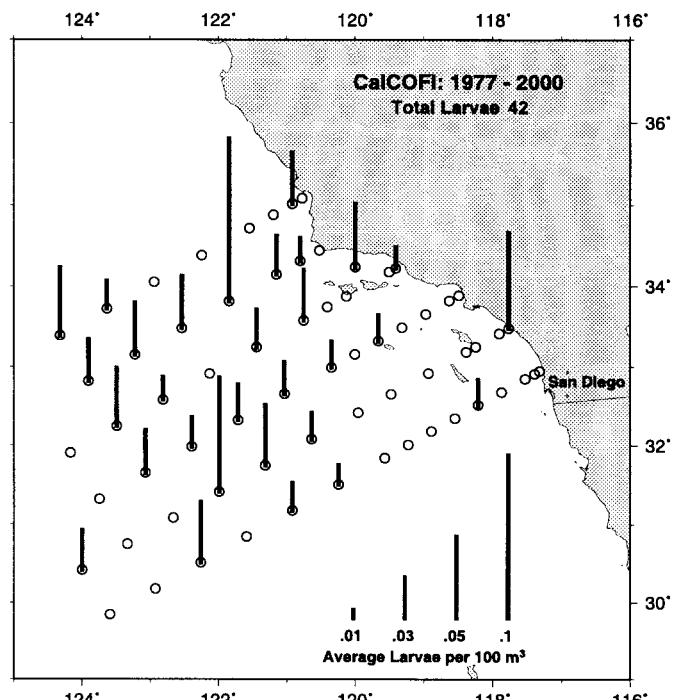
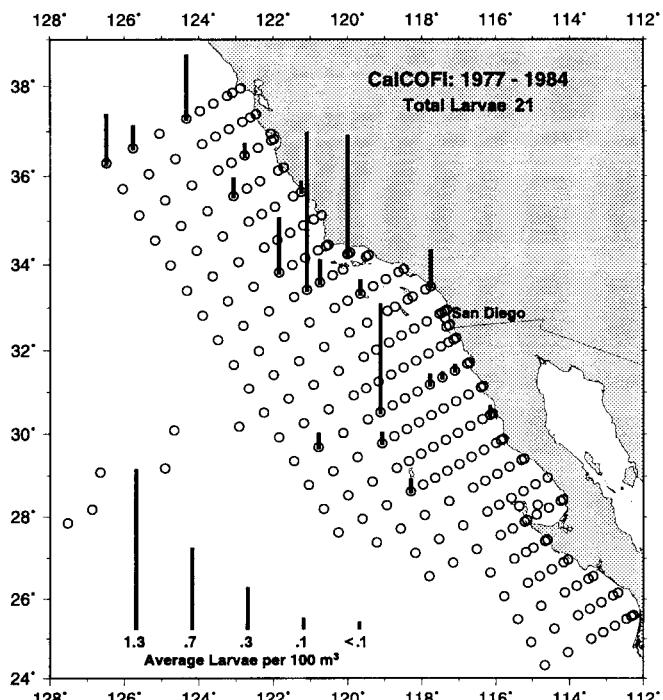
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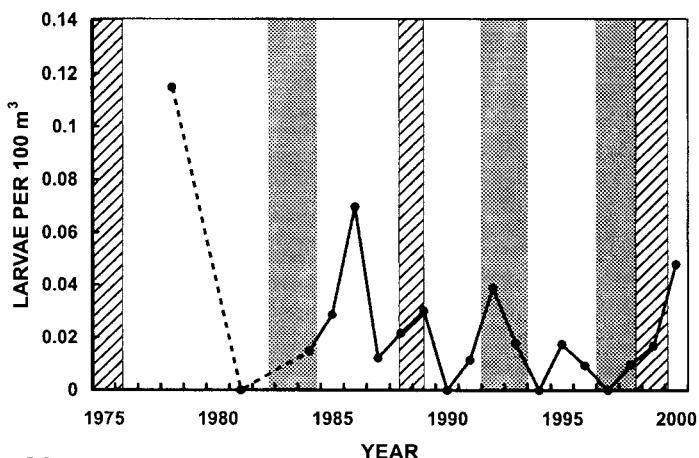
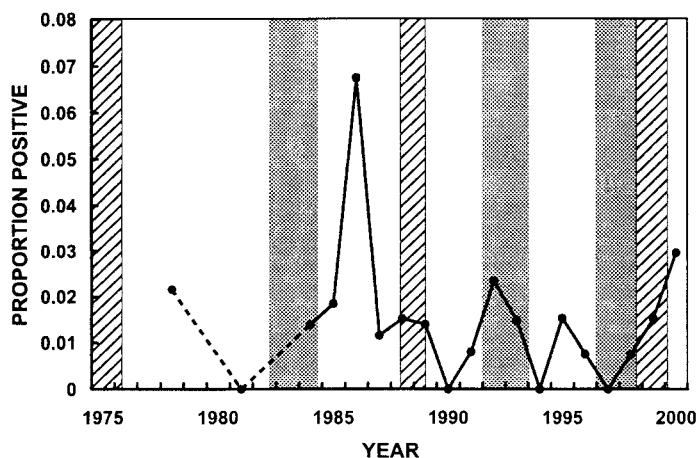
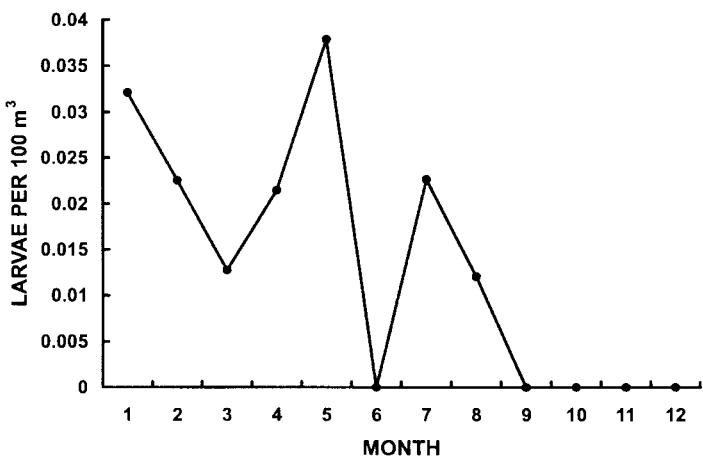
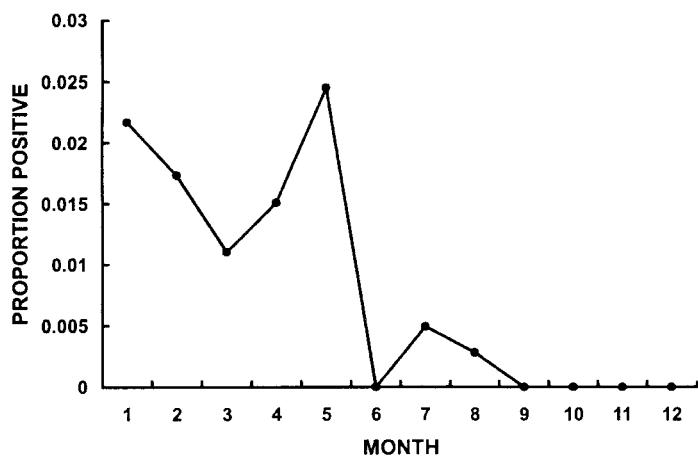
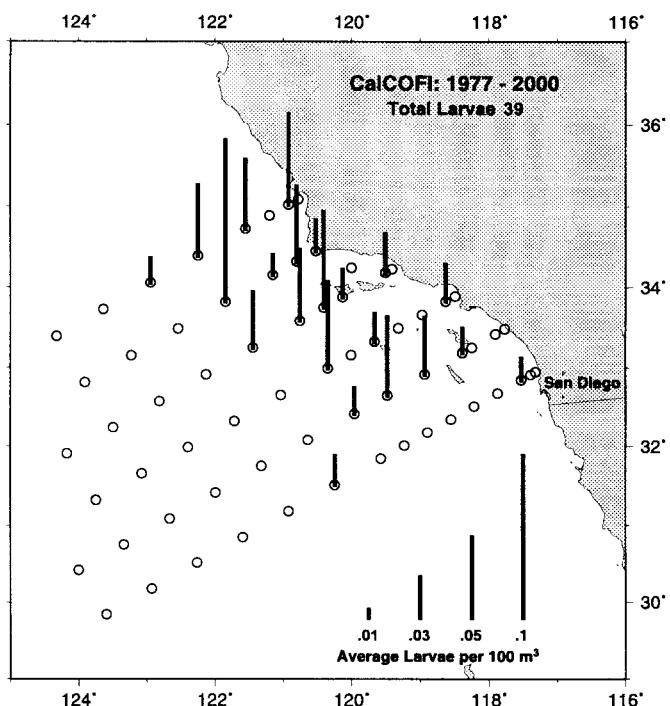
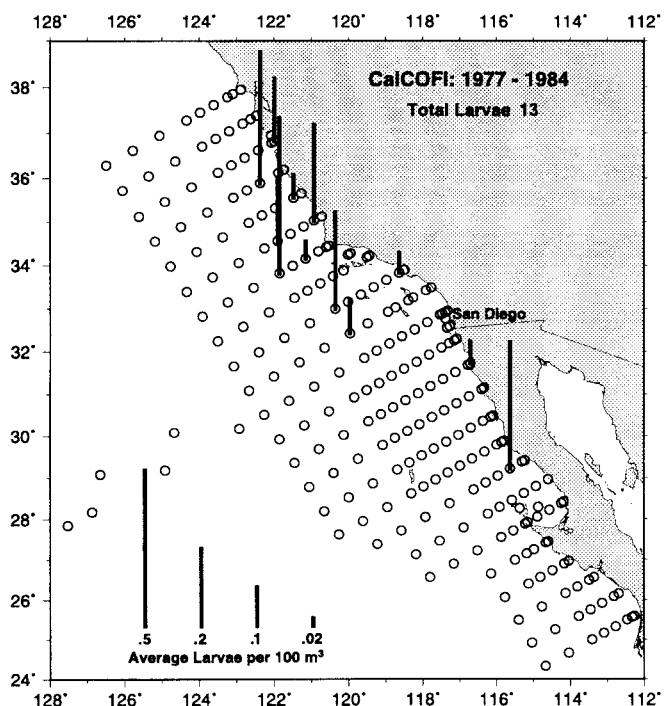


MYCTOPHIDAE

Broadfin lampfish

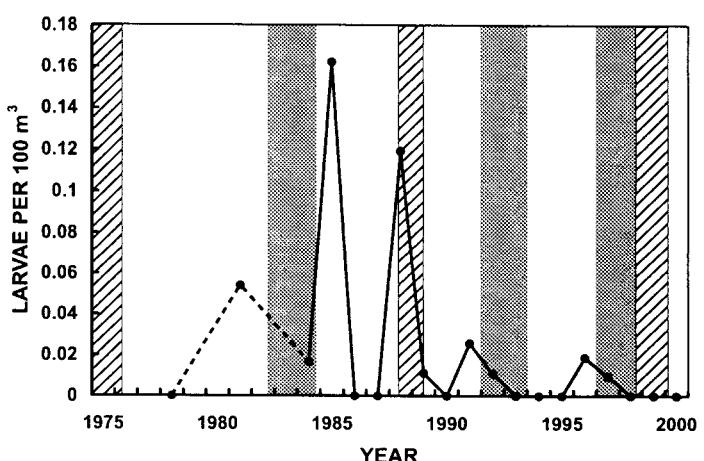
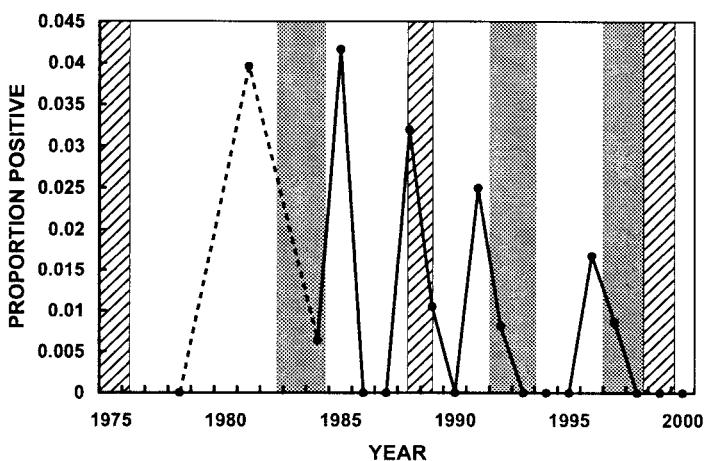
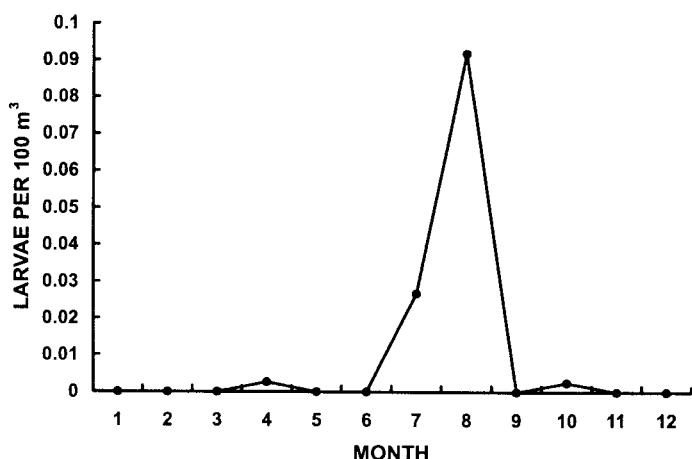
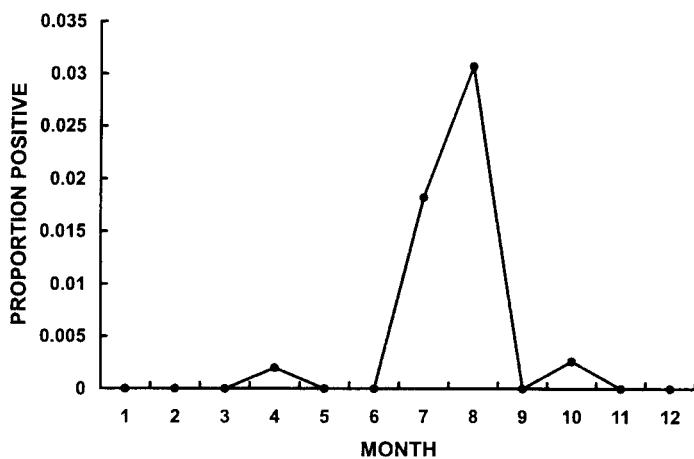
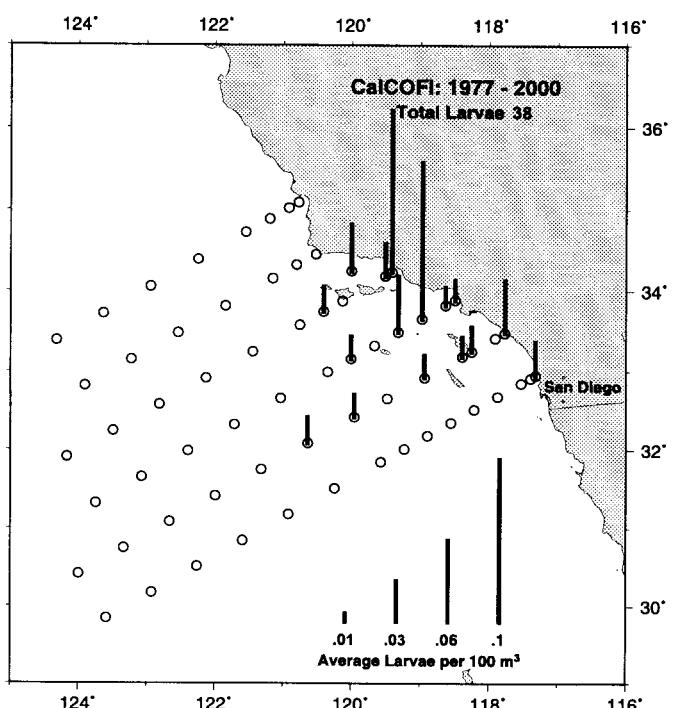
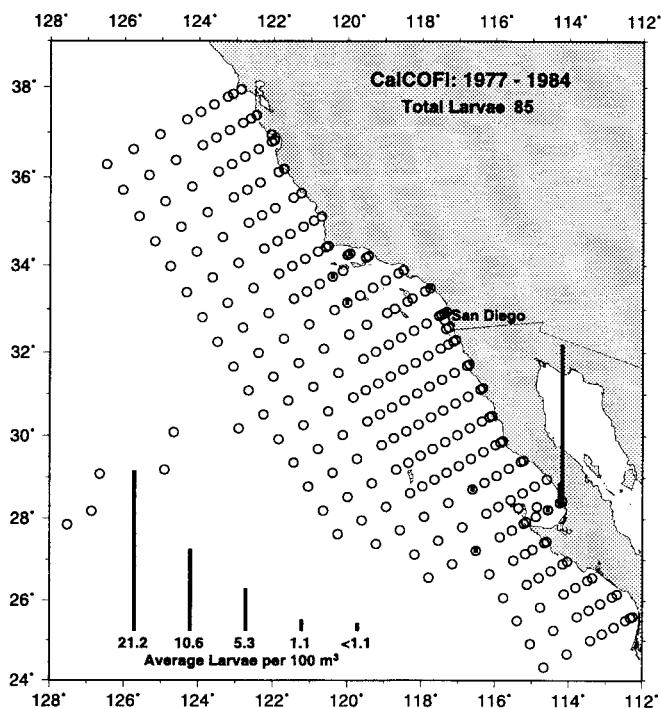
Nannobrachium ritteri



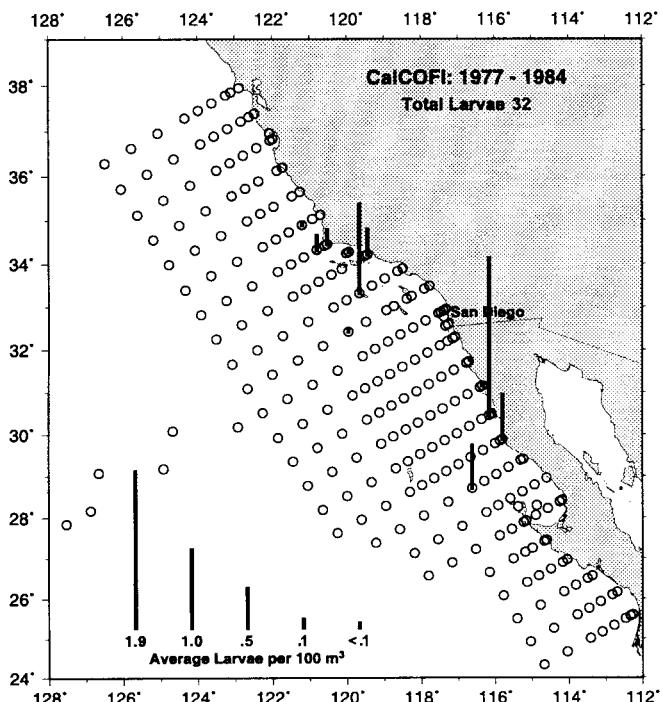
Sebastes aurora**Aurora rockfish****SEBASTIDAE**

LABRIDAE

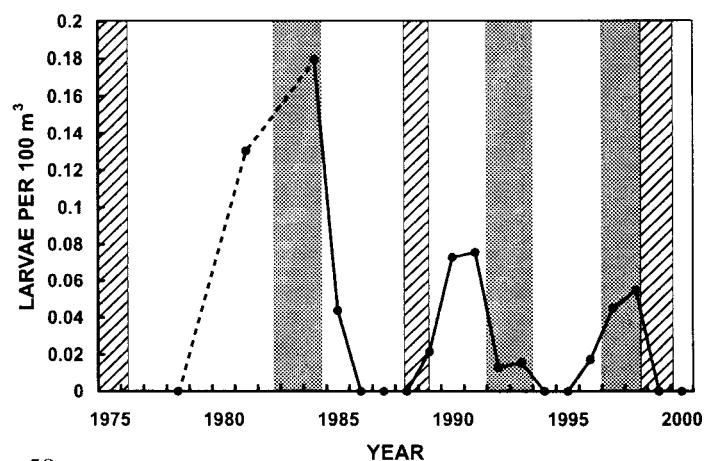
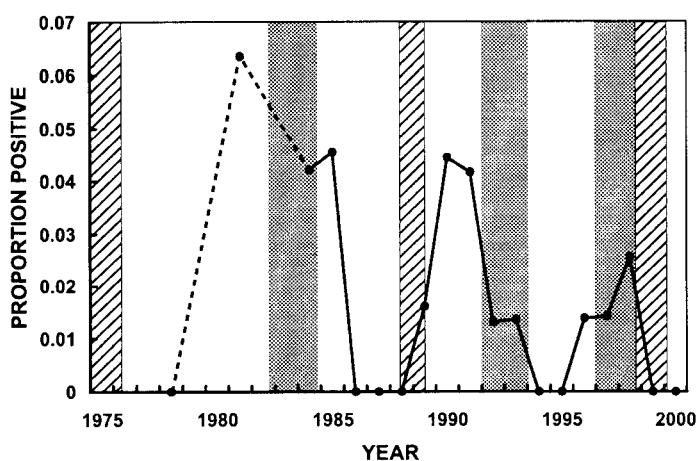
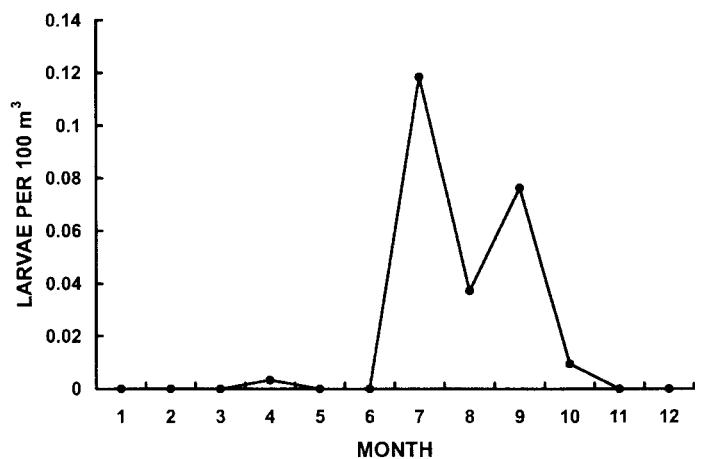
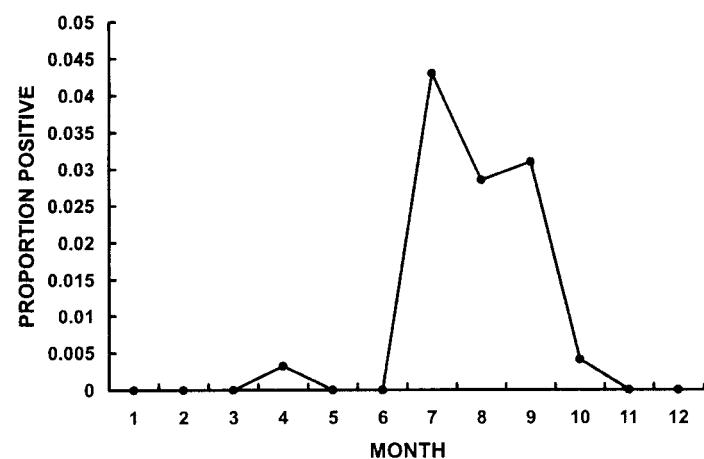
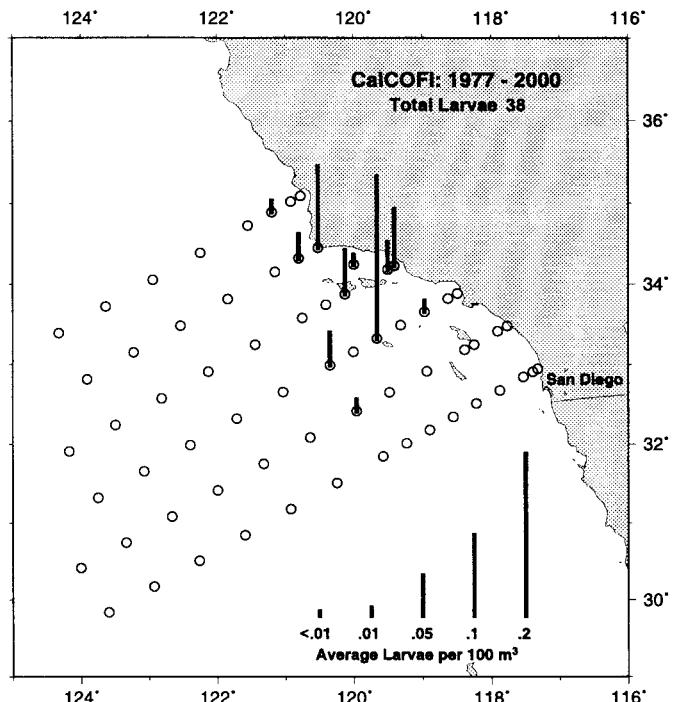
Rock wrasse

Halichoeres semicinctus

Semicossyphus pulcher

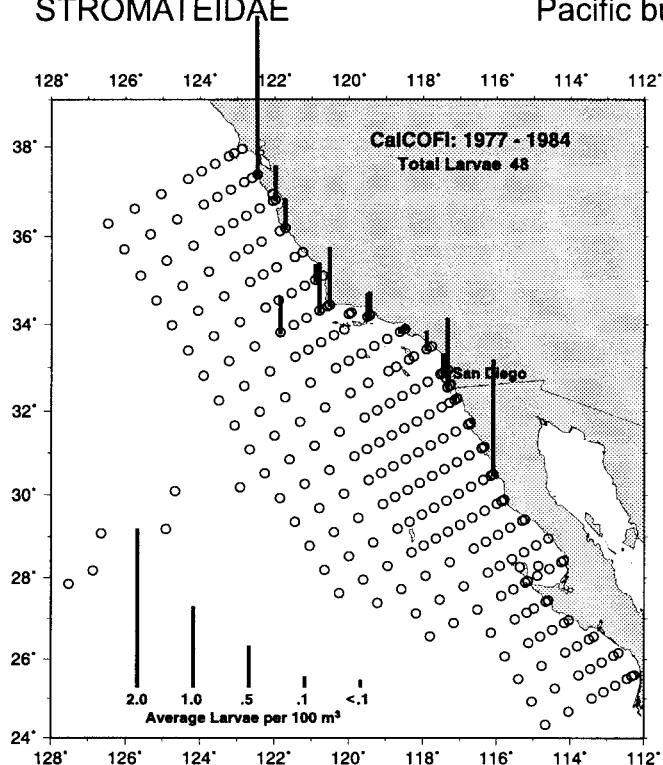


Sheephead

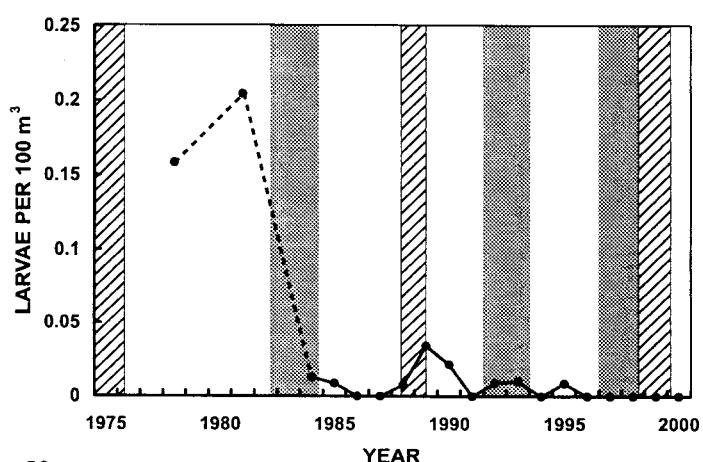
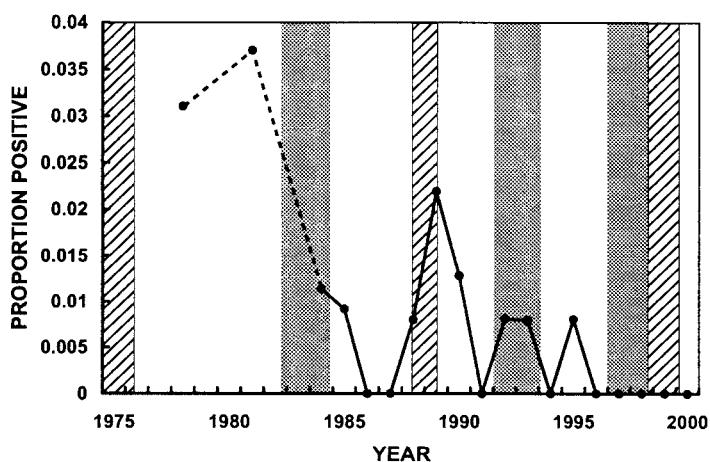
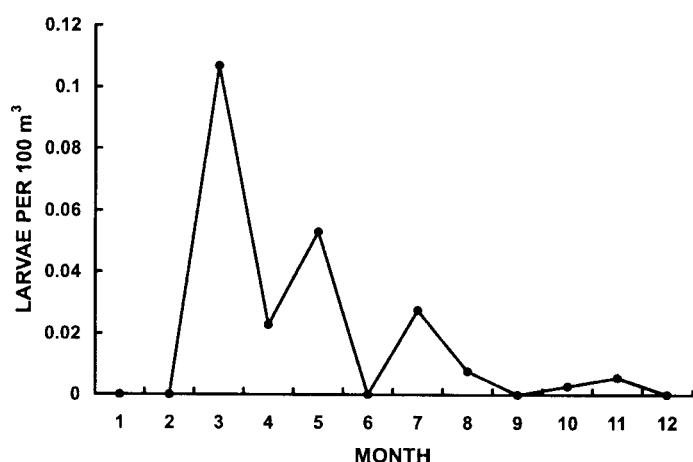
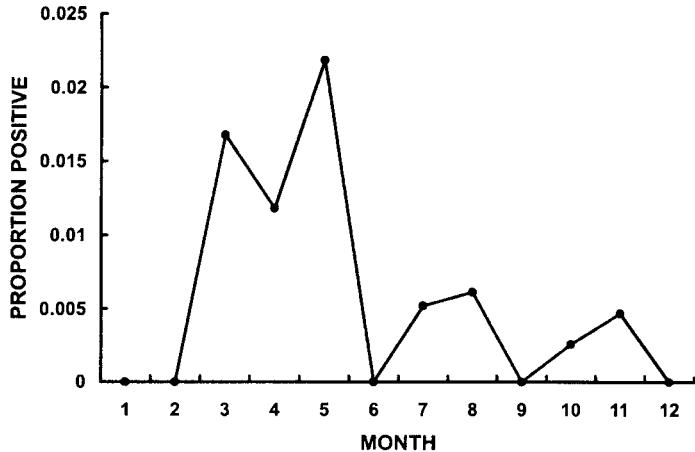
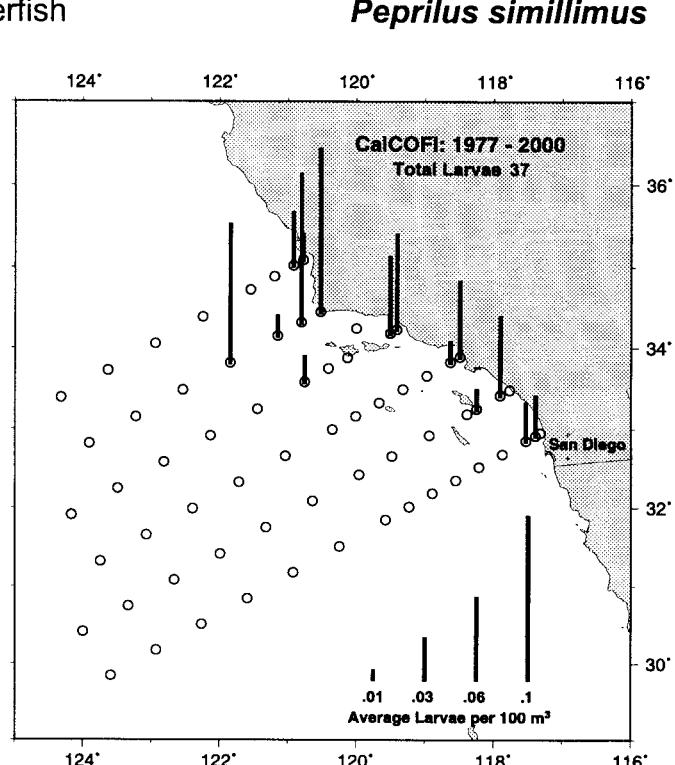


LABRIDAE

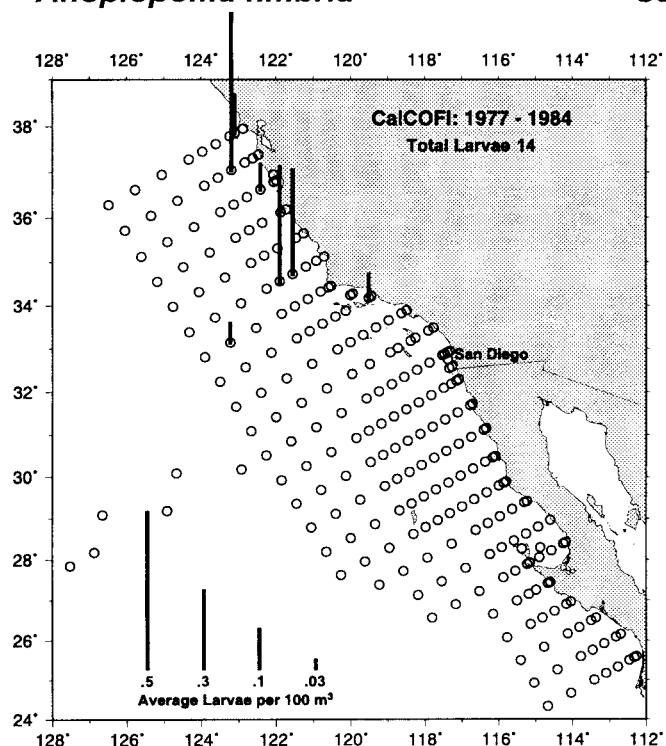
STROMATEIDAE



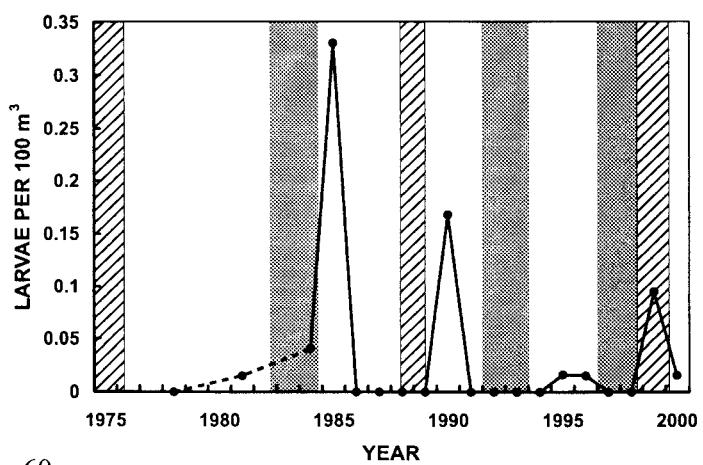
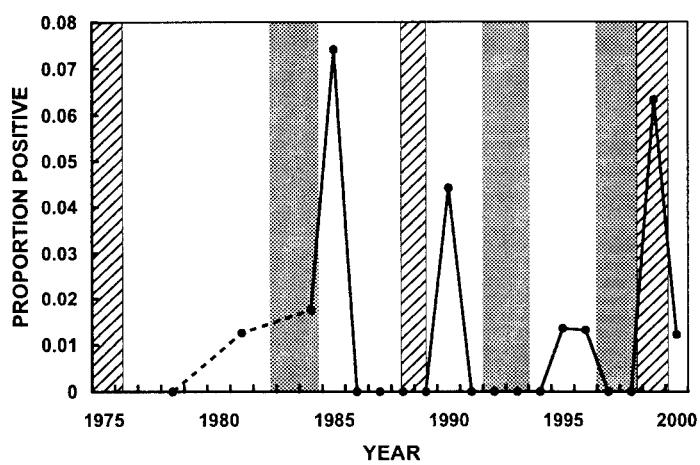
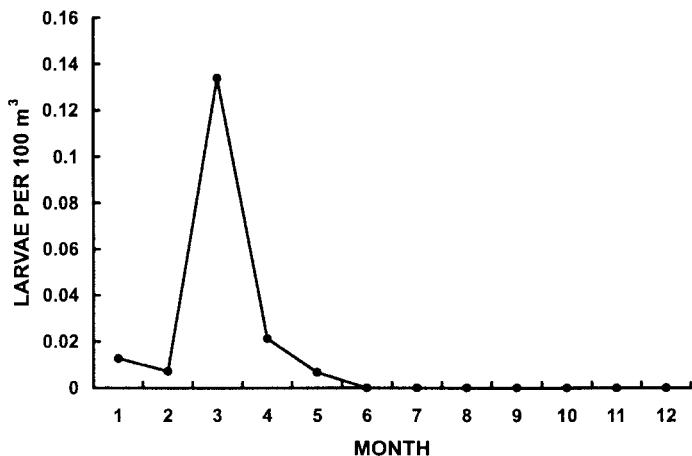
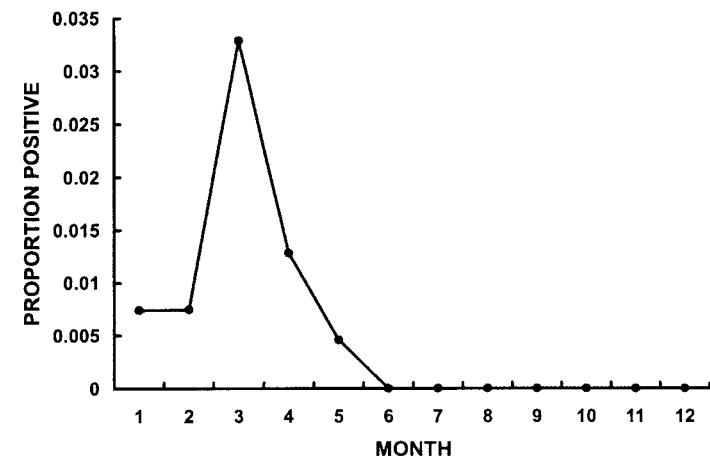
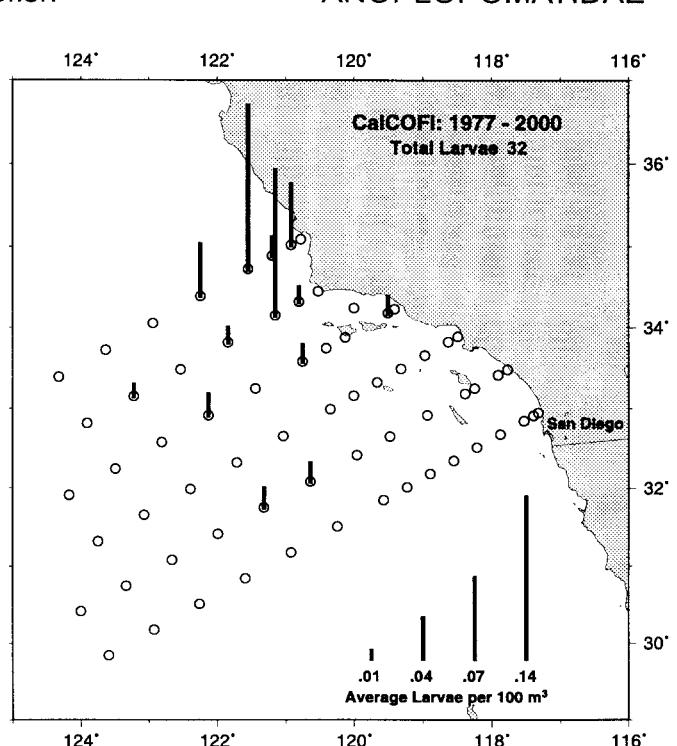
Pacific butterfish



Anoplopoma fimbria



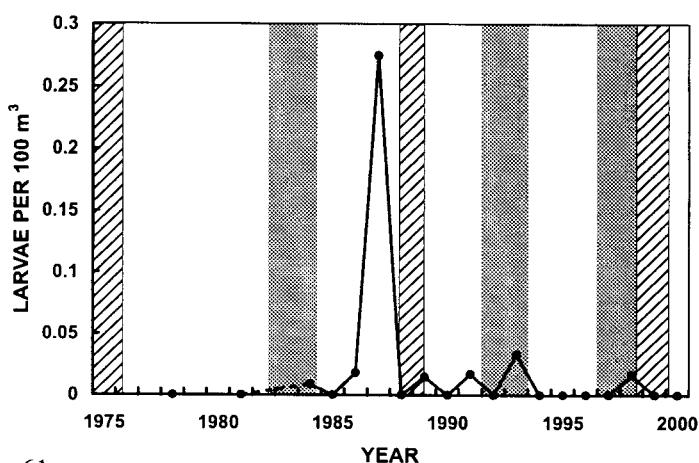
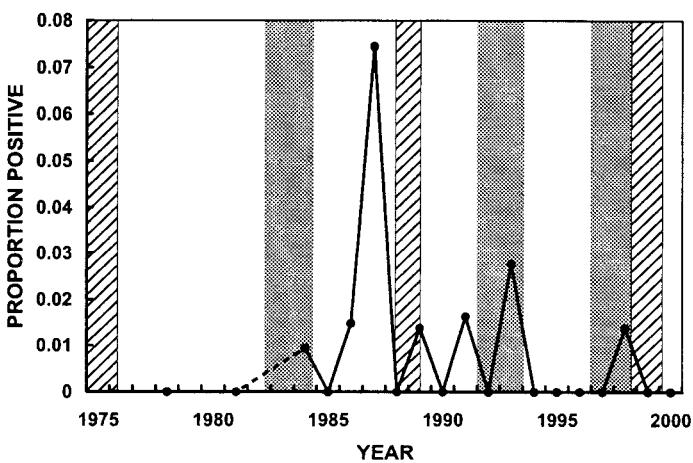
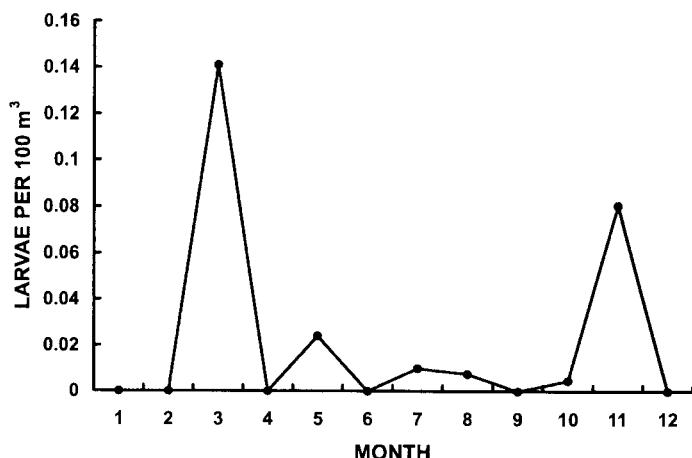
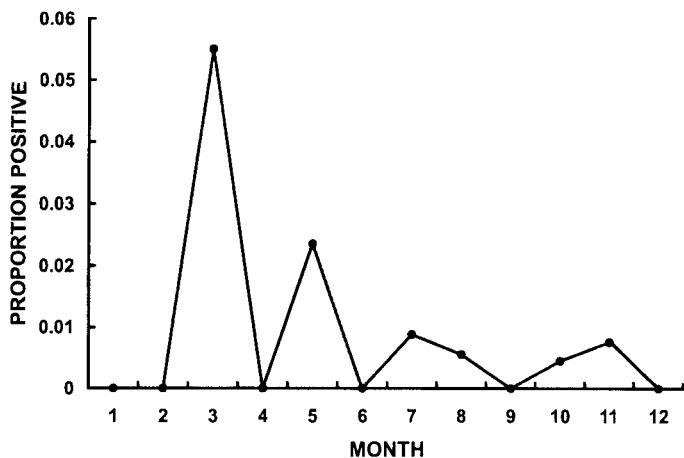
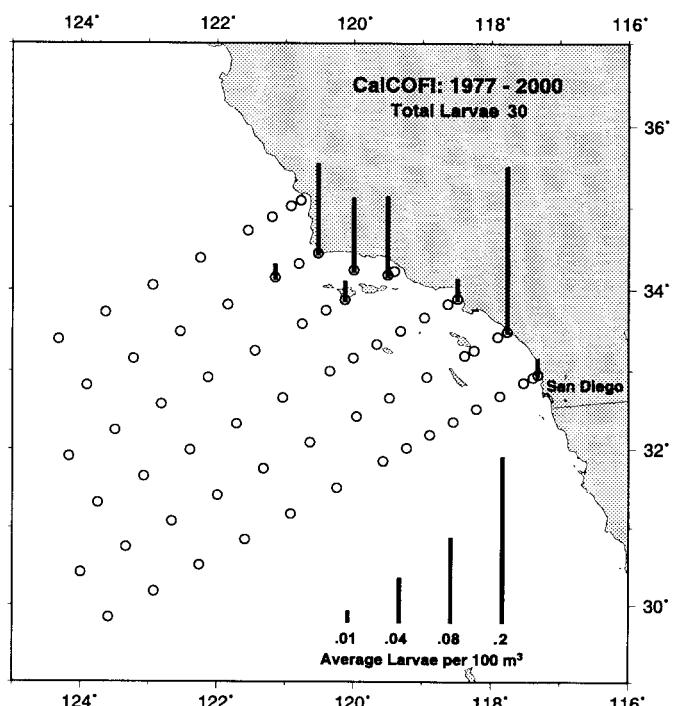
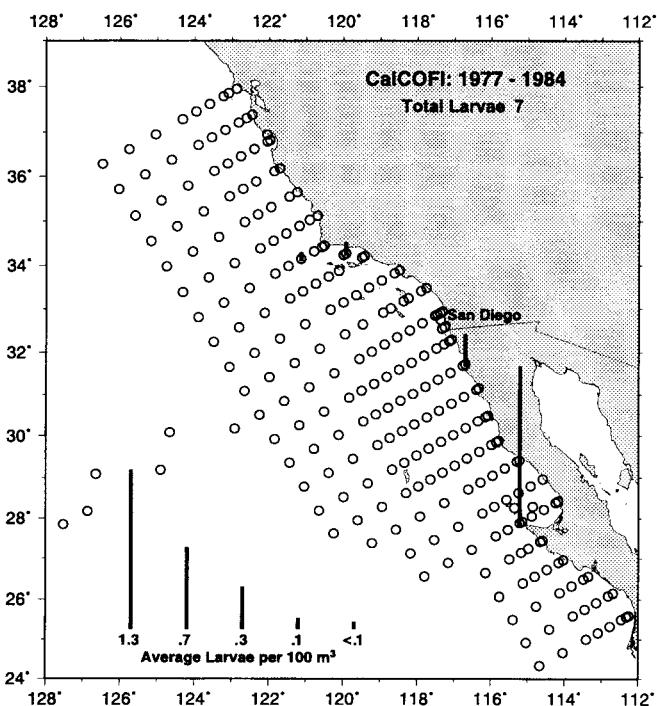
Sablefish



ATHERINIDAE

Topsmelt

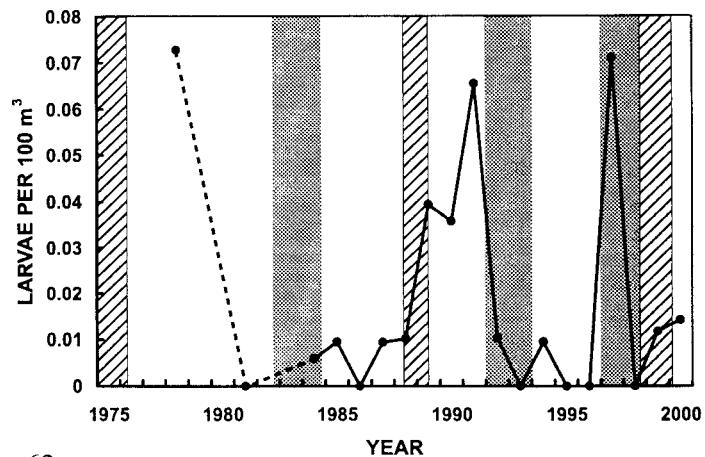
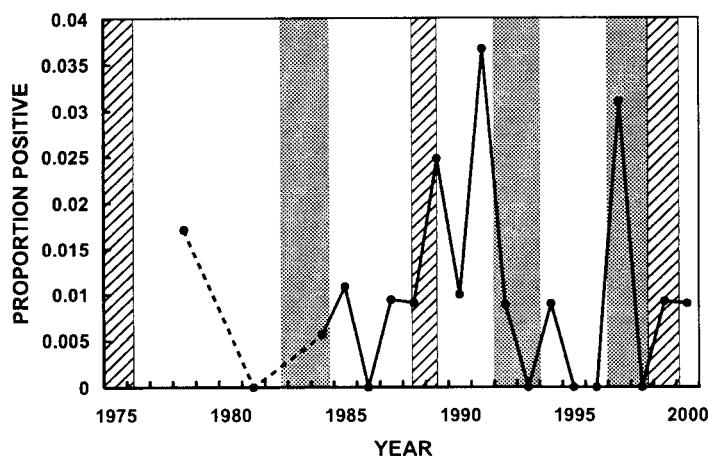
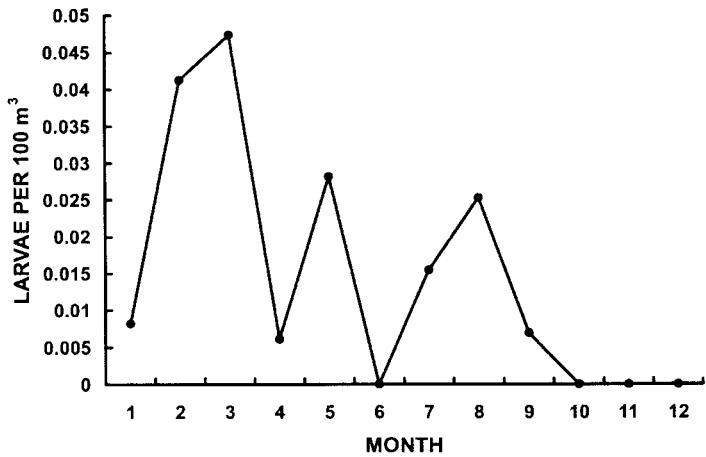
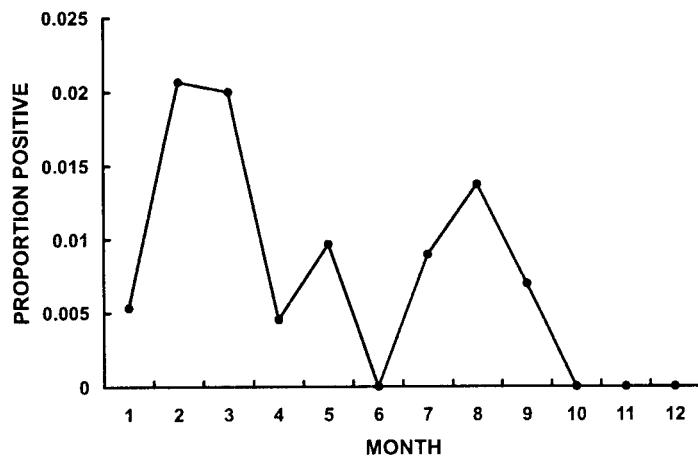
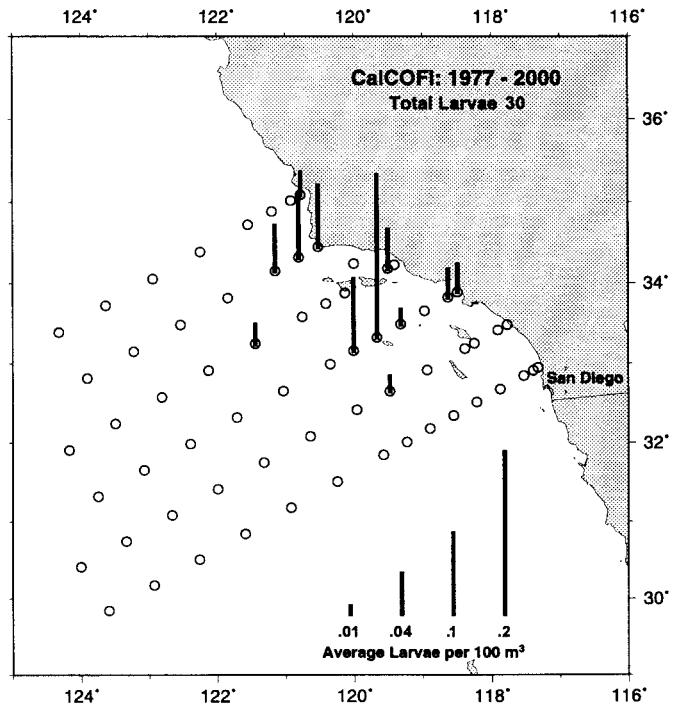
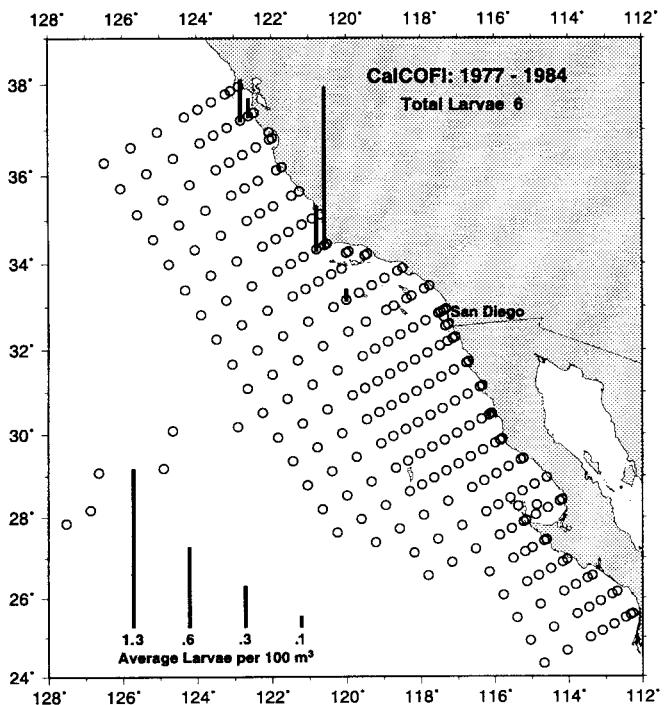
Atherinops affinis



Coryphopterus nicholsii

Blackeye goby

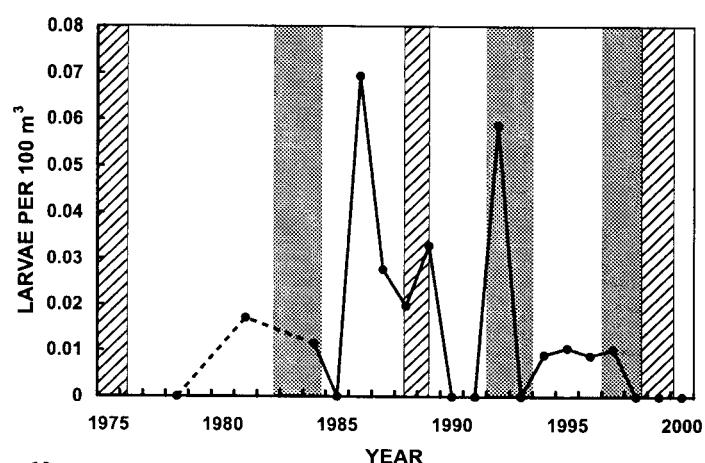
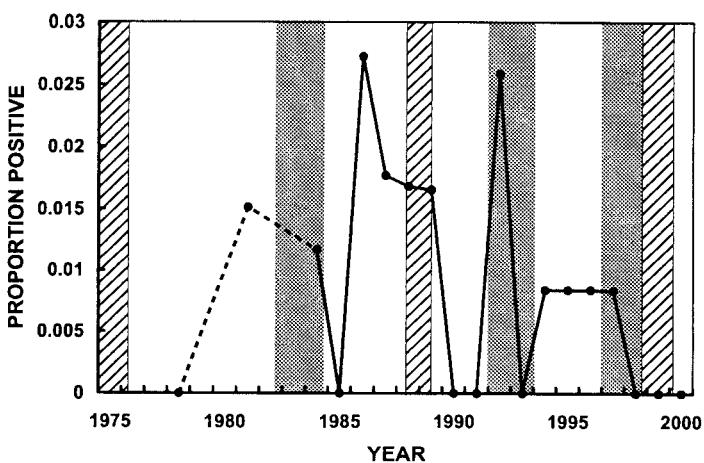
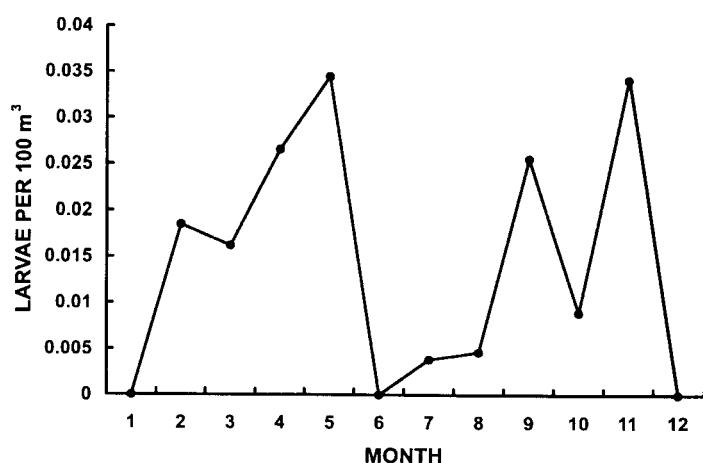
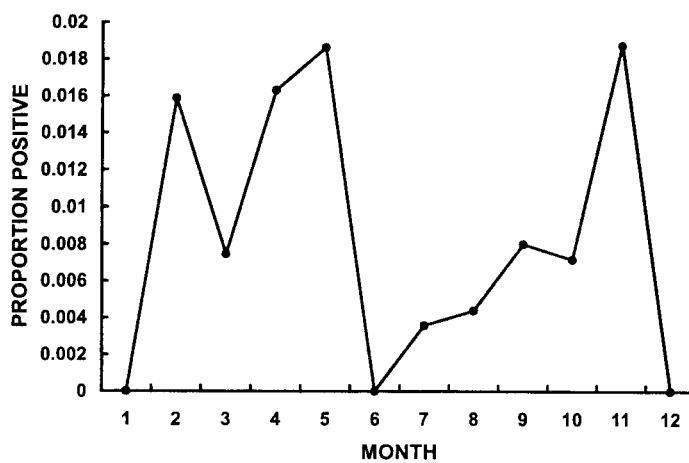
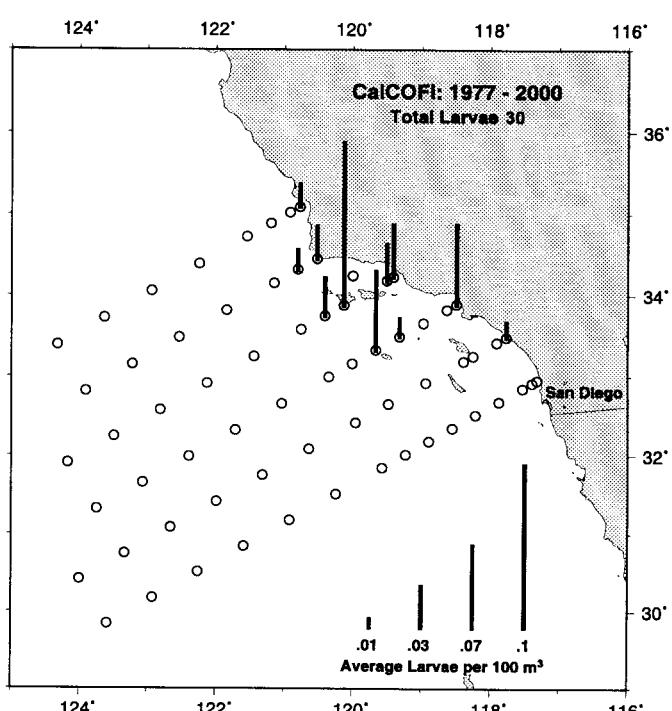
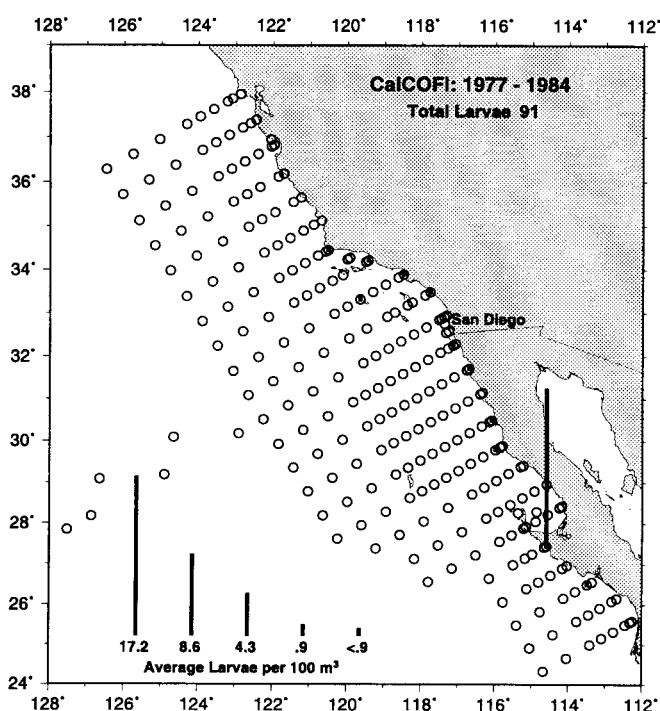
GOBIIDAE



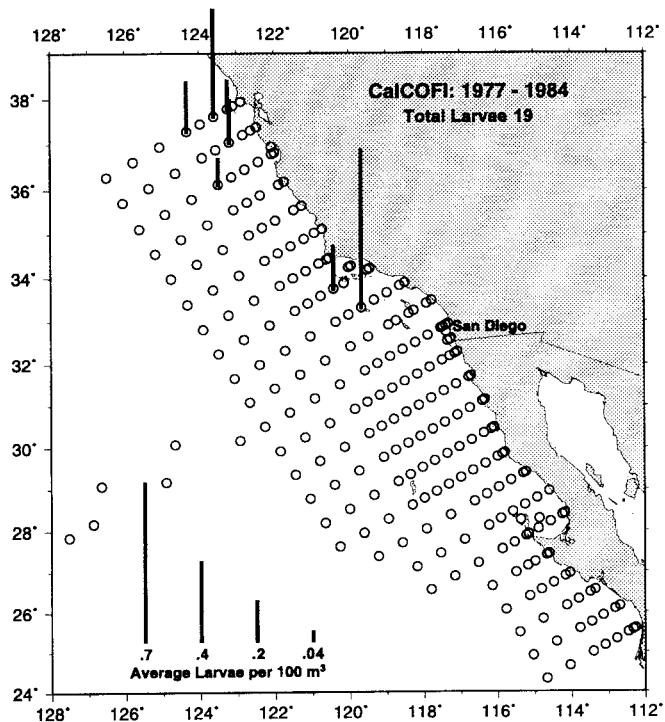
CHAENOPSIDAE

Yellowfin fringehead

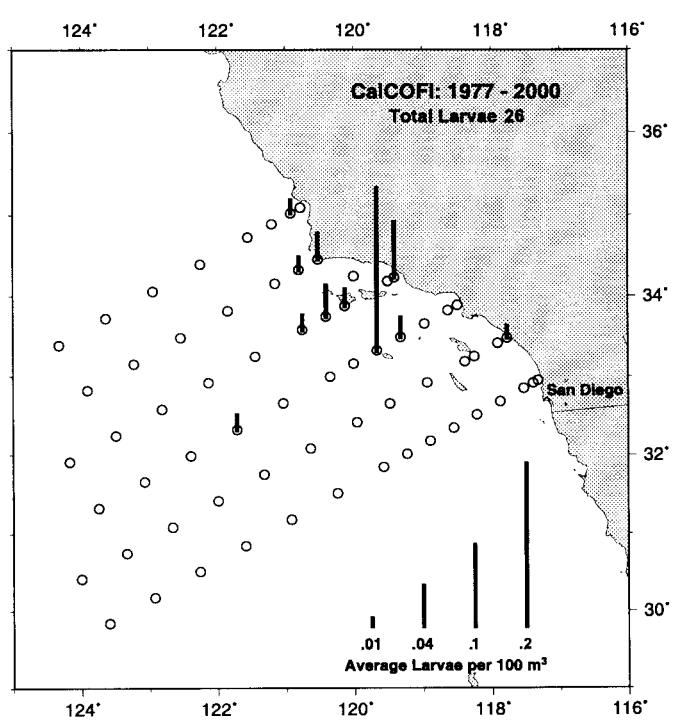
Neoclinus stephensae



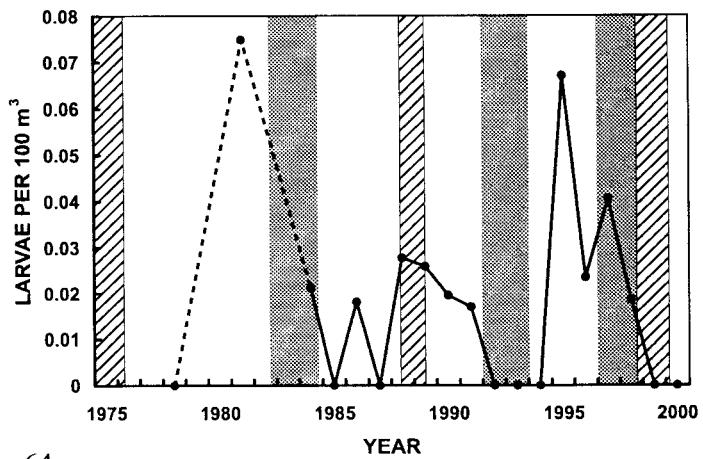
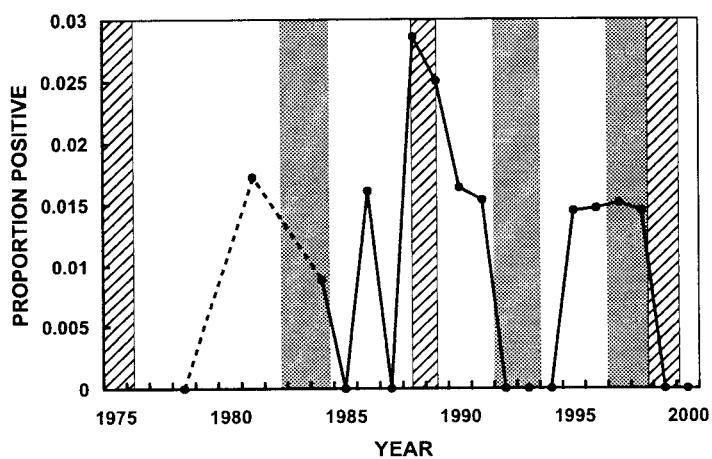
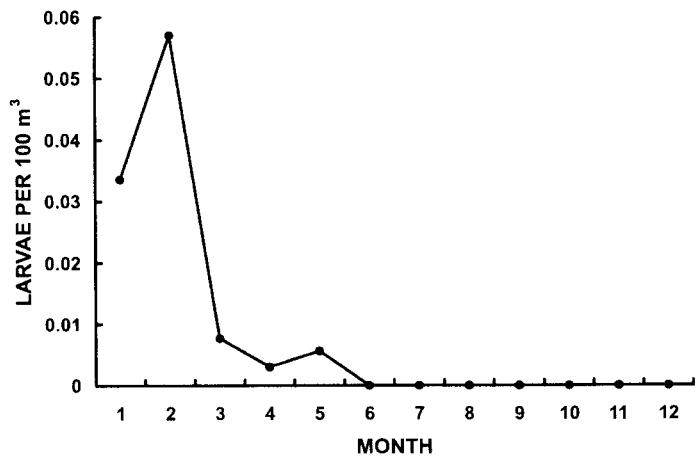
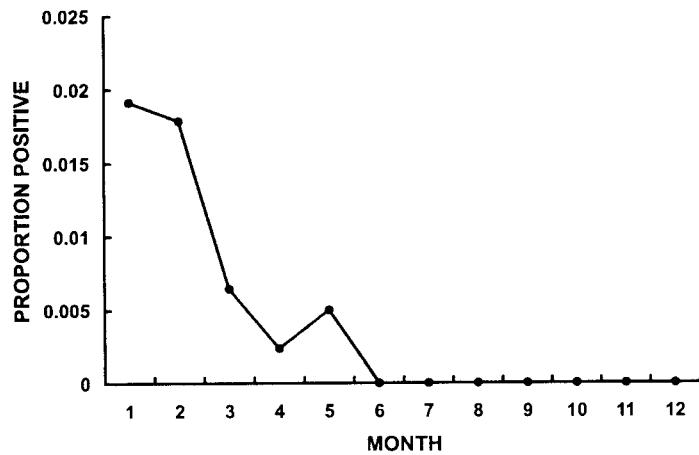
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Bocaccio



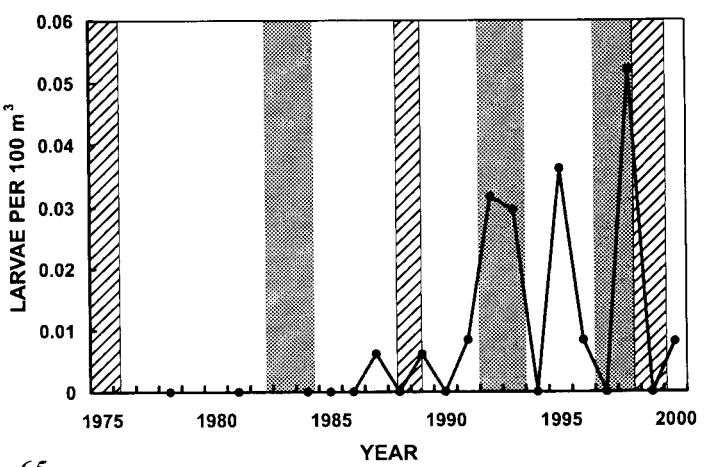
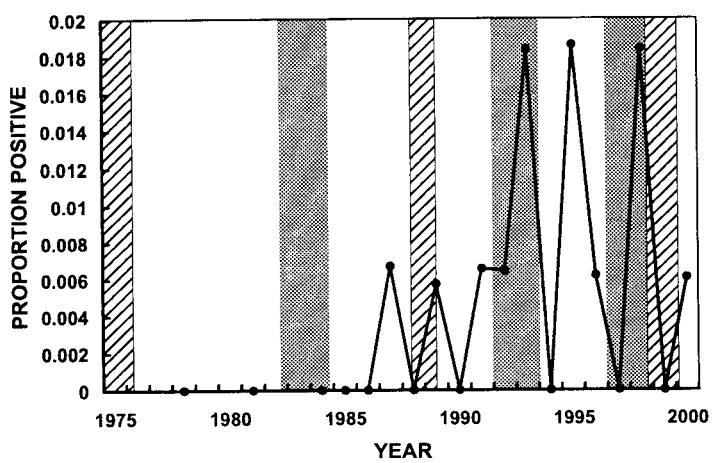
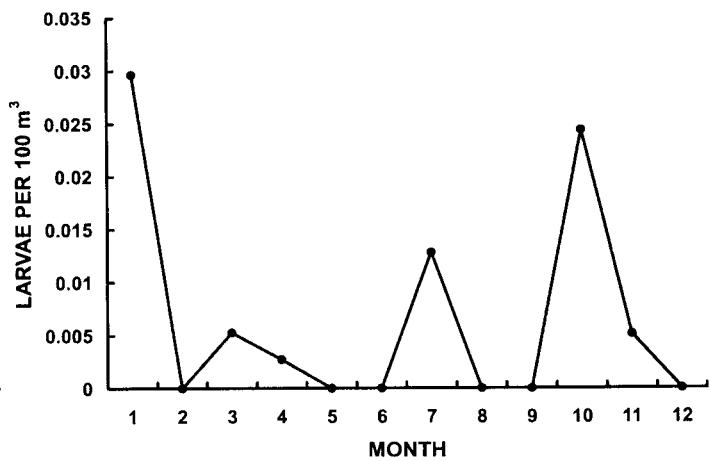
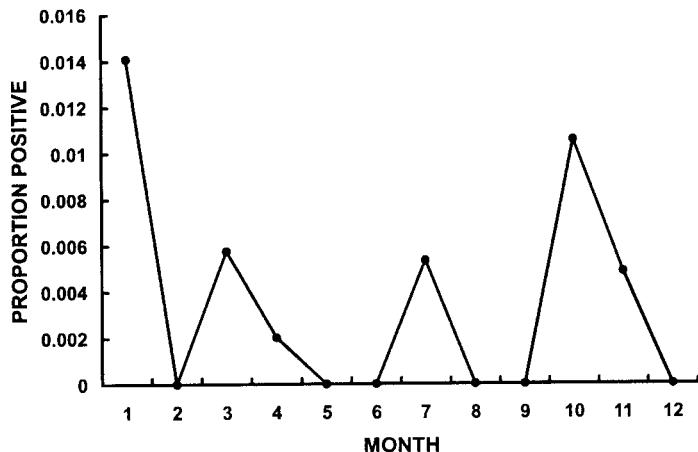
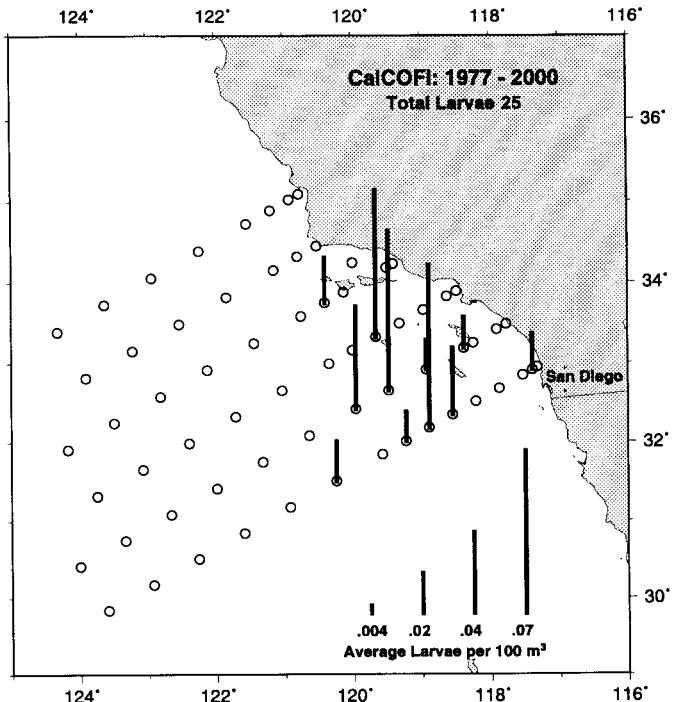
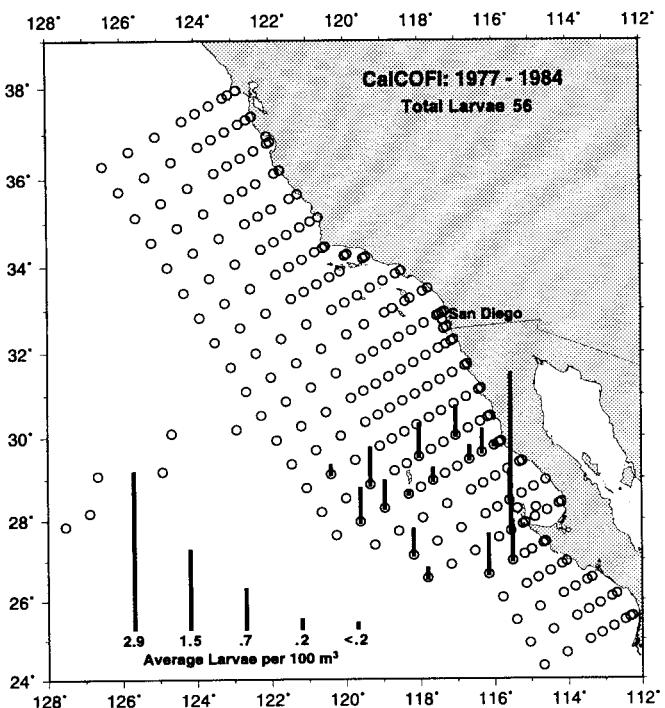
SEBASTIDAE



CENTRISCIDAE

Snipefish

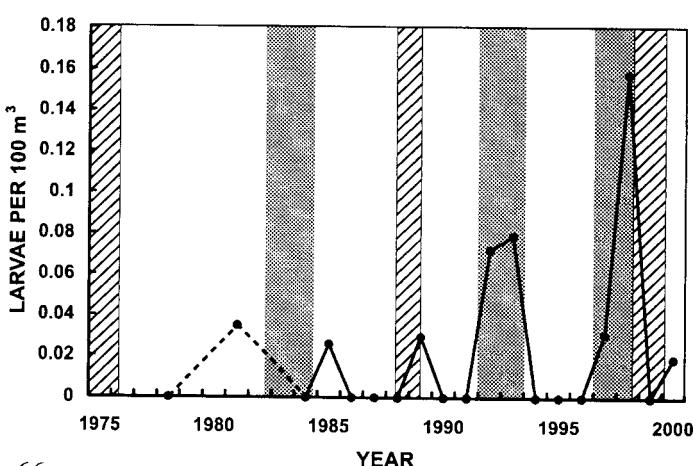
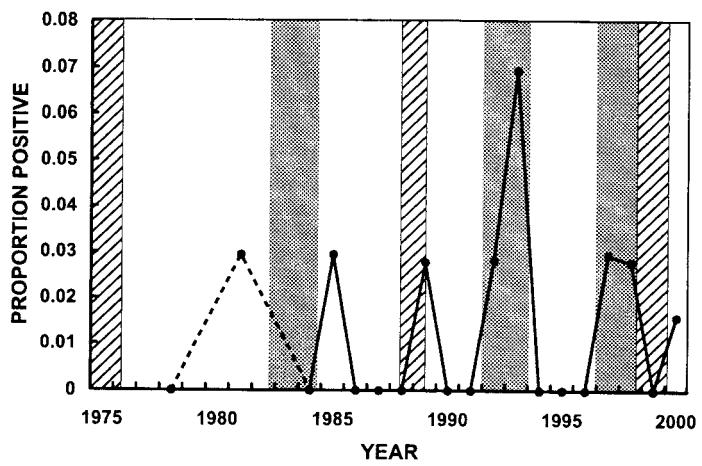
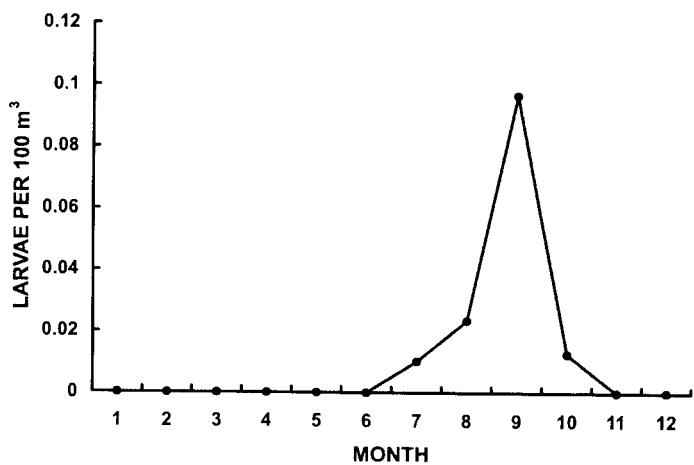
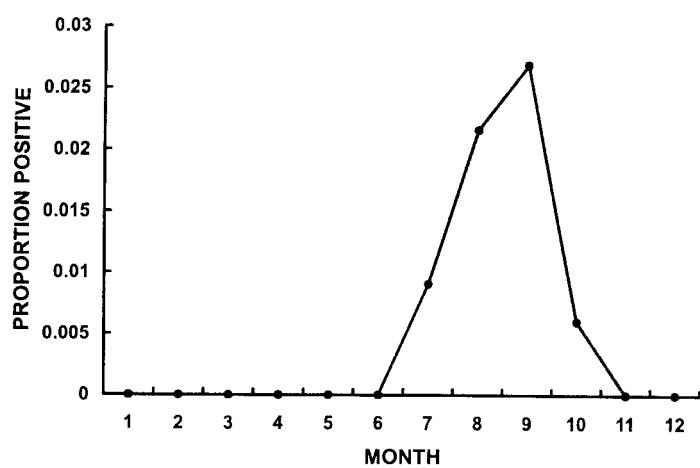
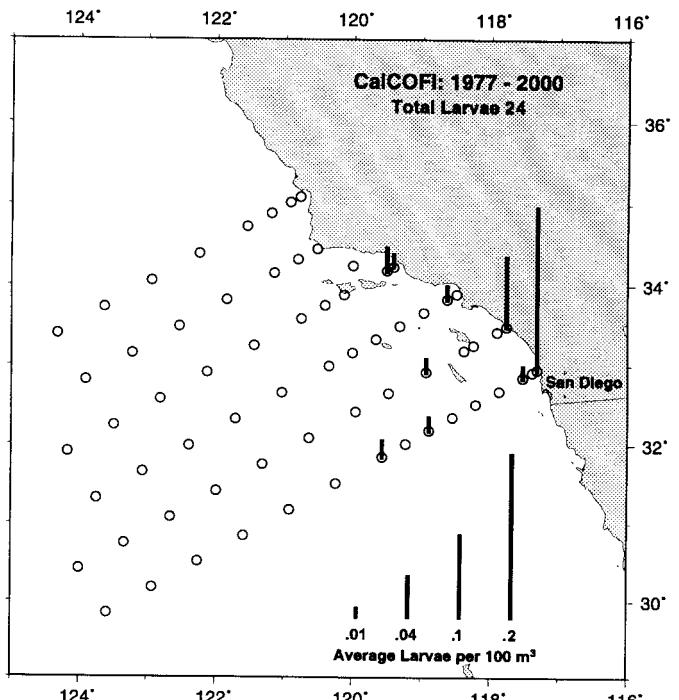
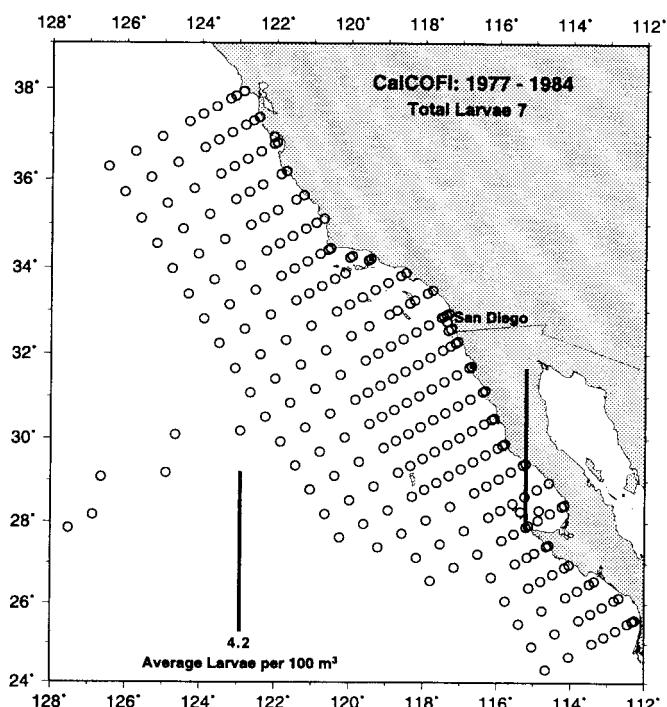
Macroramphosus gracilis



Fodiator acutus

Sharpchin flyingfish

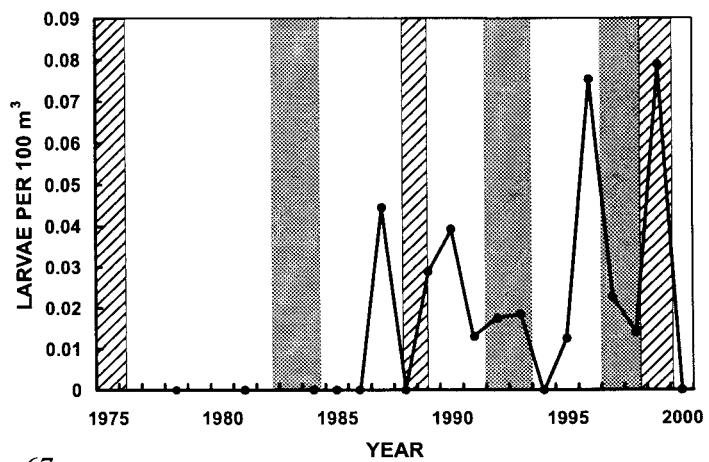
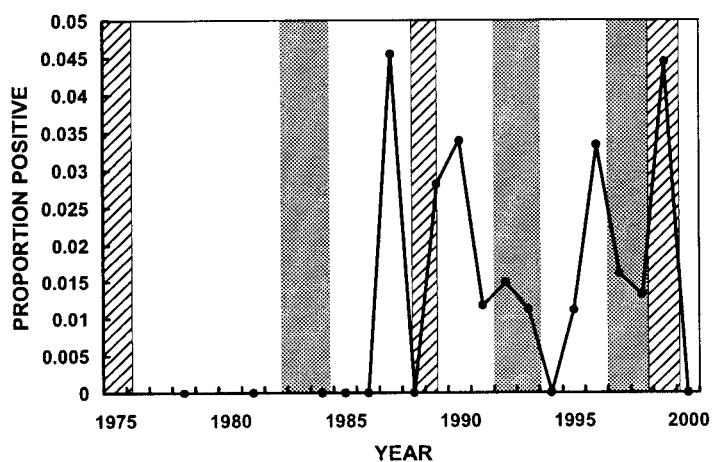
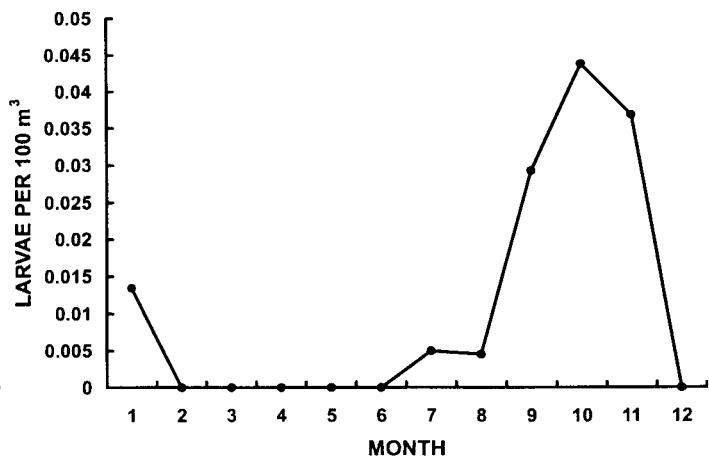
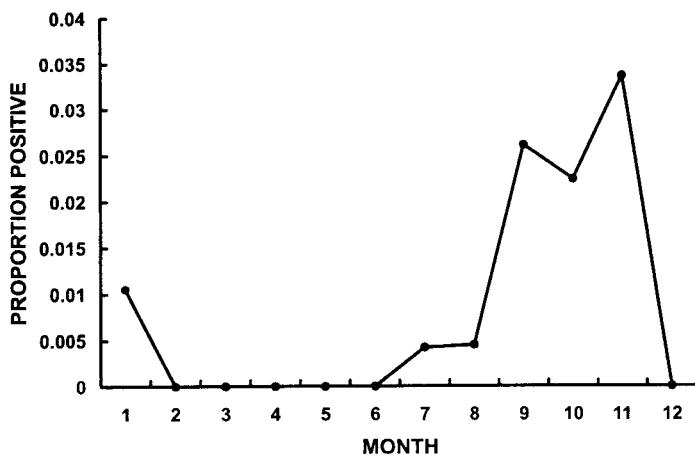
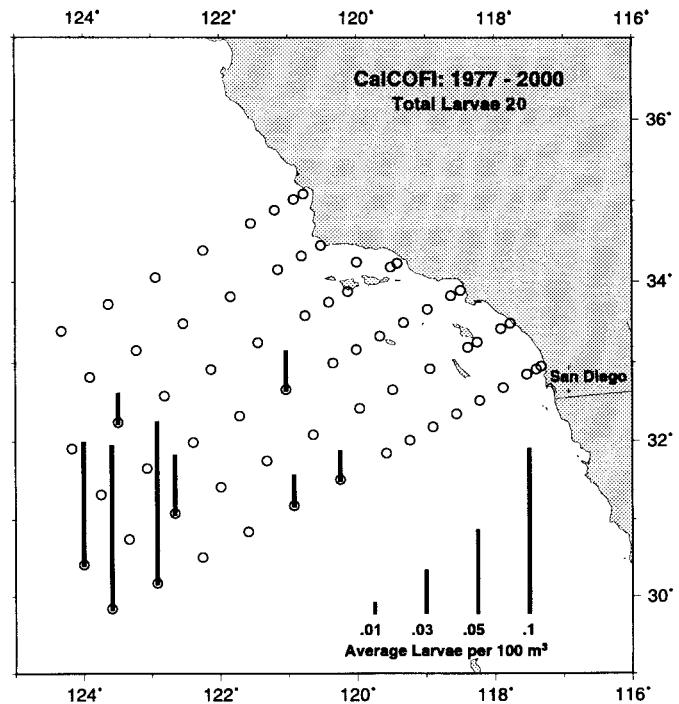
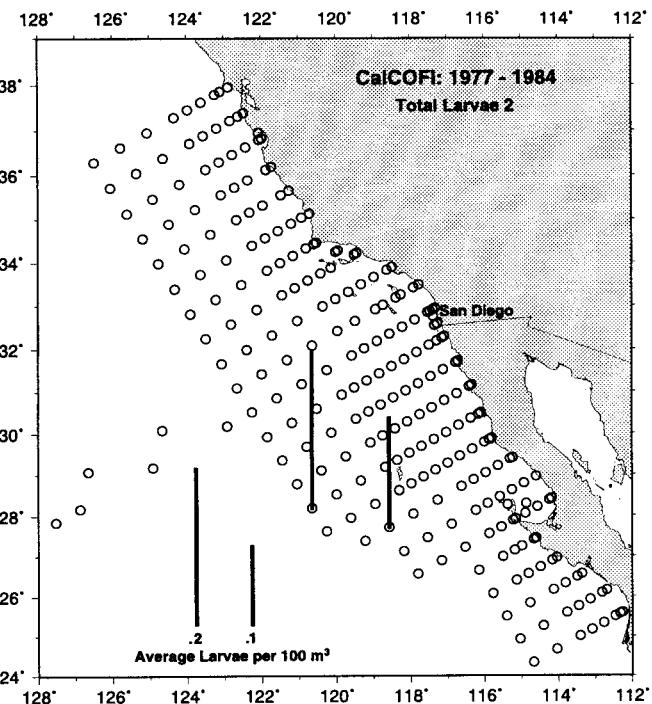
EXOCOETIDAE

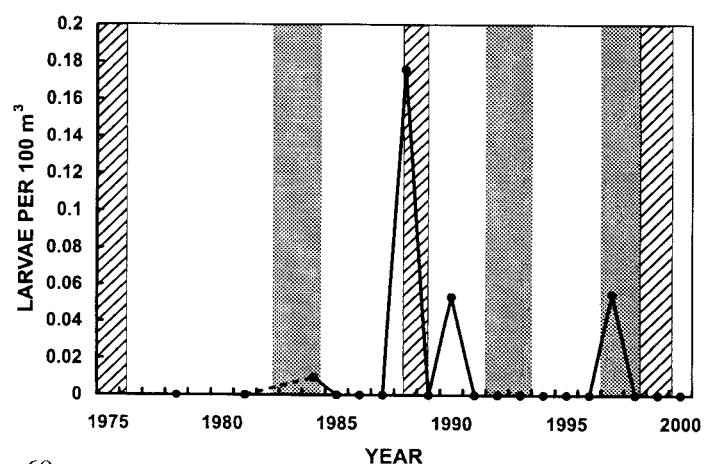
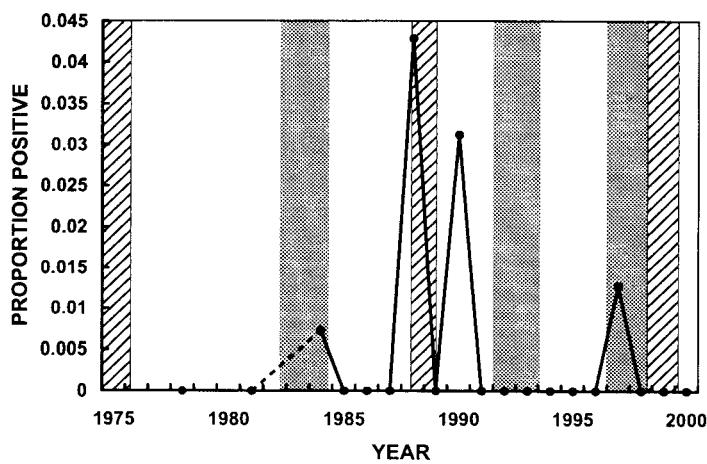
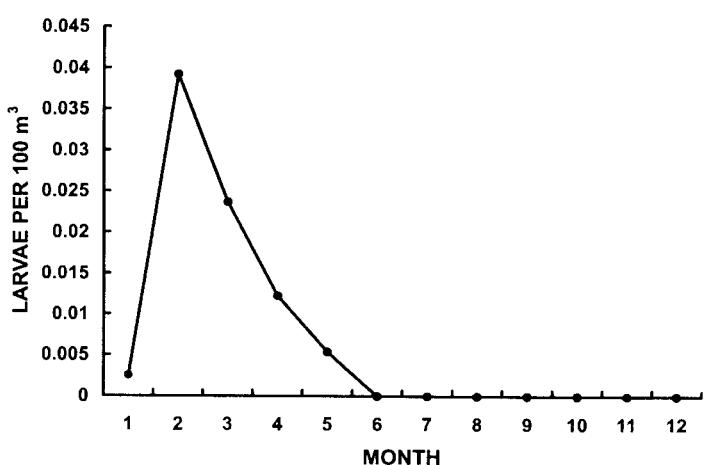
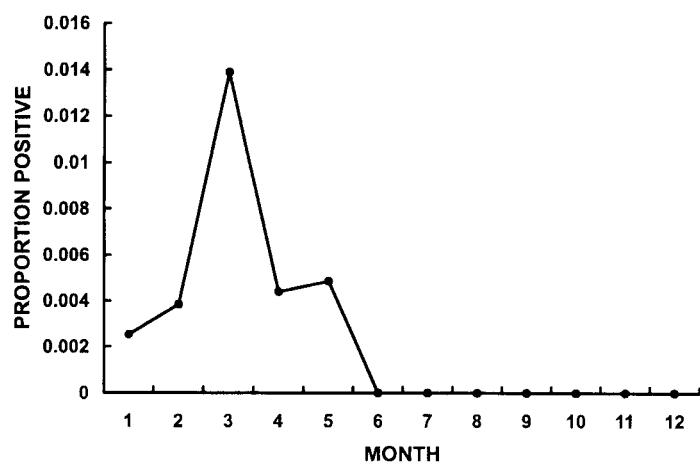
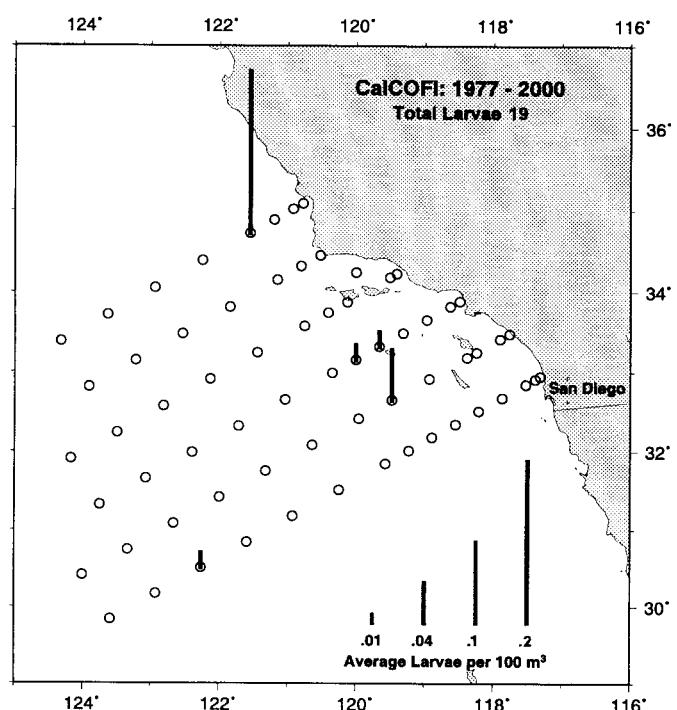
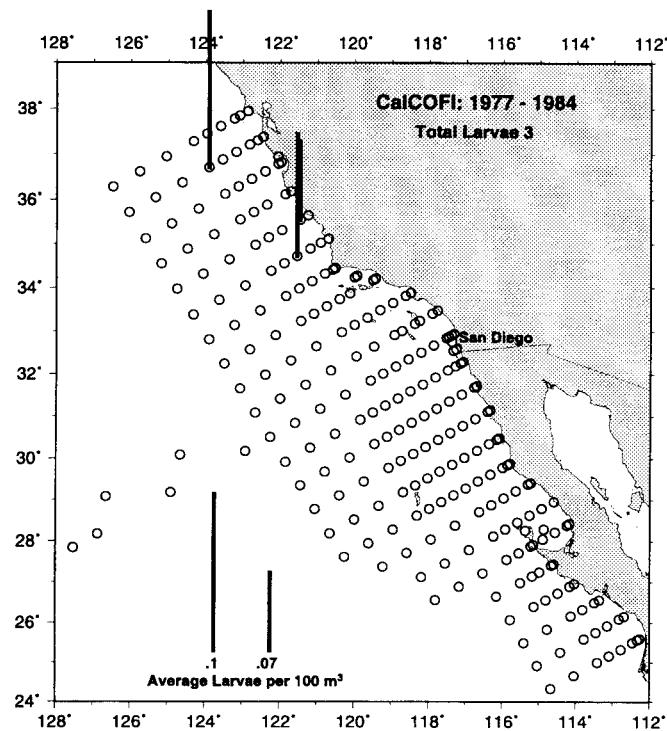


GIGANTACTINIDAE

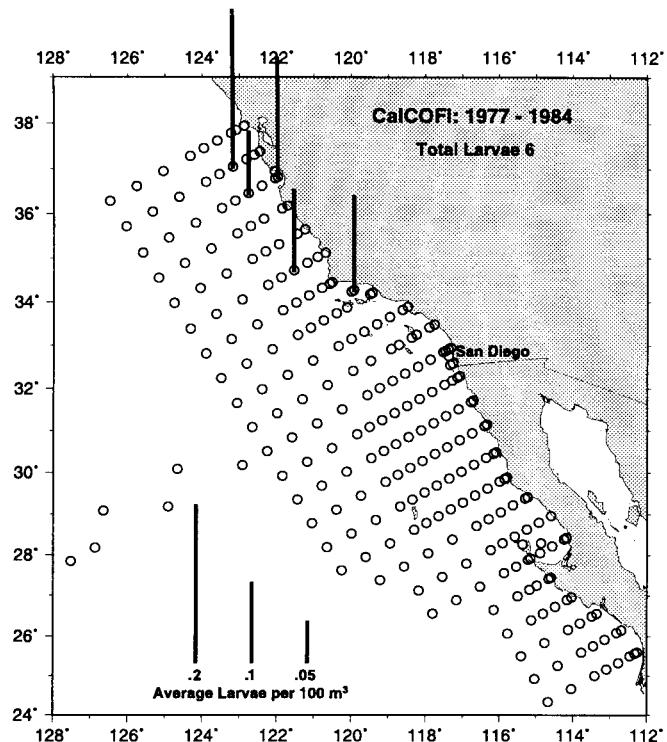
Whipnoses

Gigantactis spp.

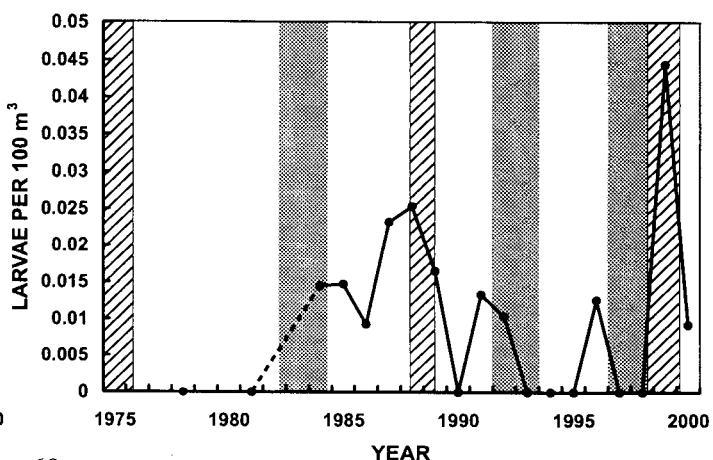
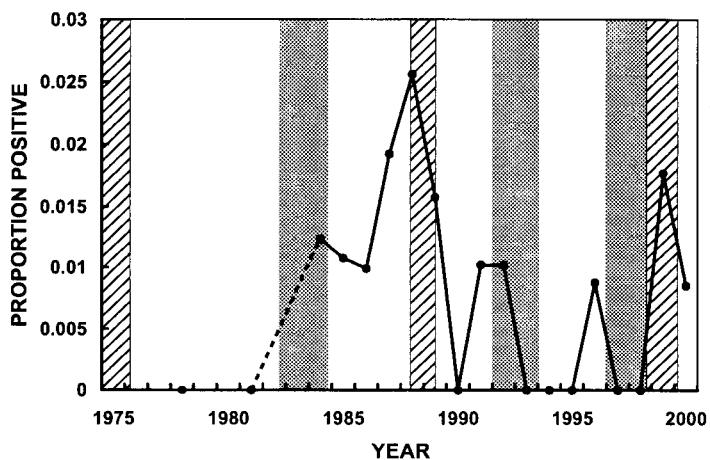
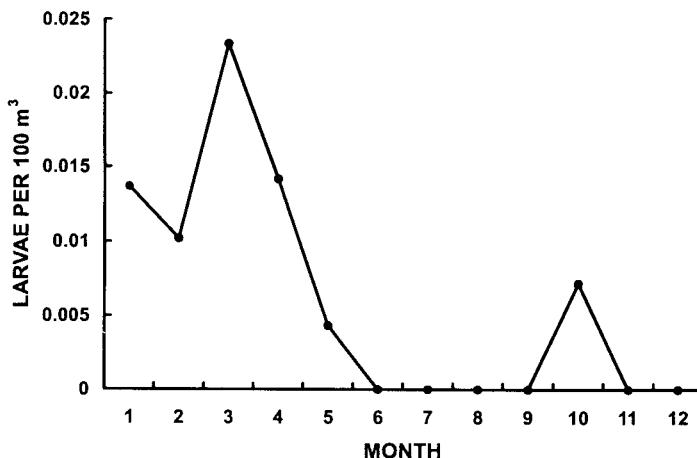
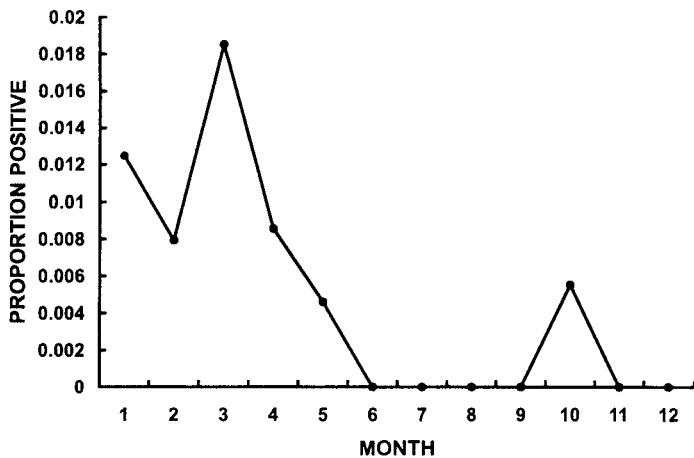
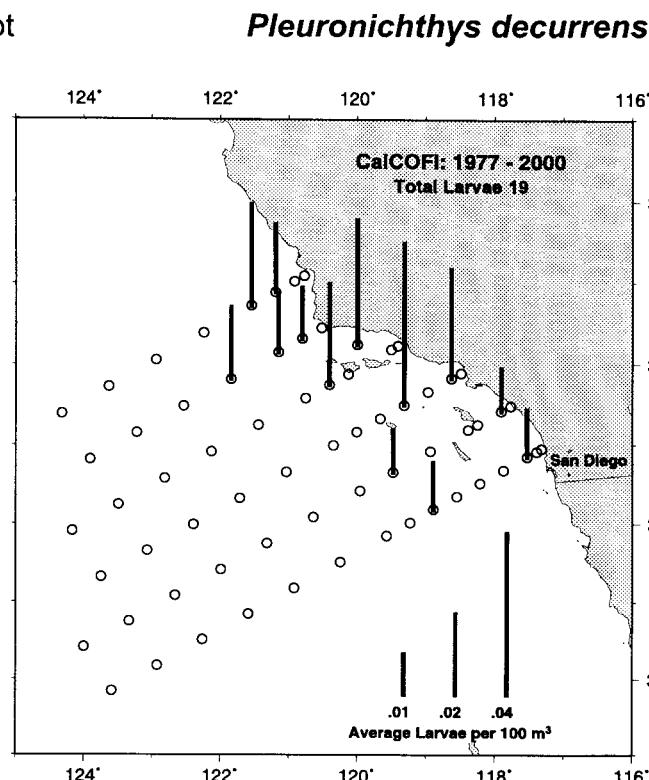


Bathylags ochotensis**Popeye blacksmelt****BATHYLAGIDAE**

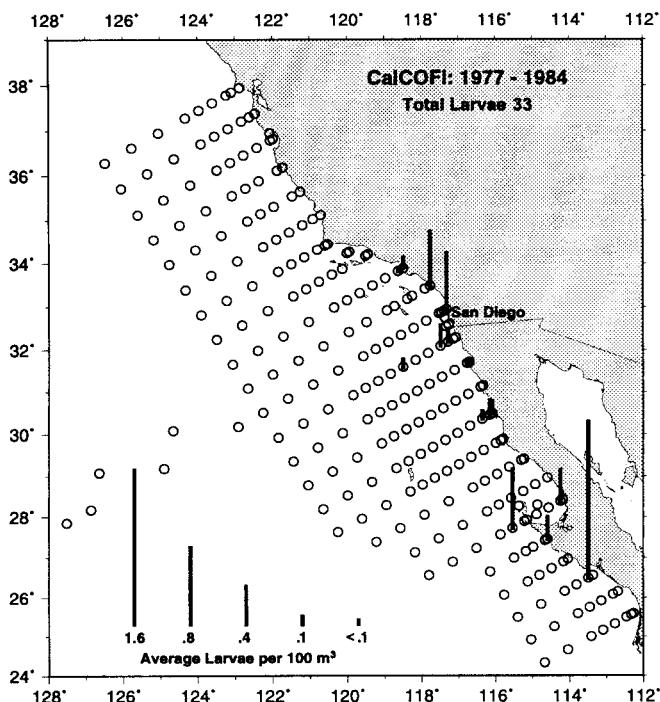
PLEURONECTIDAE



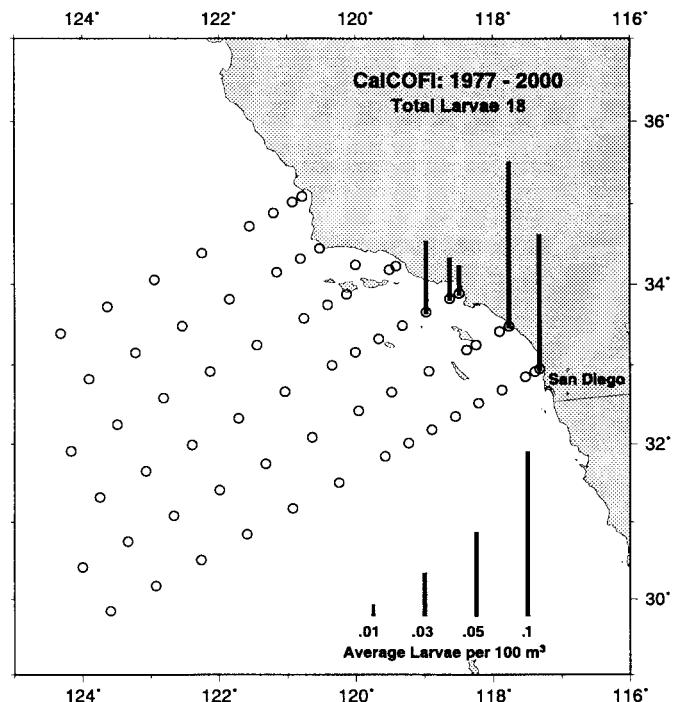
Curlfin turbot



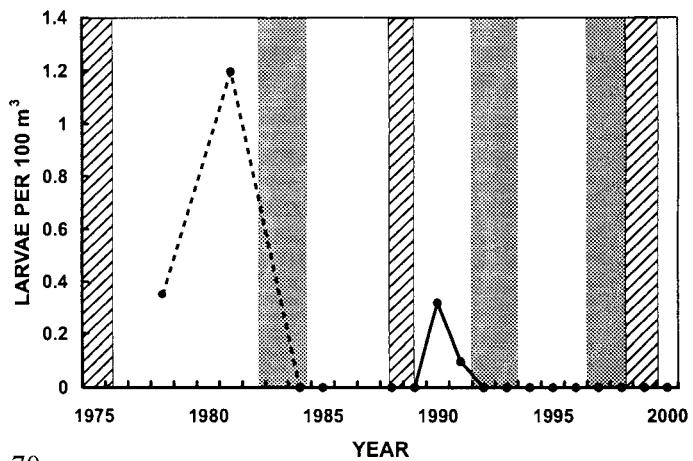
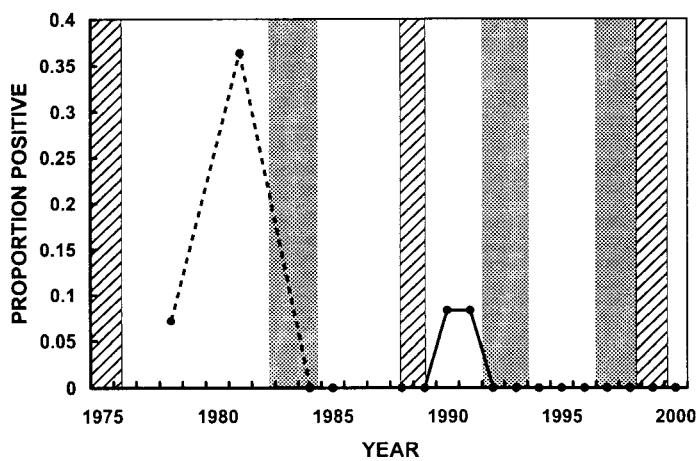
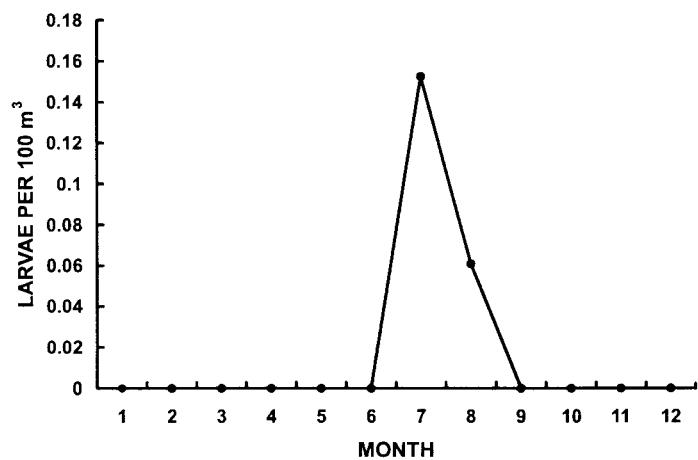
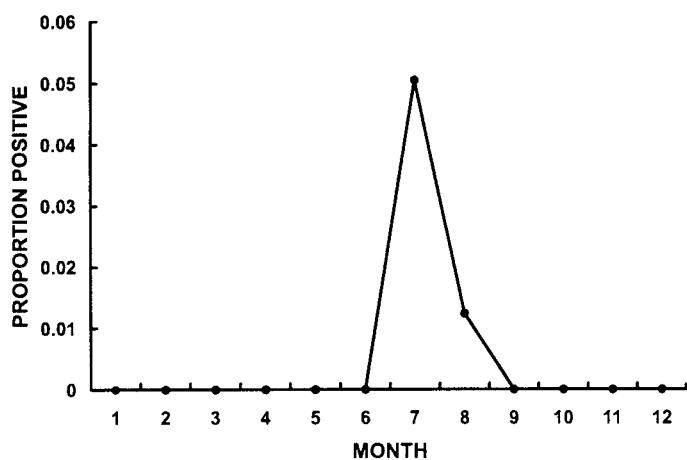
Hermosilla azurea



Zebraperch



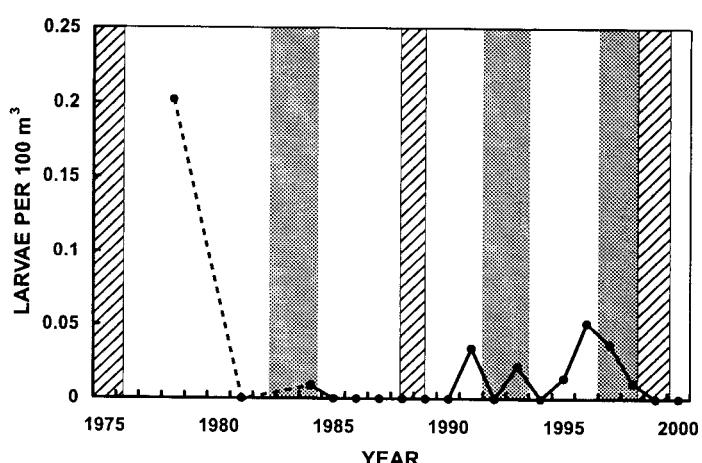
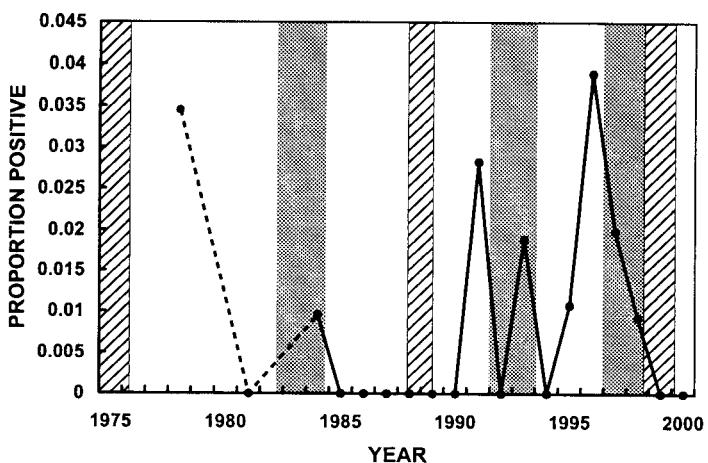
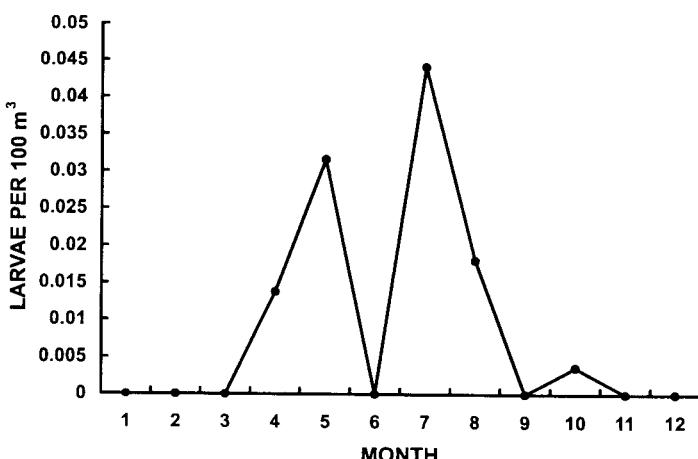
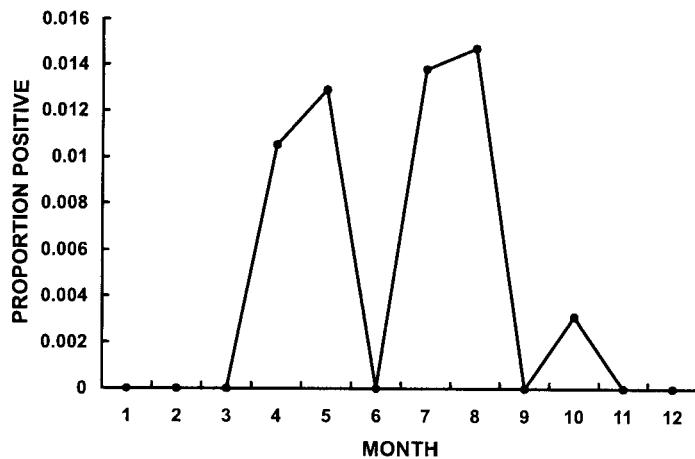
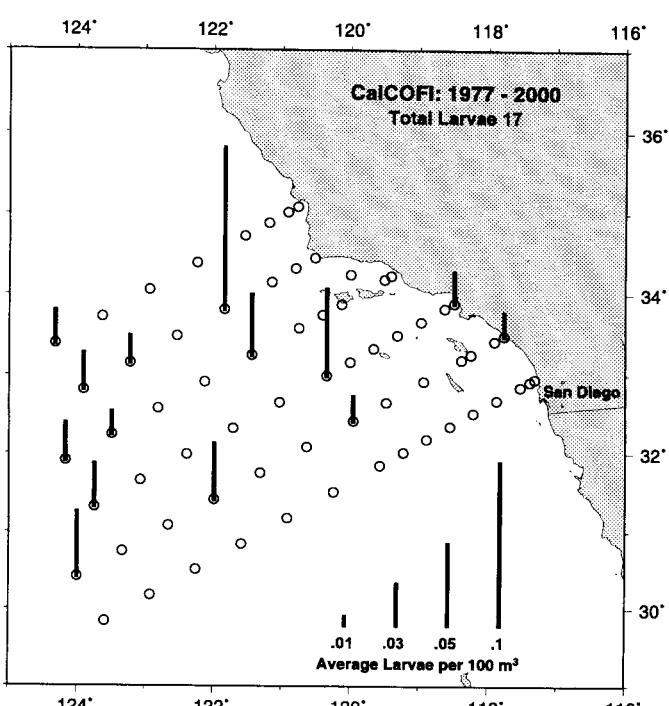
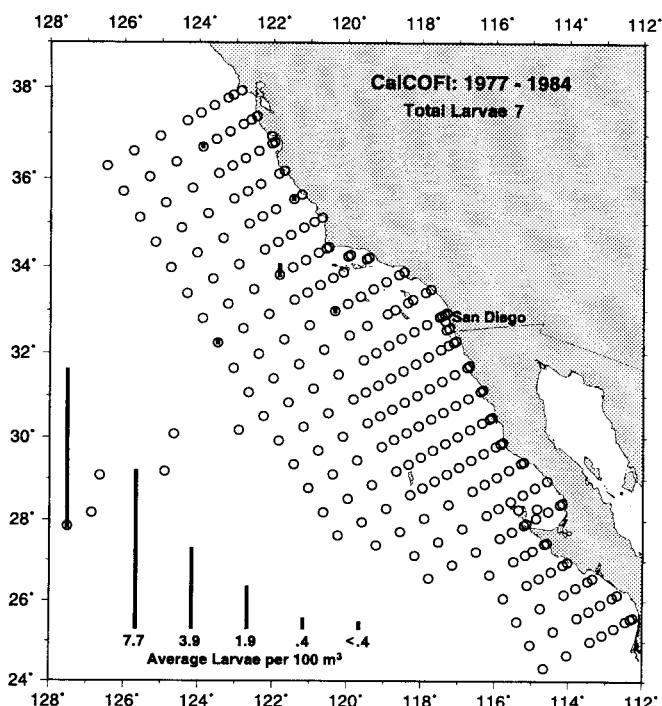
KYPHOSIDAE



MYCTOPHIDAE

Headlightfishes

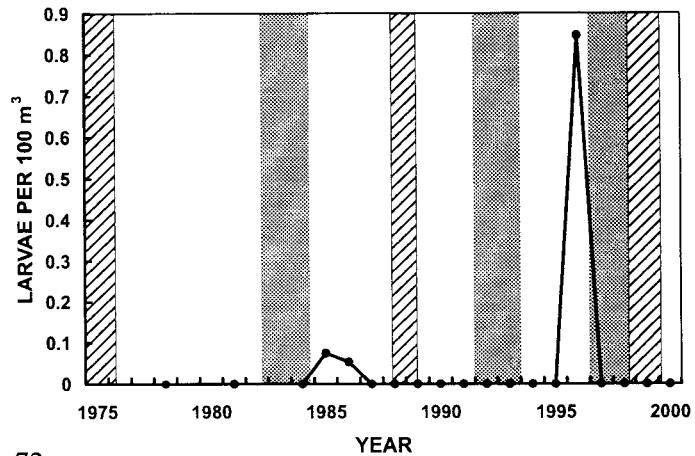
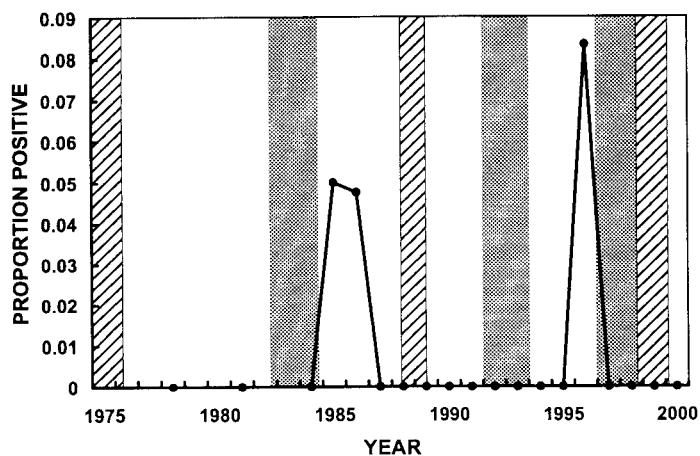
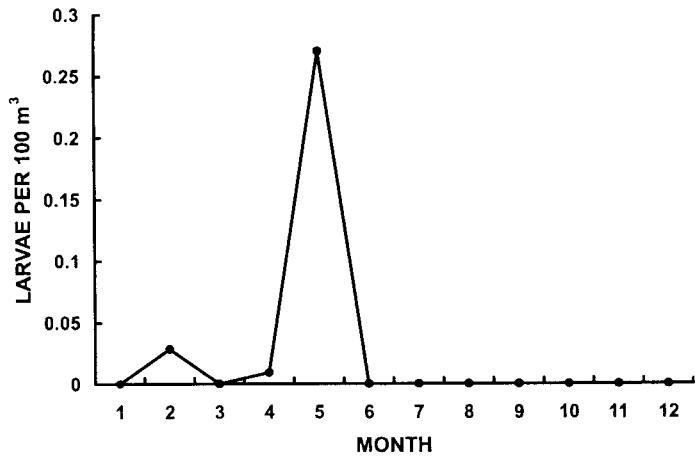
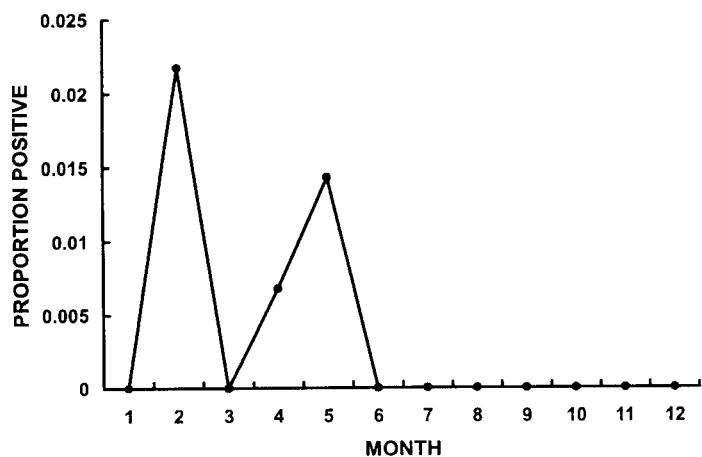
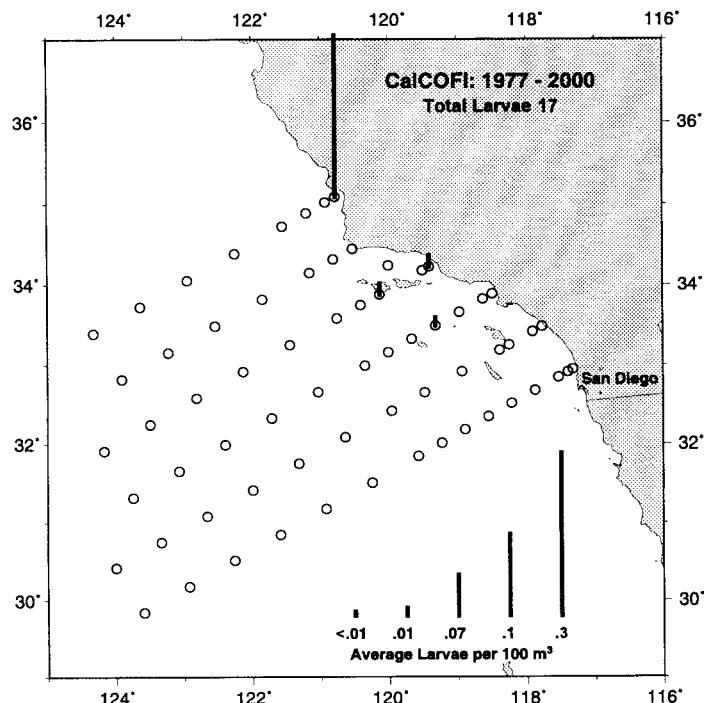
Diaphus spp.



Rathbunella allenii

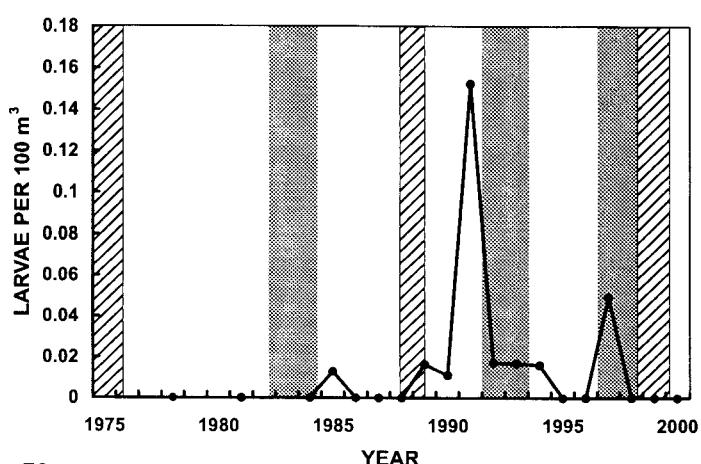
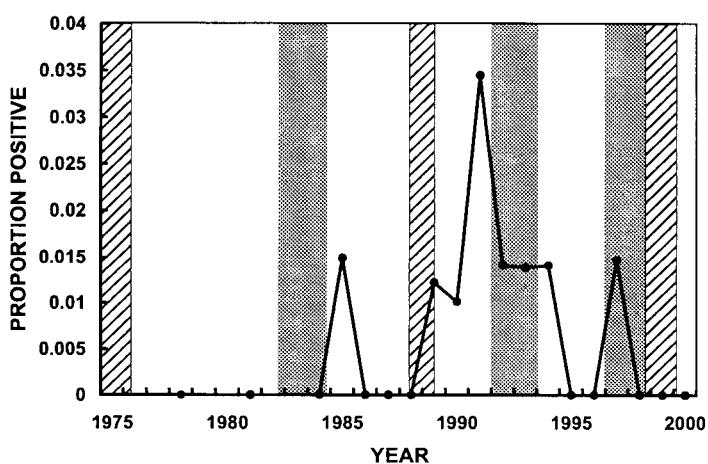
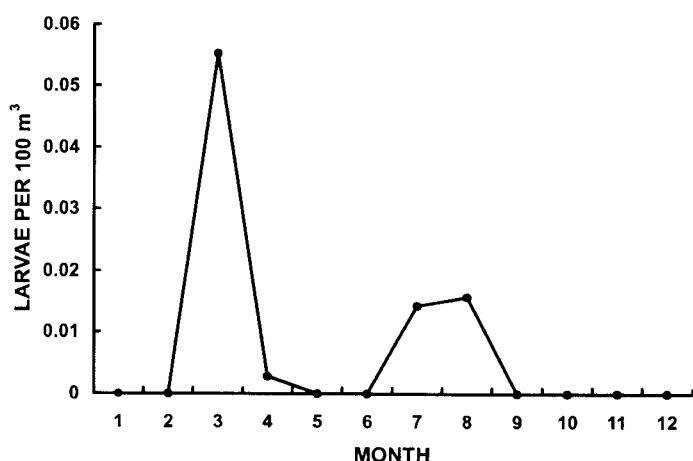
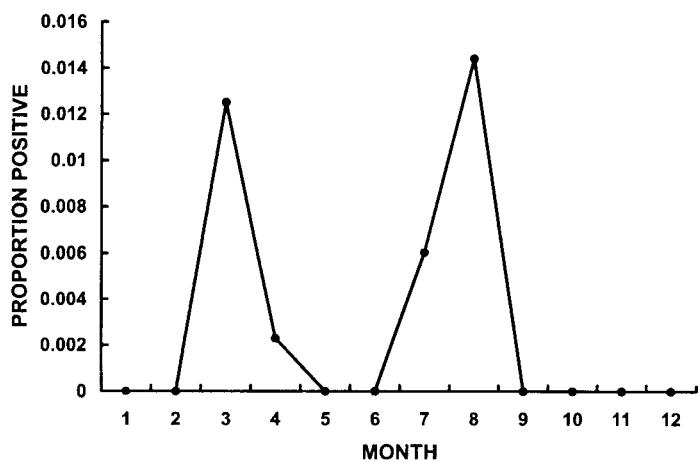
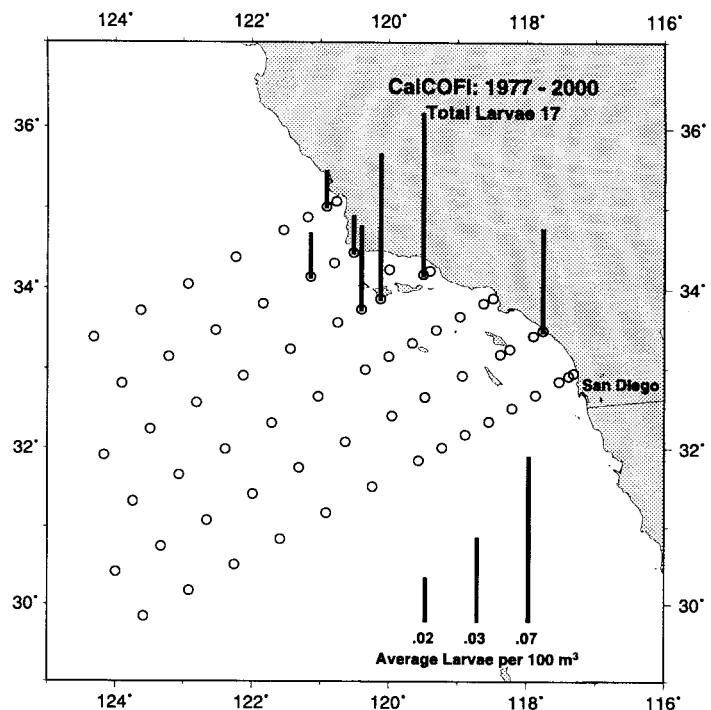
Rough ronquil

BATHYMASTERIDAE

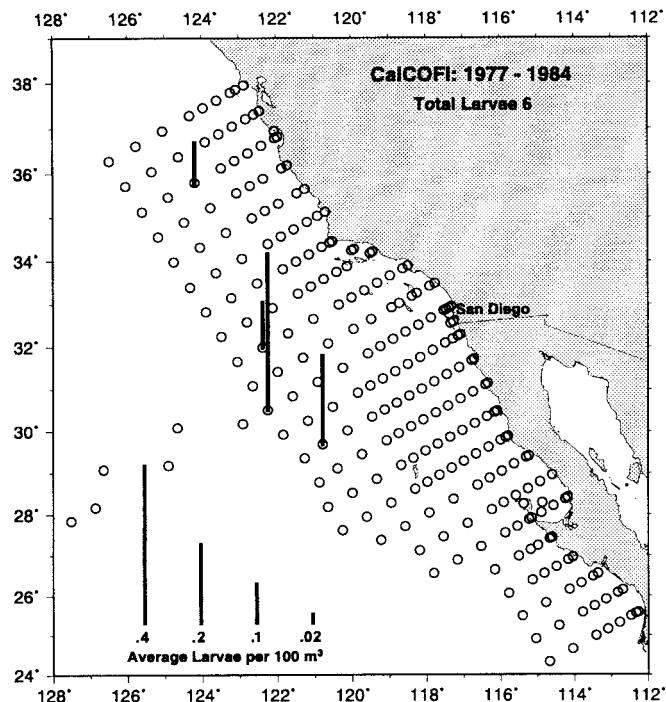


OPHIDIIDAE

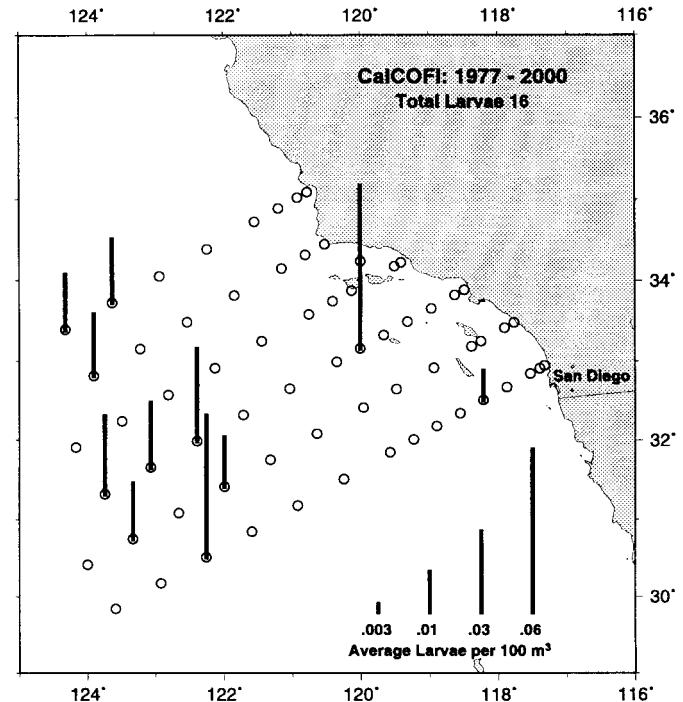
Basketweave cusk-eel

Ophidion scrippsae

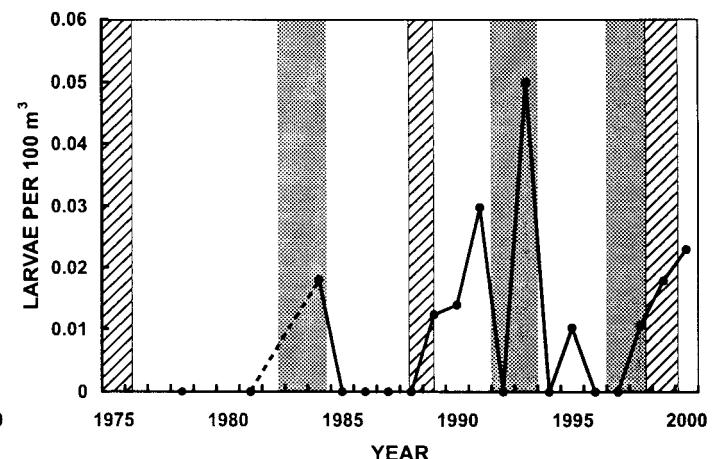
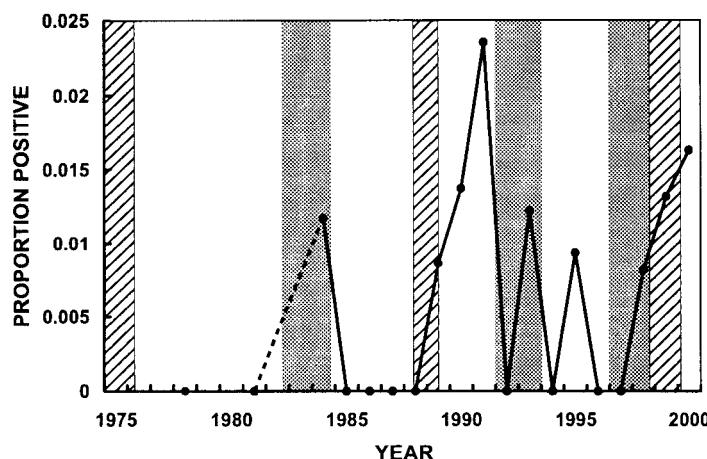
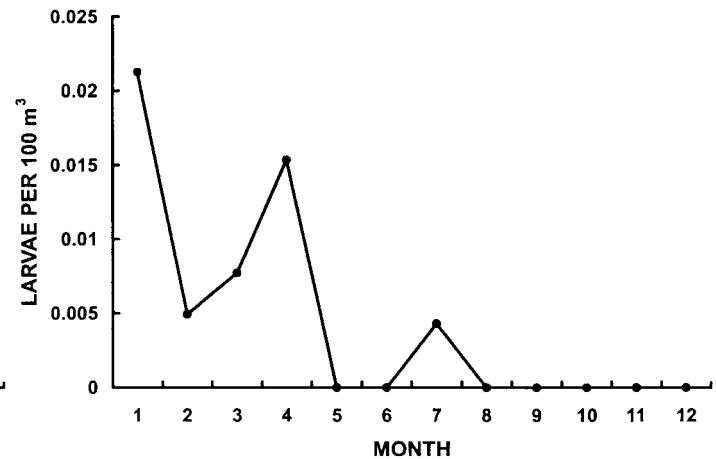
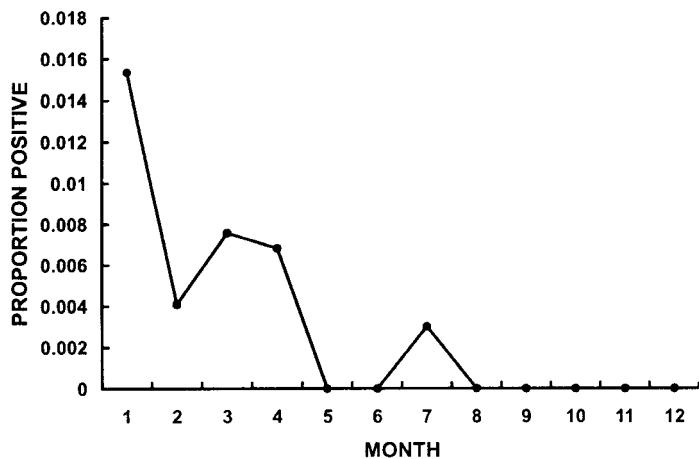
Sybolophorus californiensis



California lanternfish



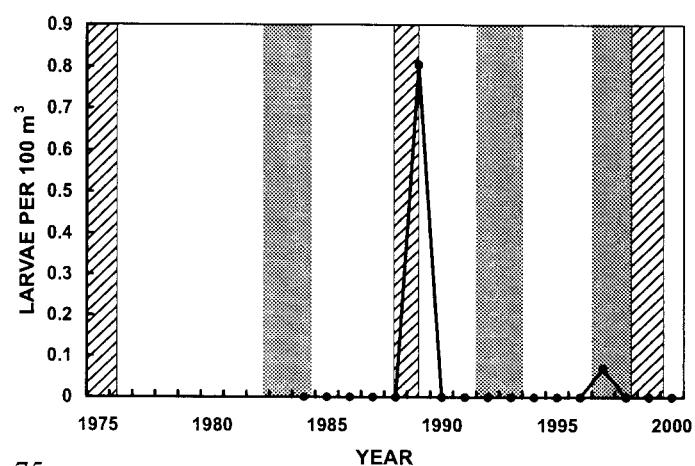
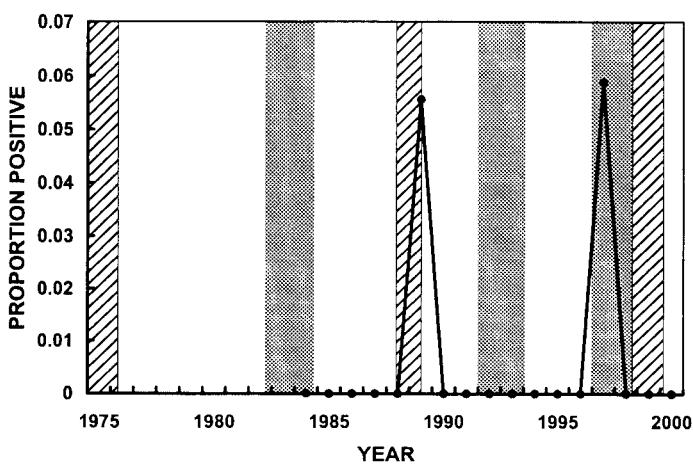
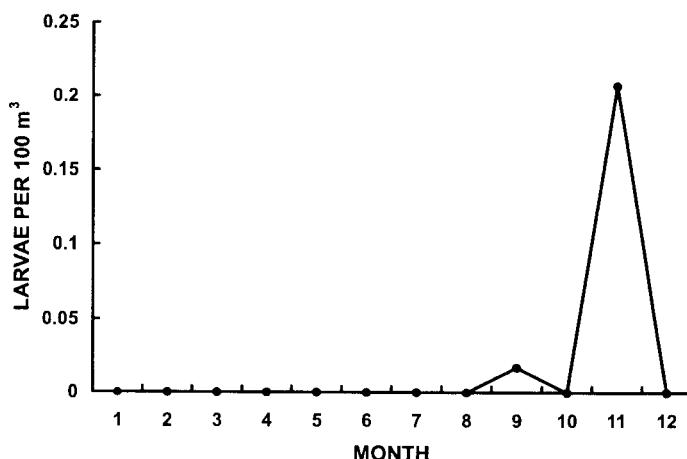
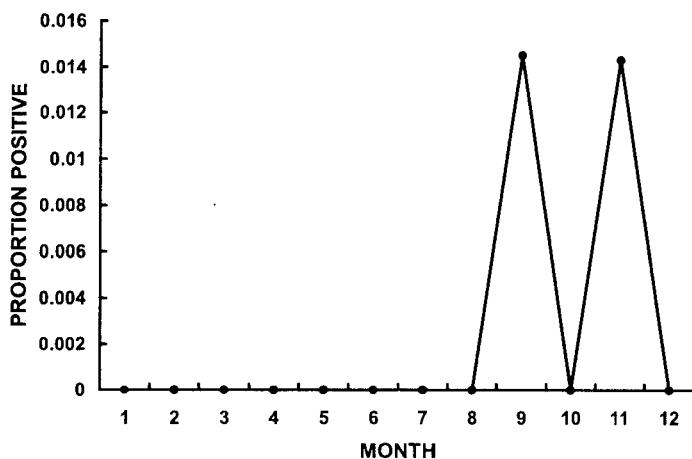
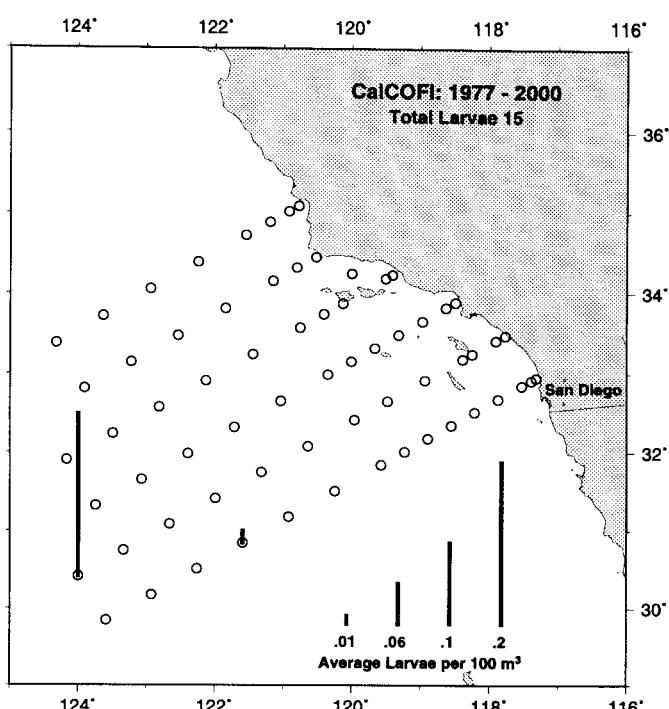
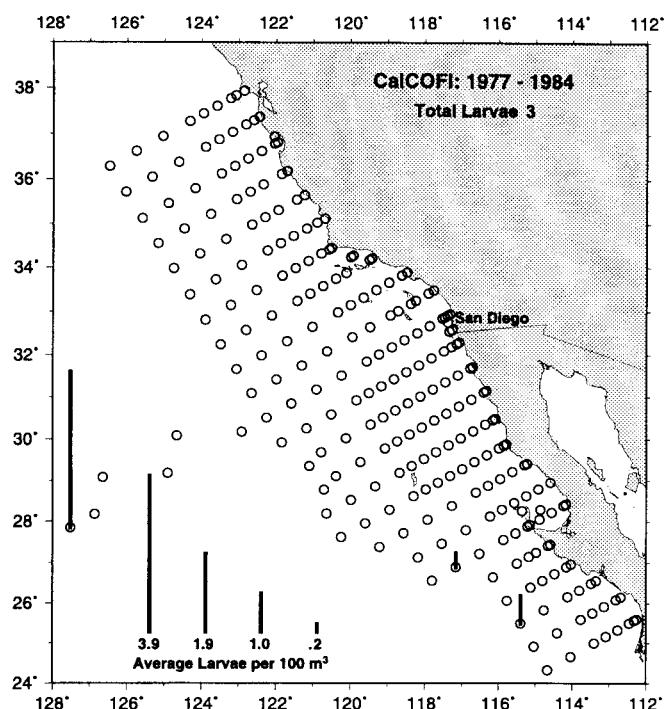
MYCTOPHIDAE



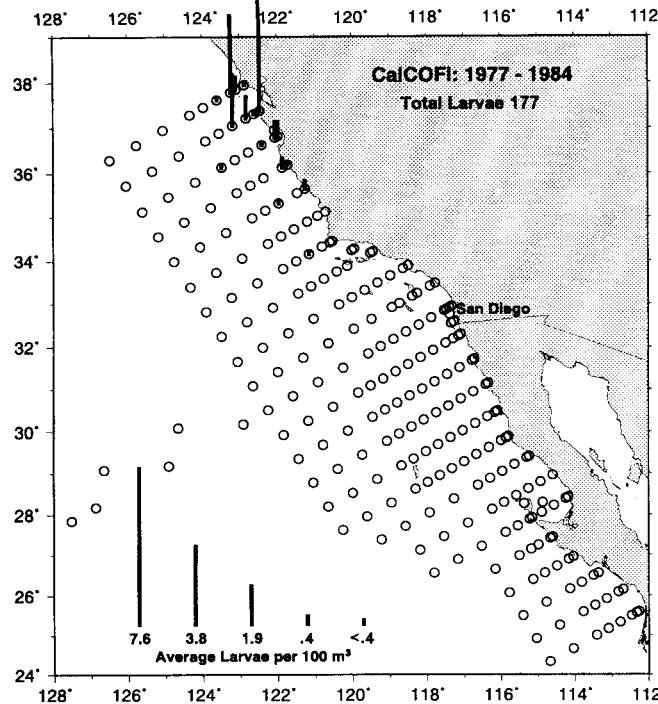
MYCTOPHIDAE

Popeye lampfish

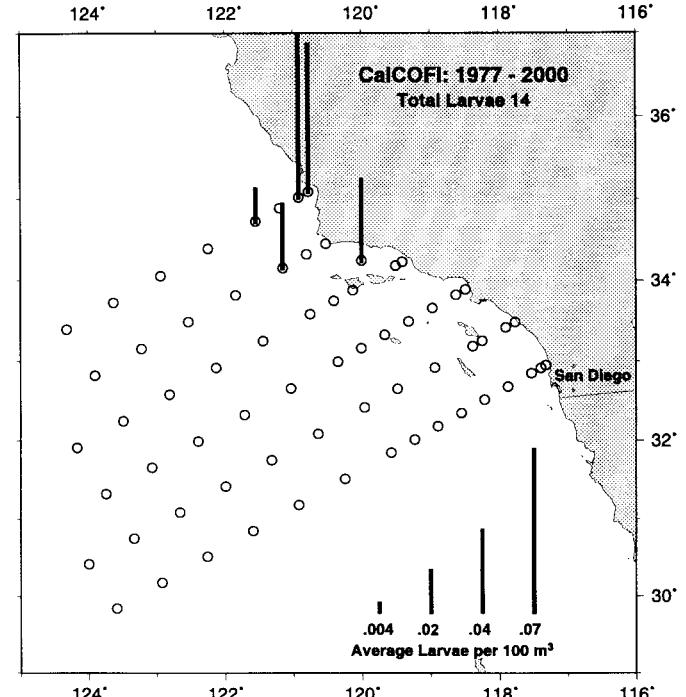
Bolinichthys longipes



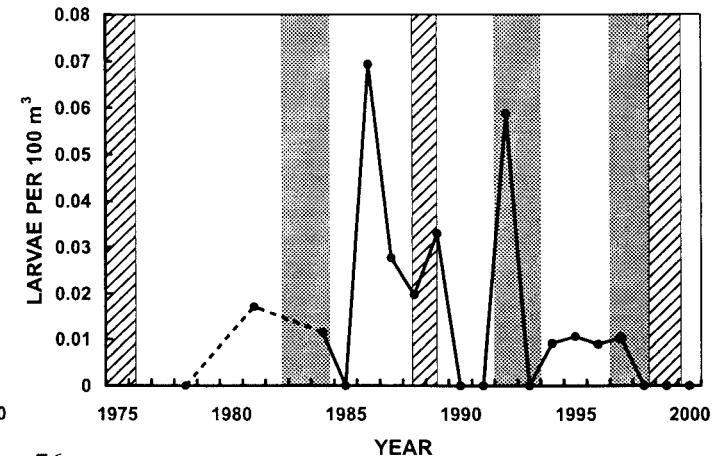
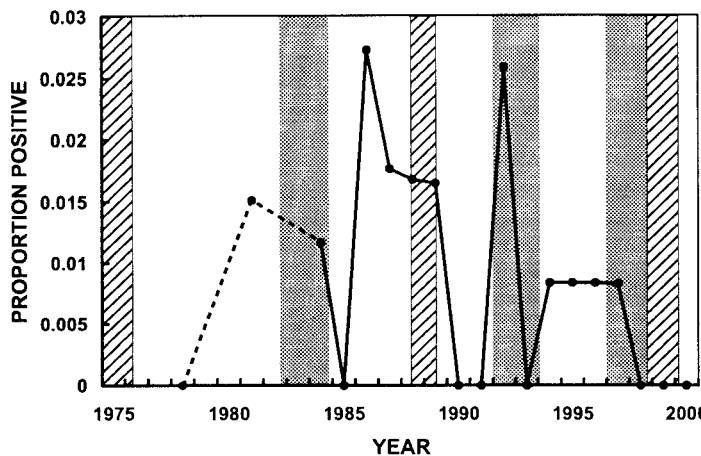
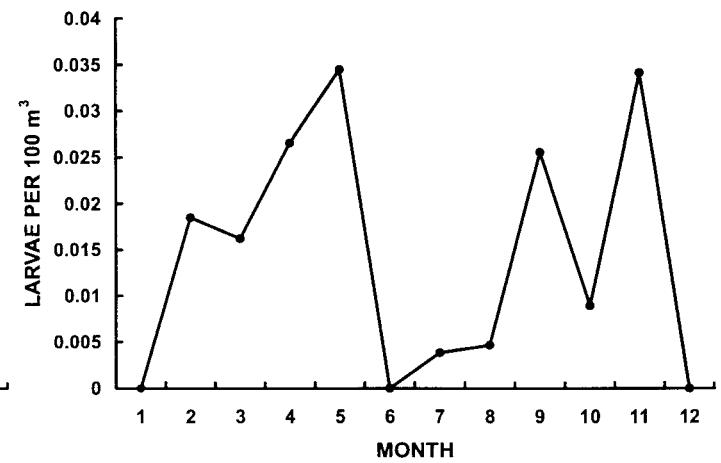
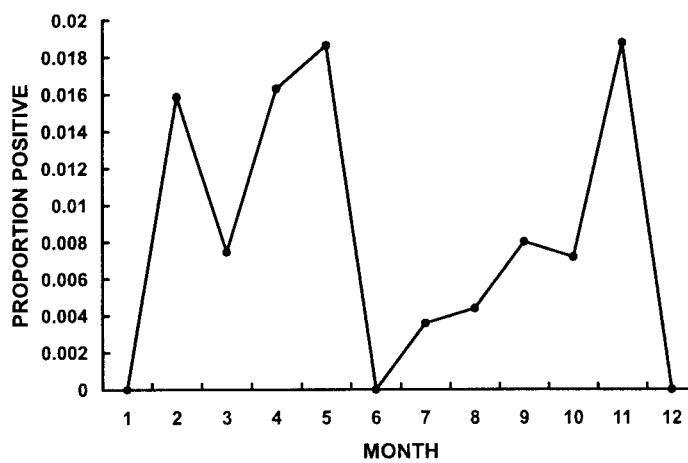
Hemilepidotus spinosus



Brown Irish Lord



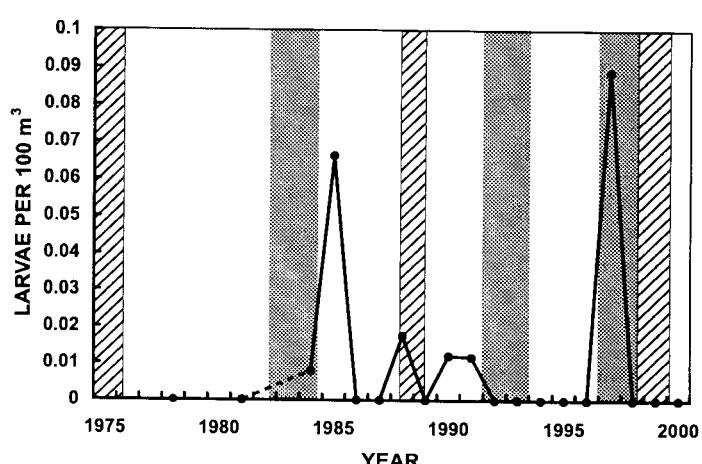
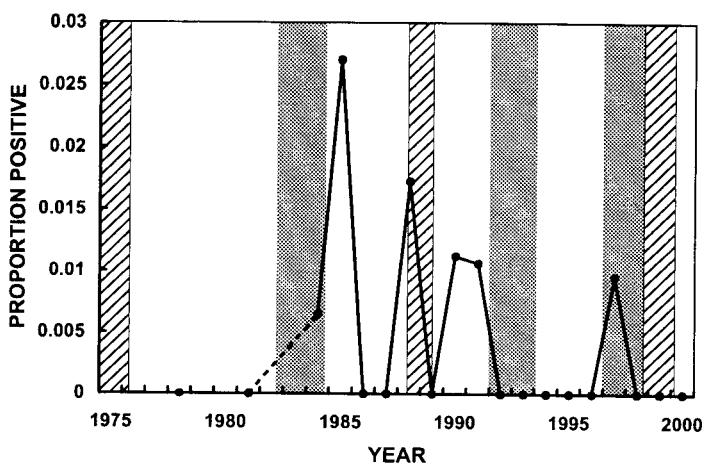
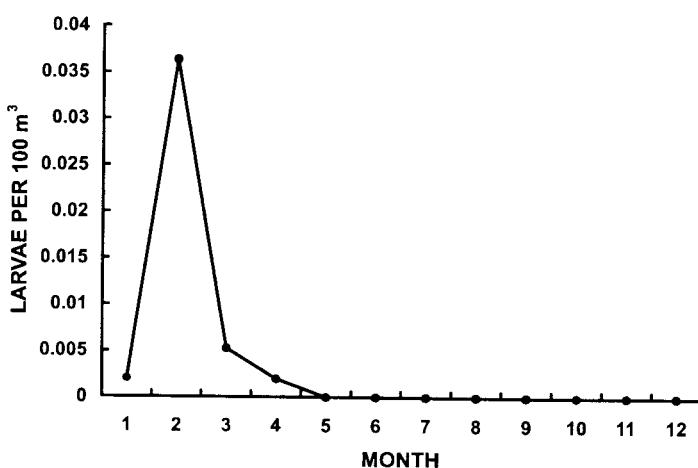
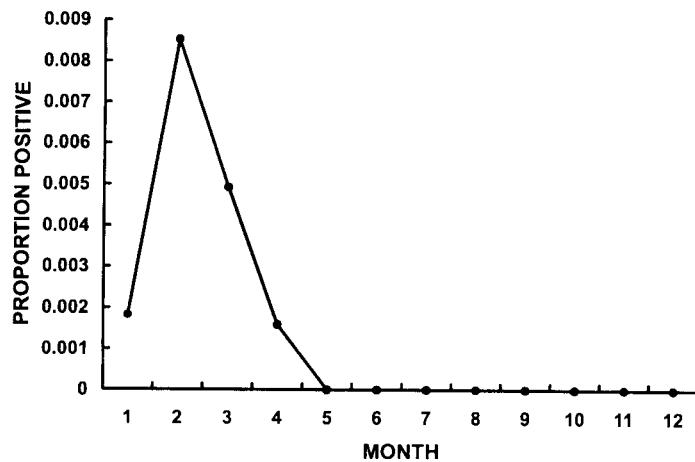
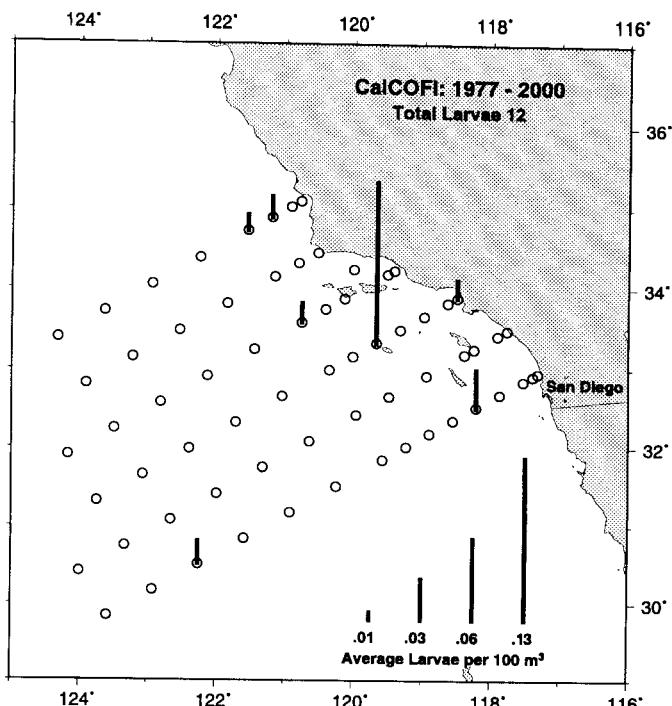
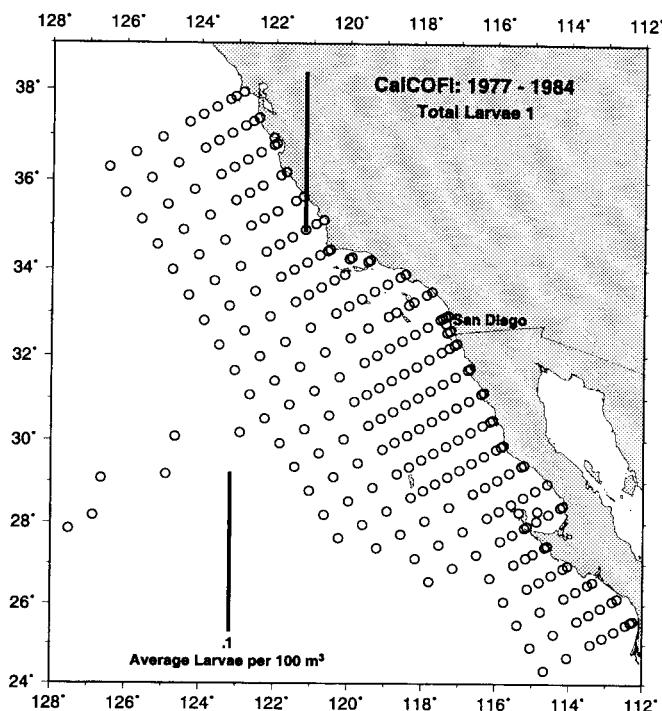
COTTIDAE



BATHYLAGIDAE

California smoothtongue

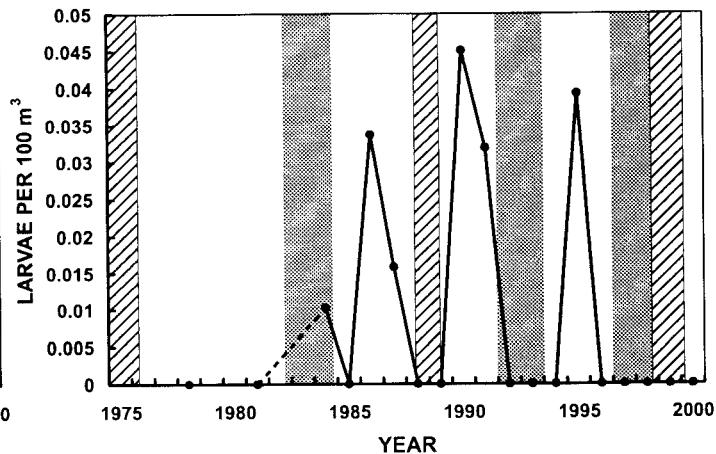
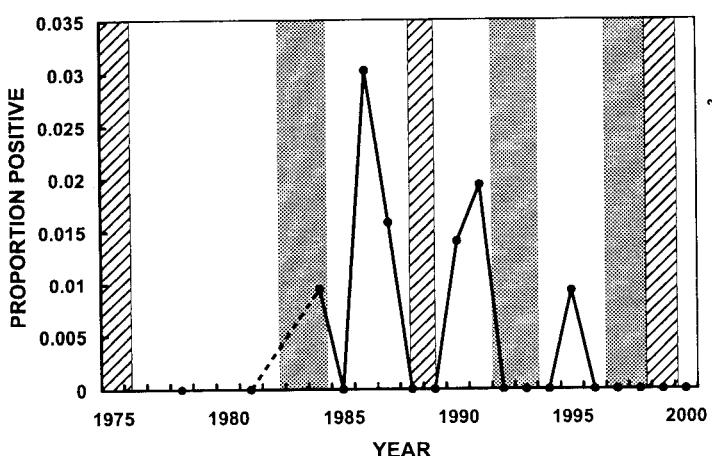
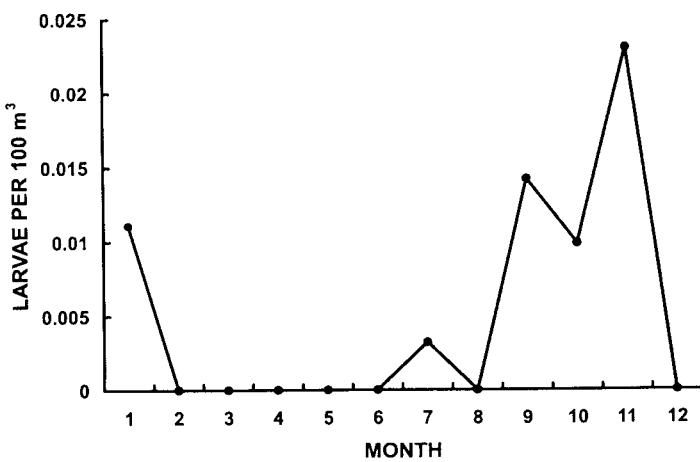
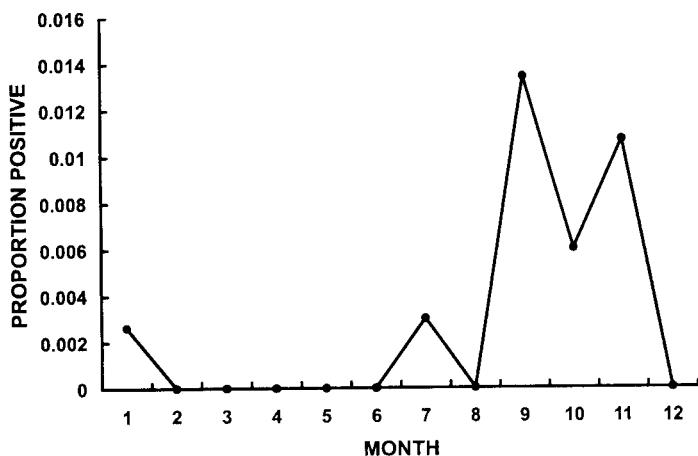
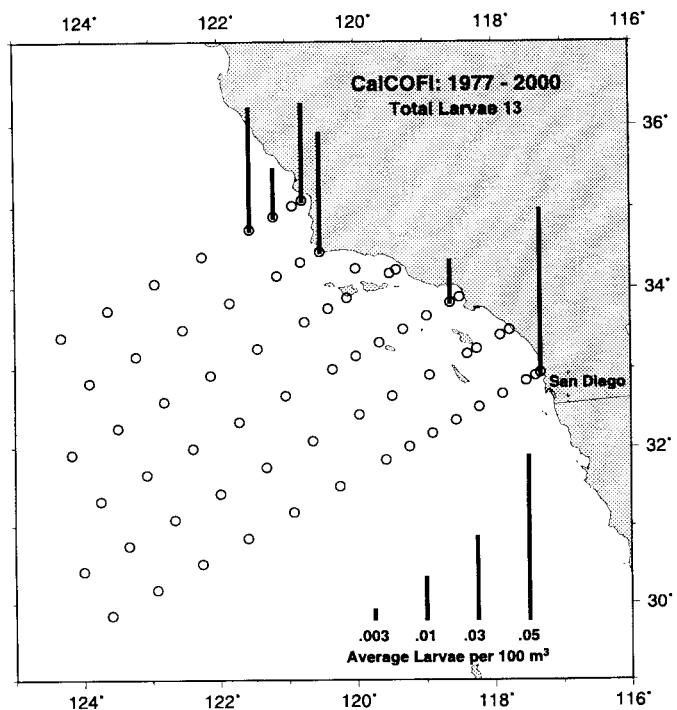
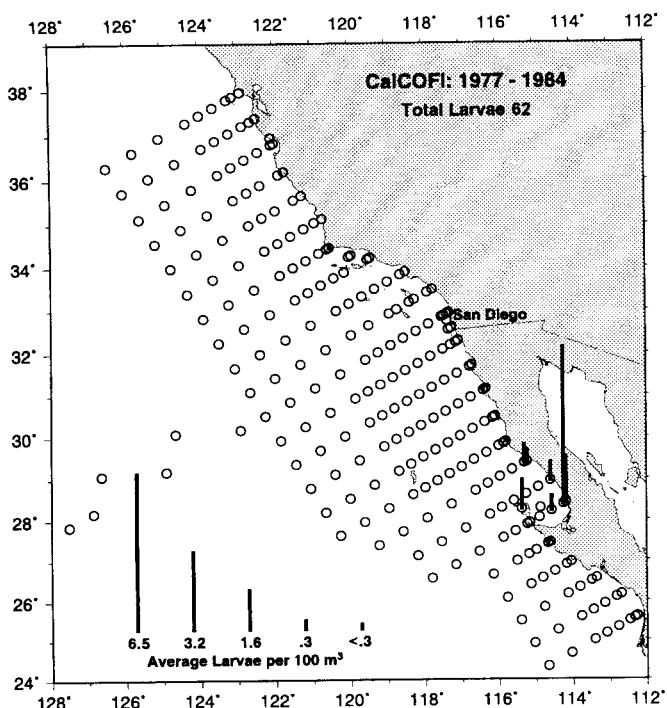
Leuroglossus stibius



Synodus lucioceps

California lizardfish

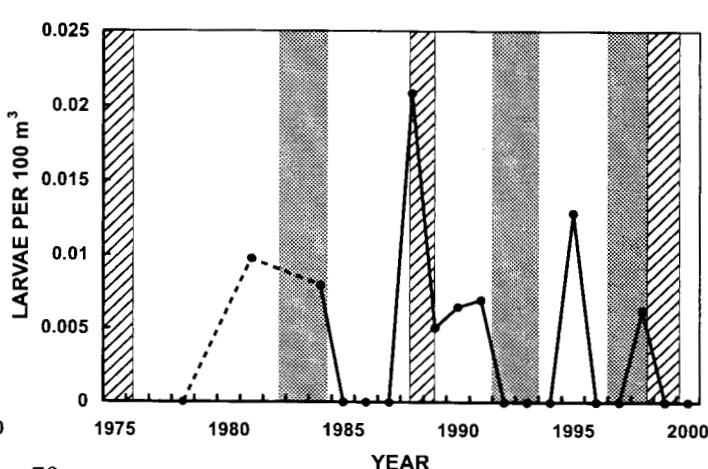
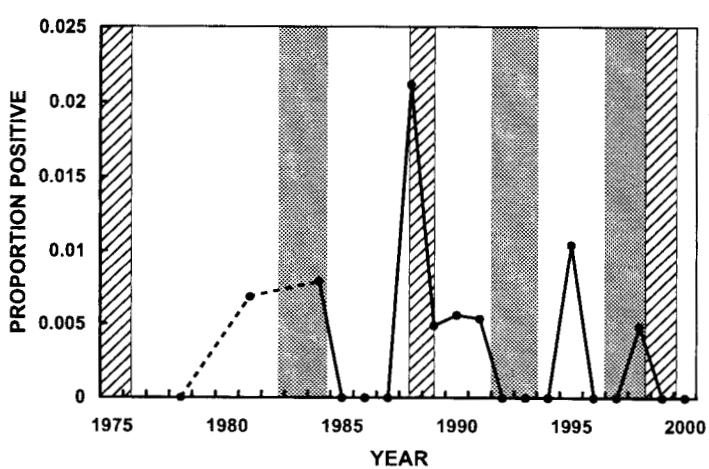
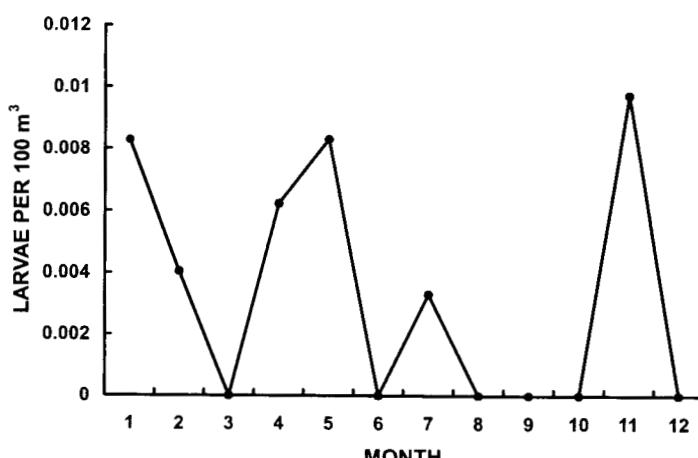
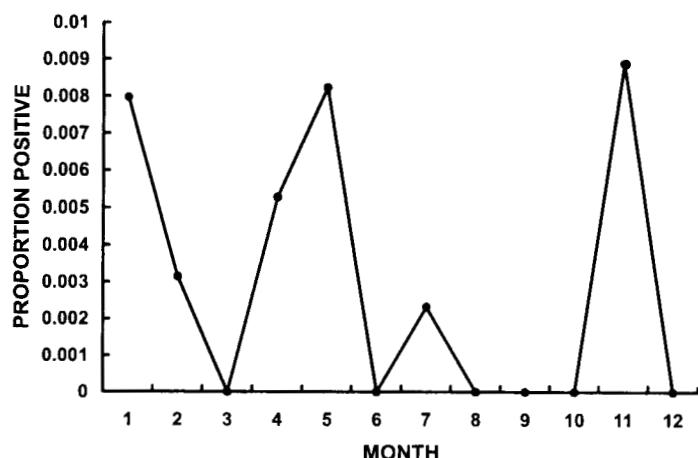
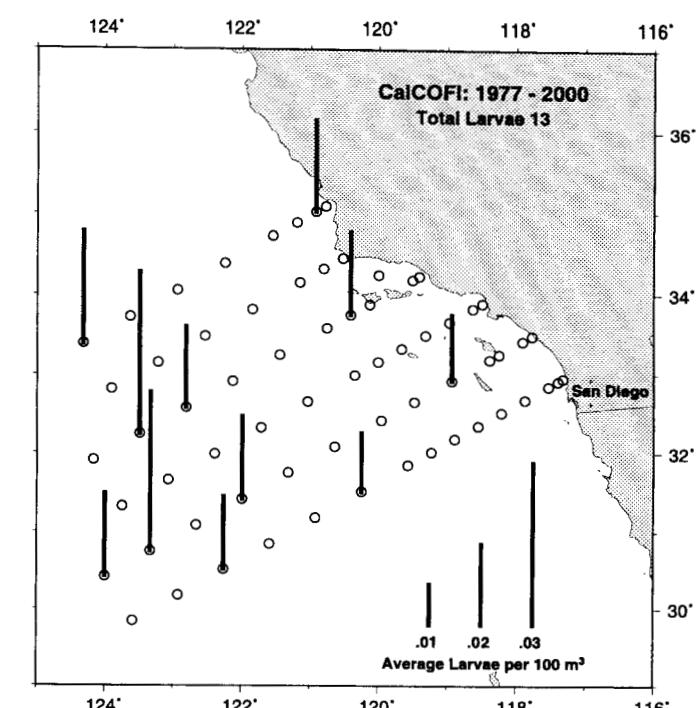
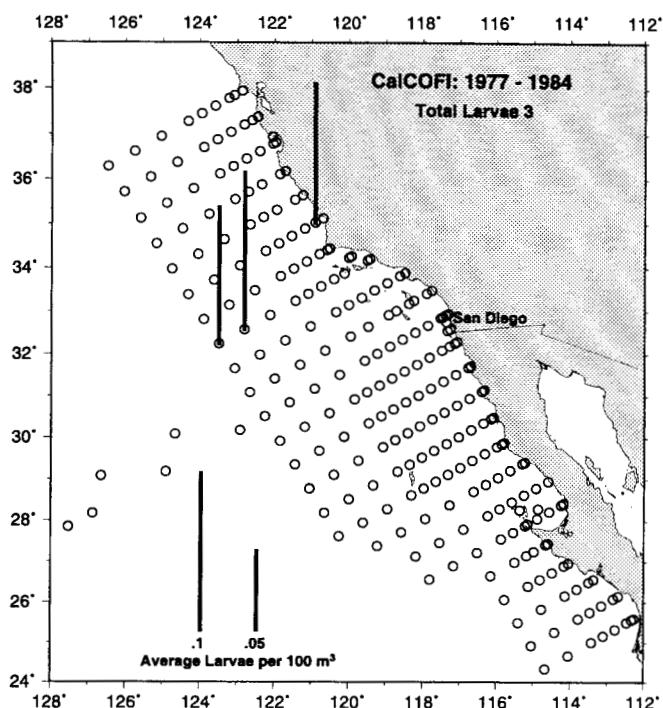
SYNODONTIDAE



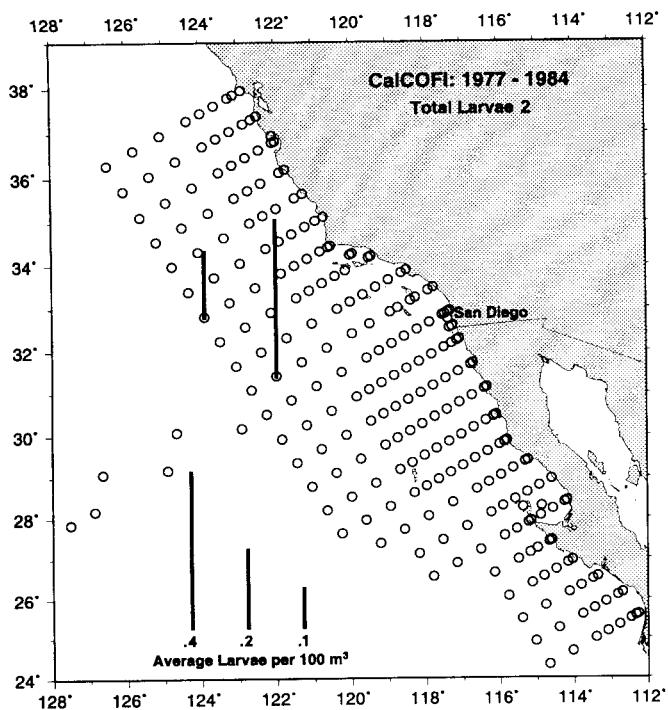
MYCTOPHIDAE

Longfin lanternfish

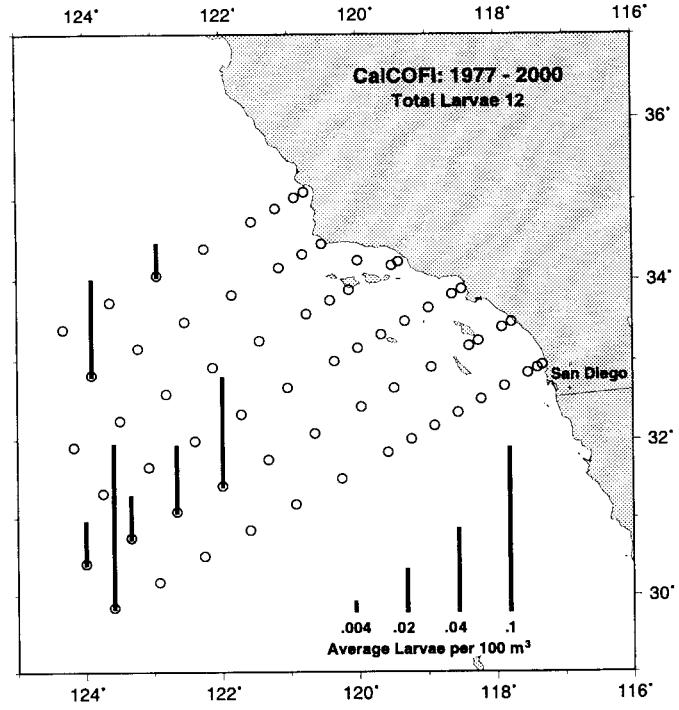
Diogenichthys atlanticus



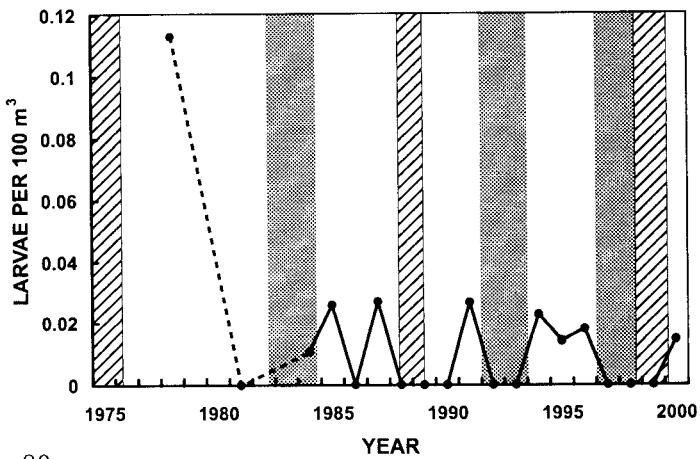
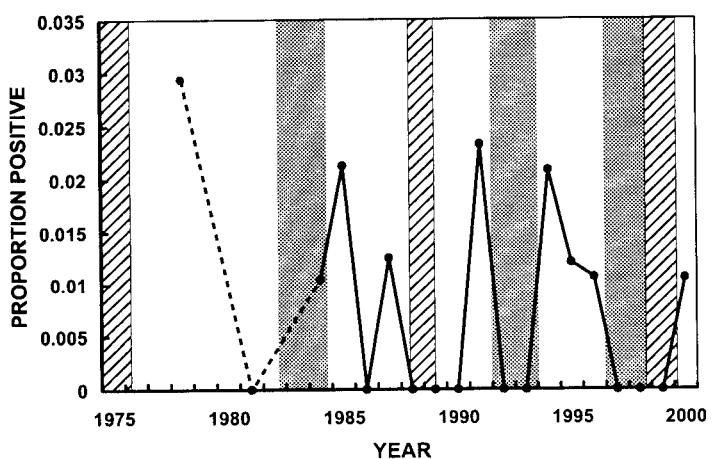
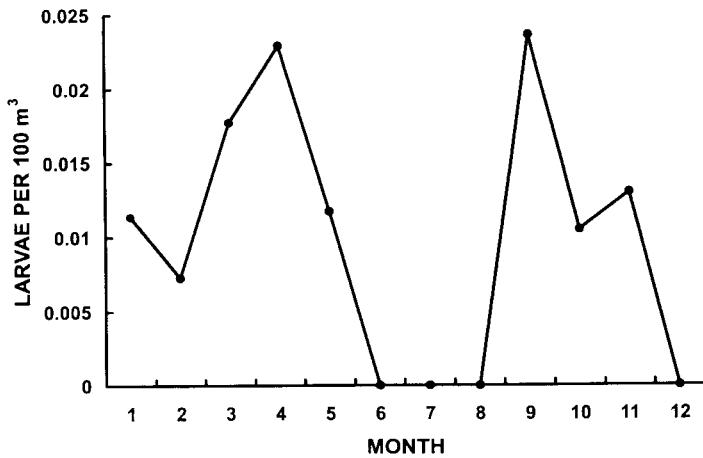
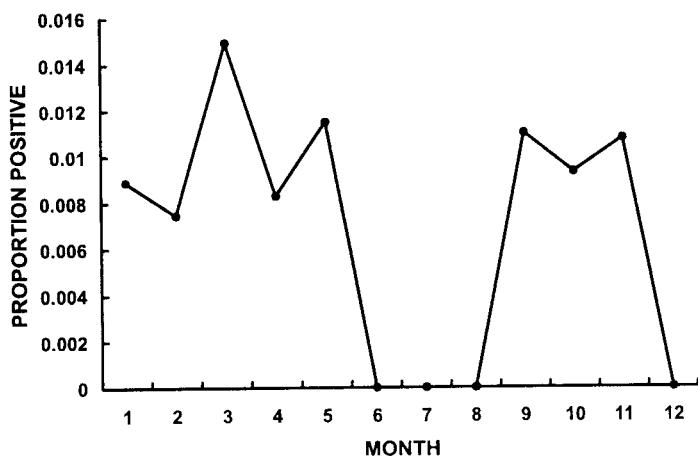
Bathophilus flemingi



Highfin dragonfish



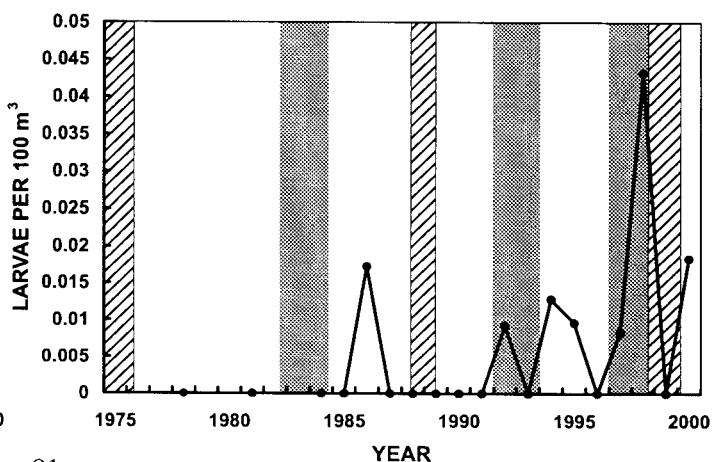
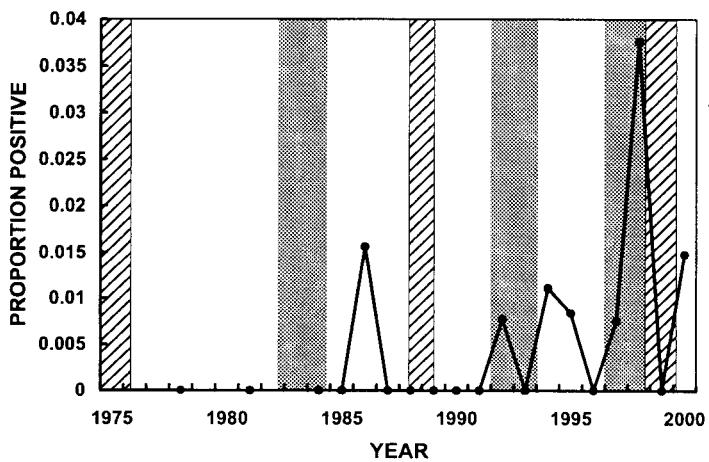
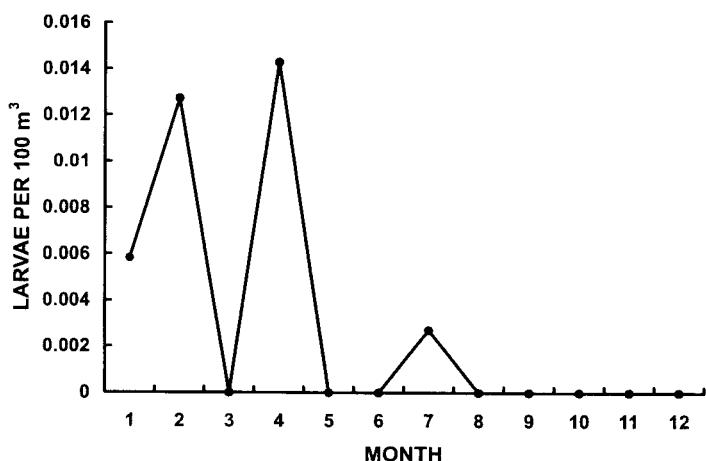
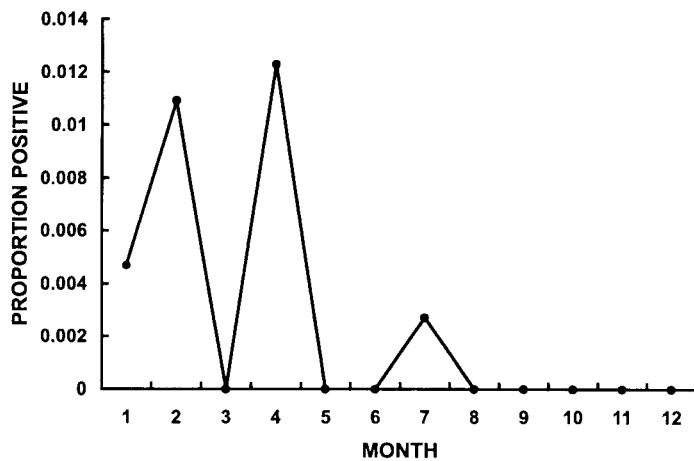
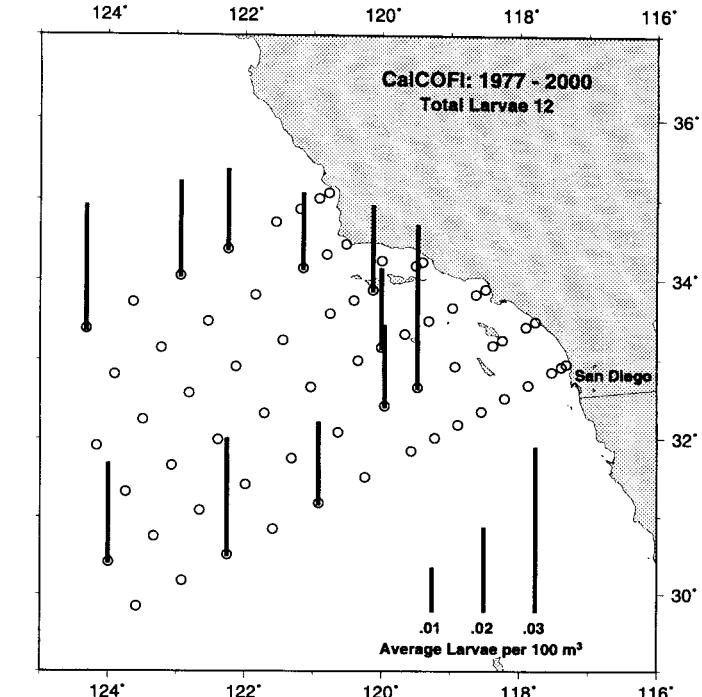
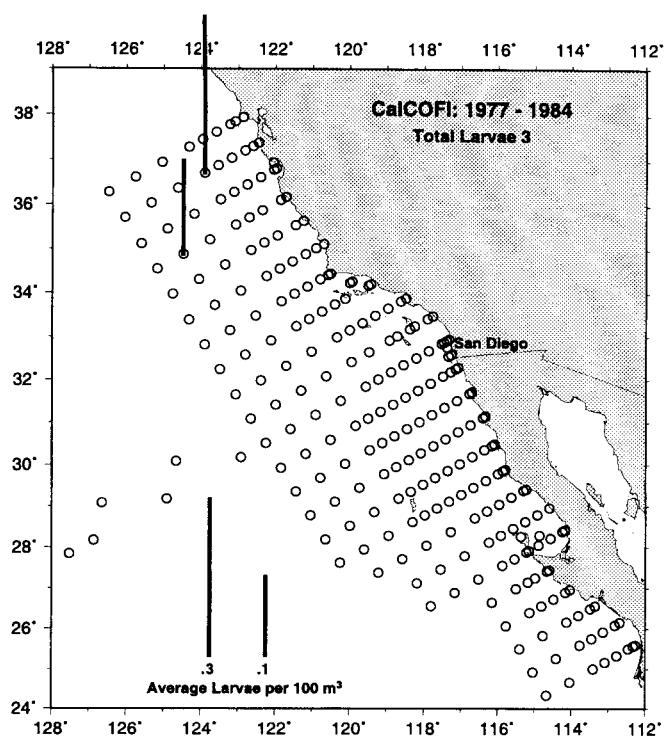
STOMIIDAE



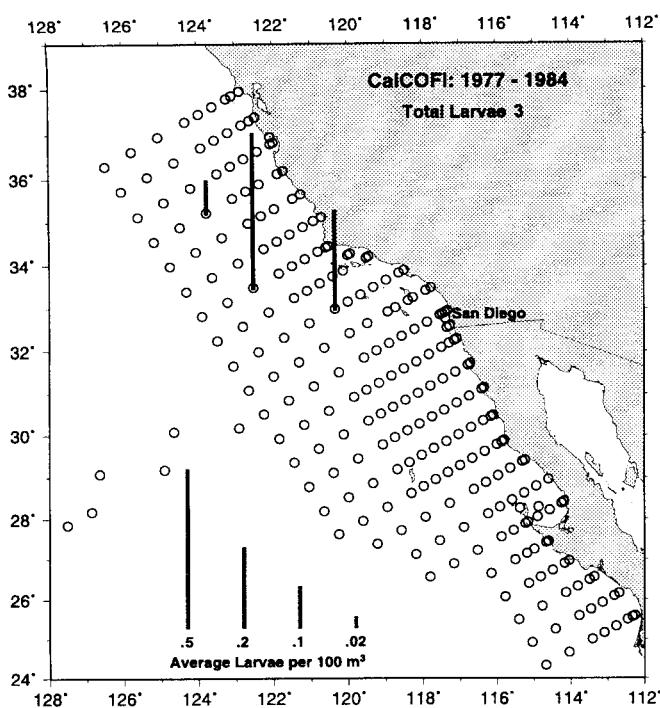
MYCTOPHIDAE

California flashlightfish

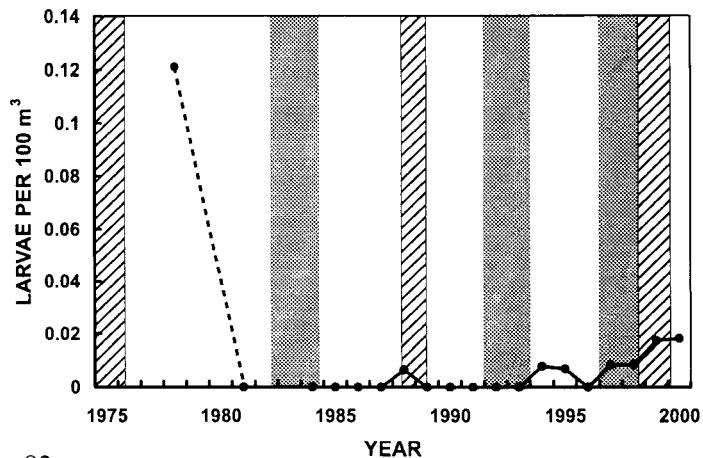
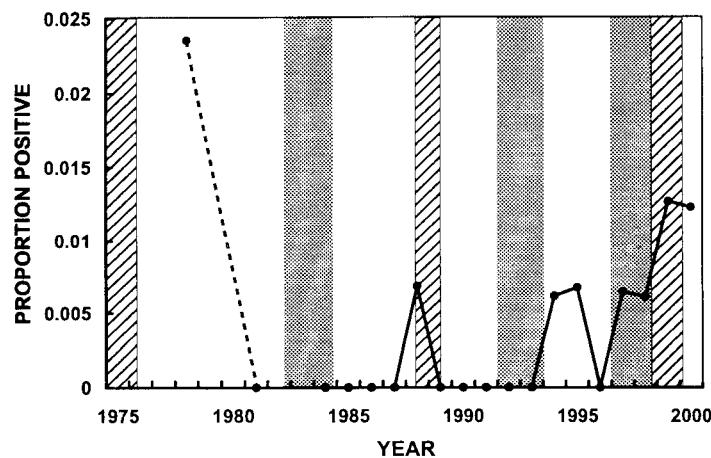
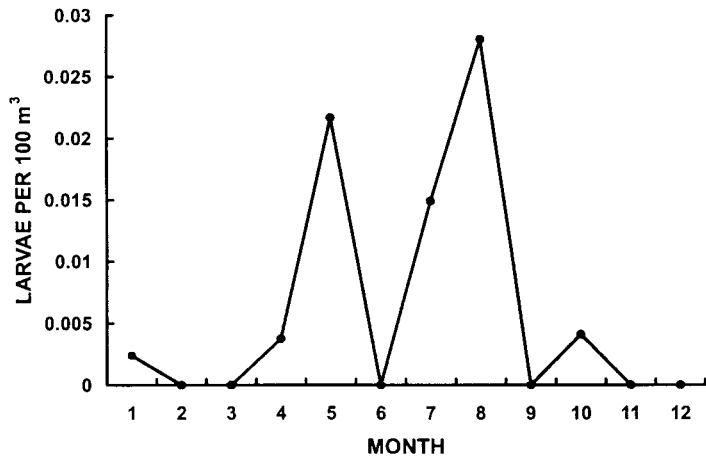
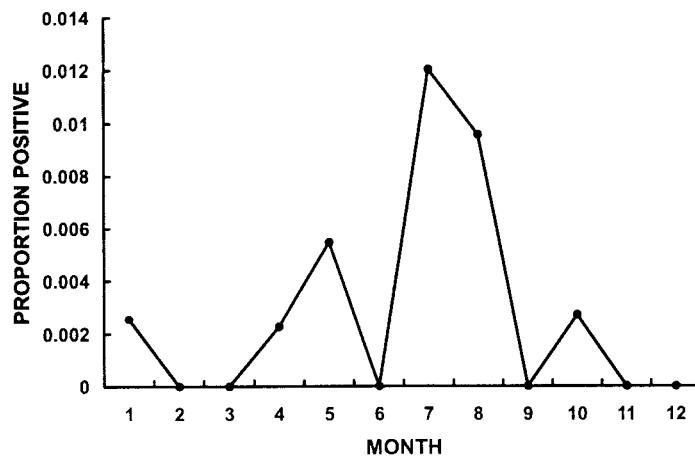
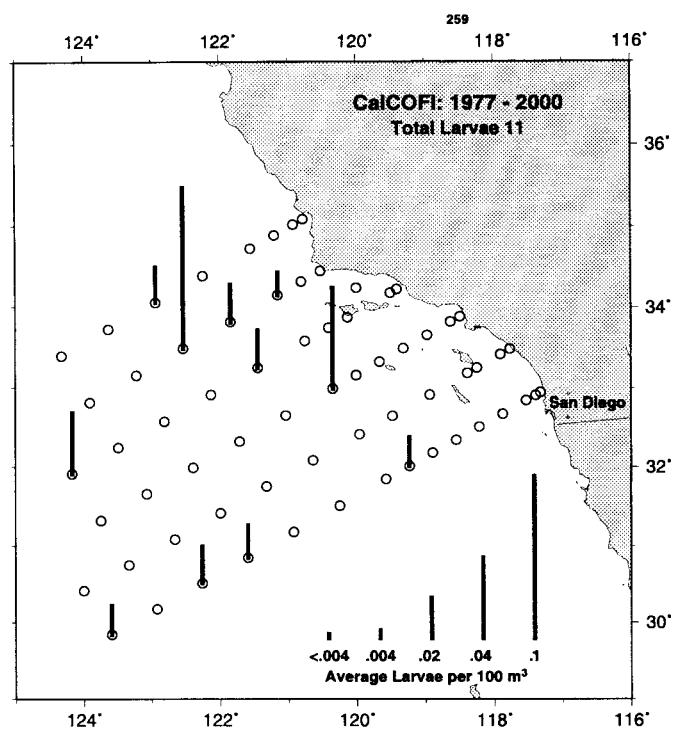
Protomyctophum crockeri



Nannobrachium regale



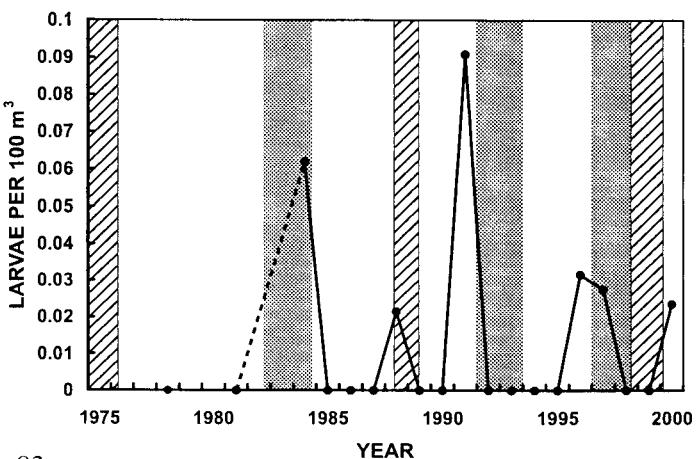
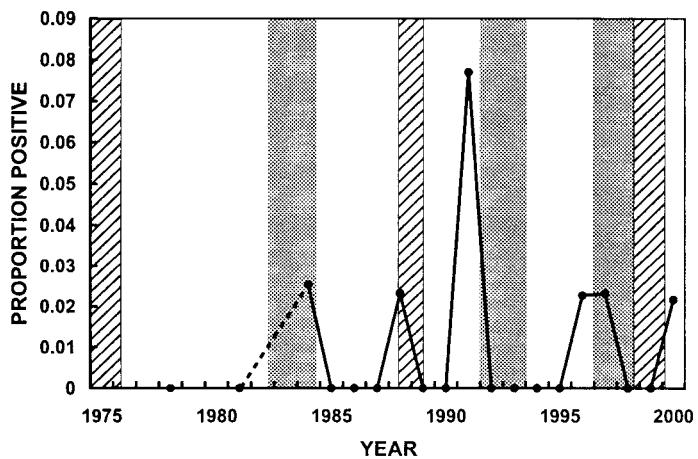
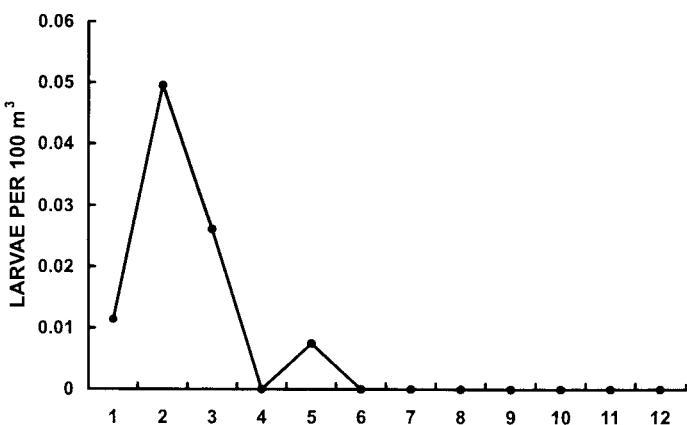
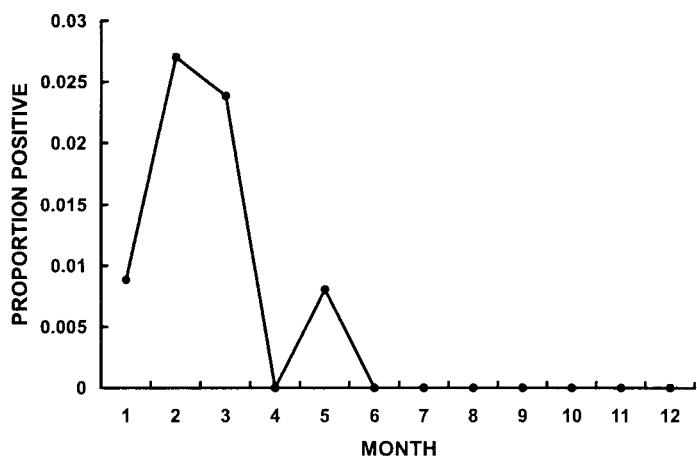
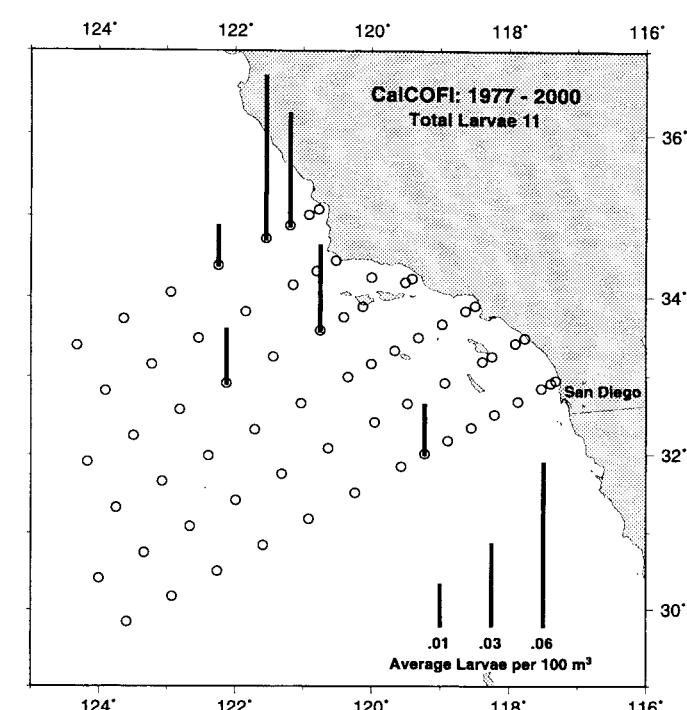
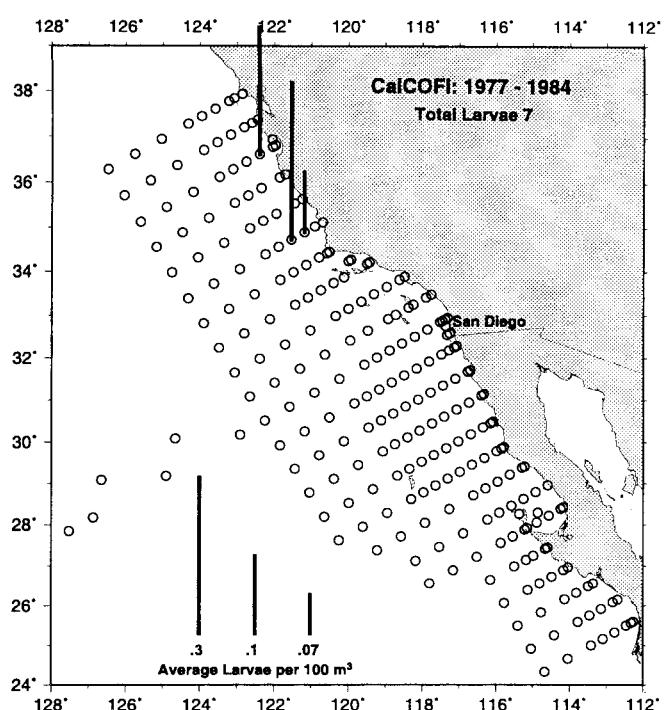
Pinpoint lightfish



ICOSTEIDAE

Ragfish

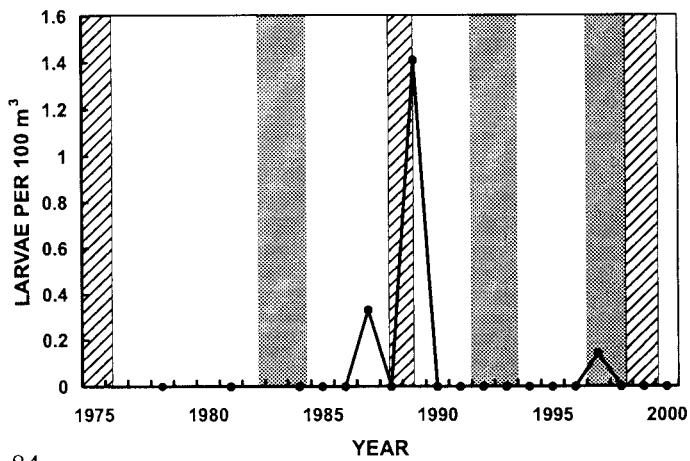
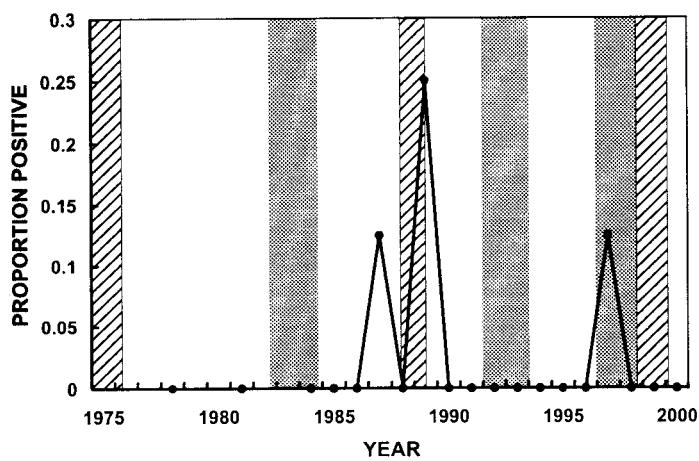
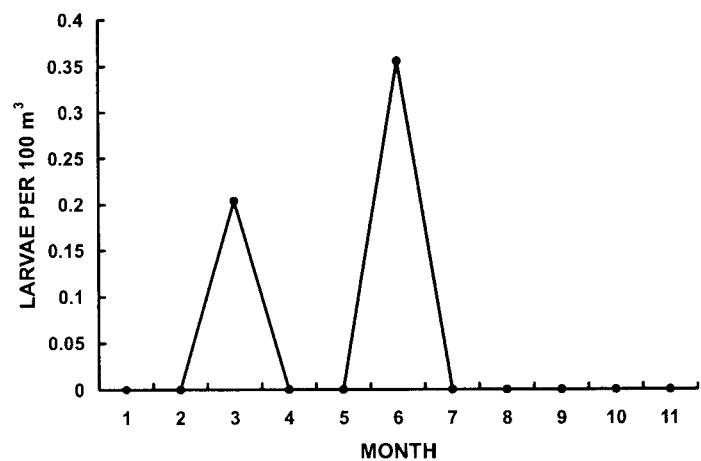
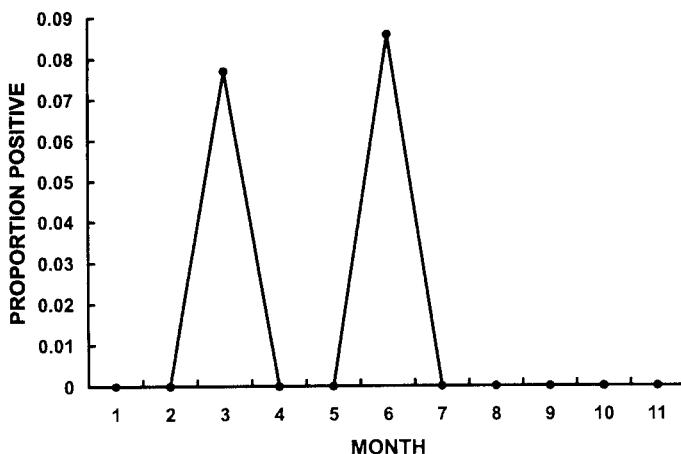
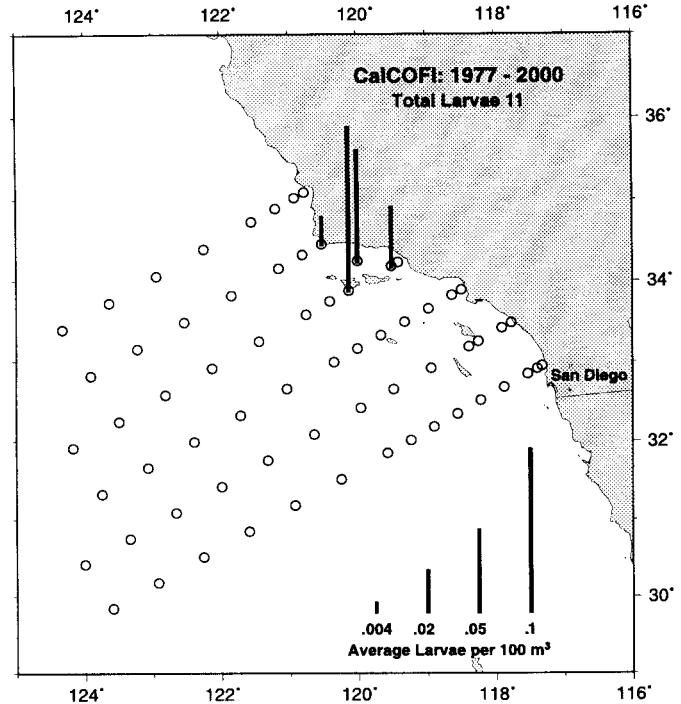
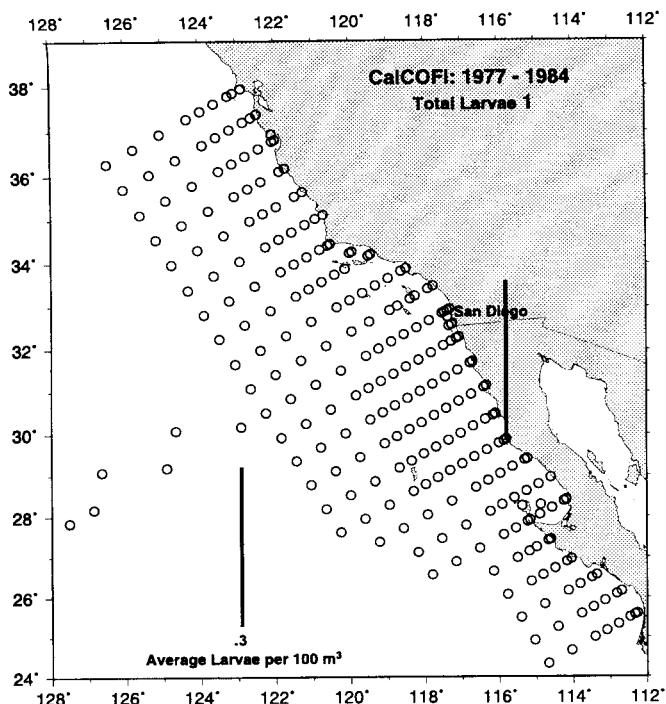
Icosteus aenigmaticus



Cryptotrema corallinum

Deepwater kelpfish

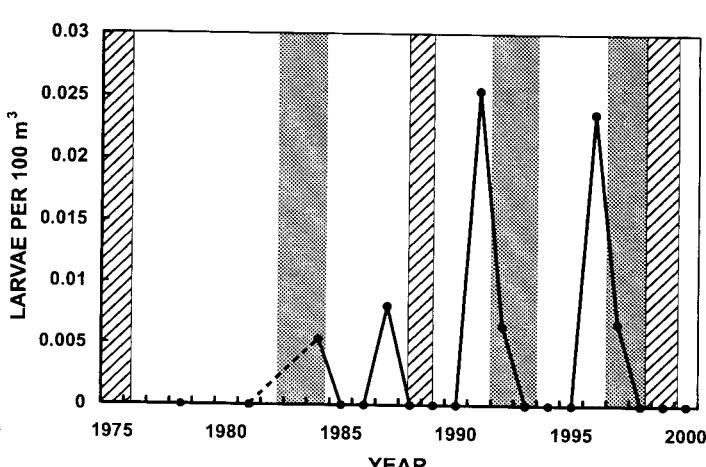
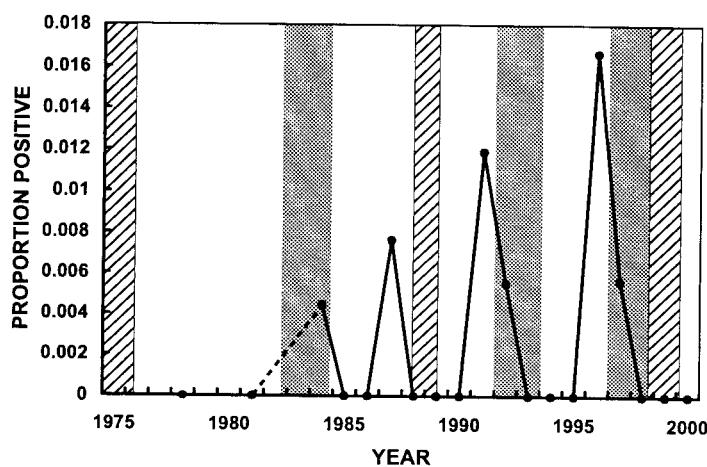
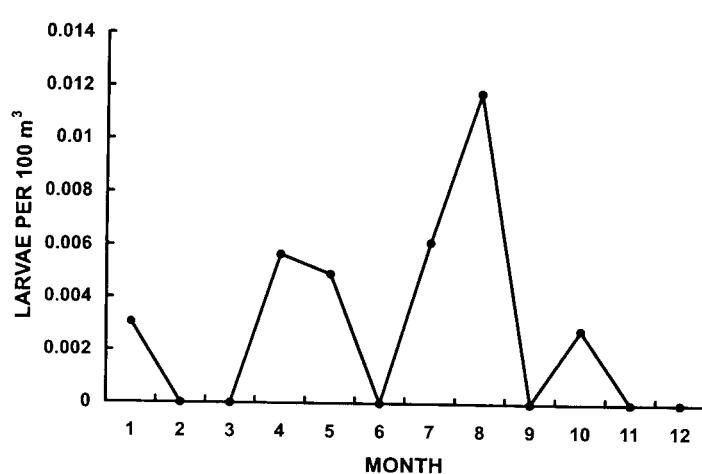
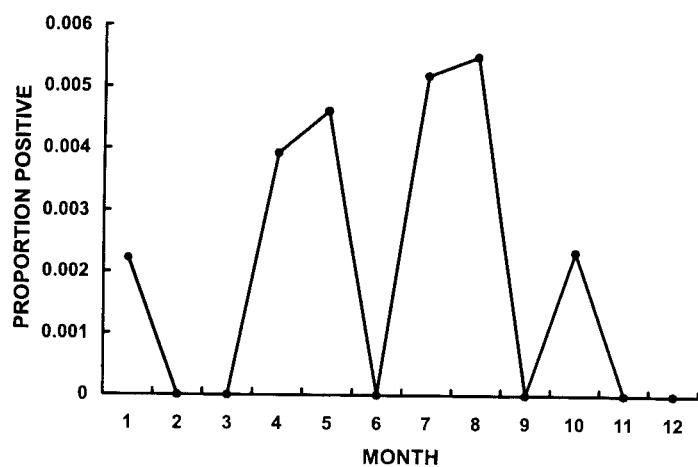
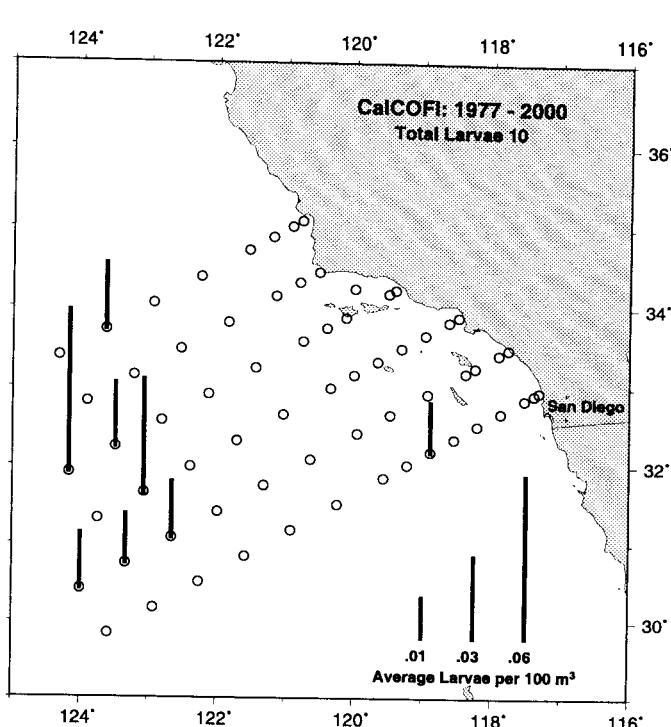
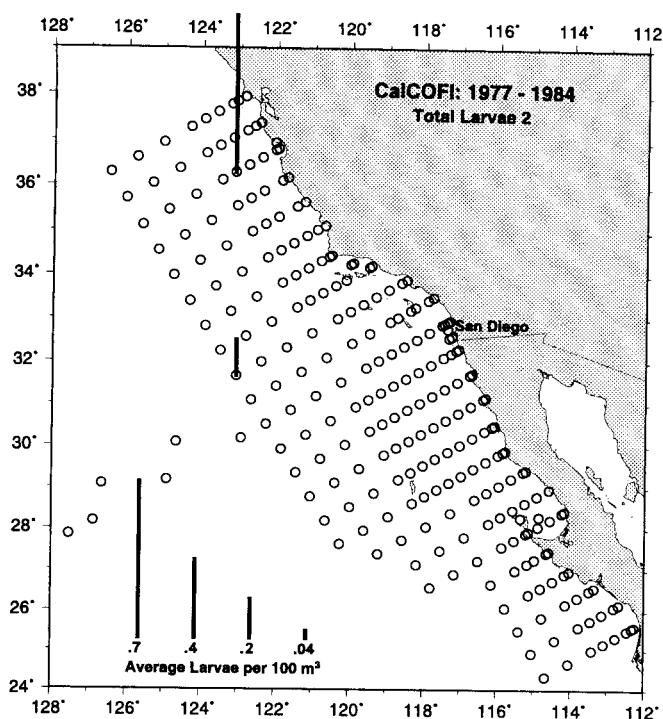
LABRISOMIDAE



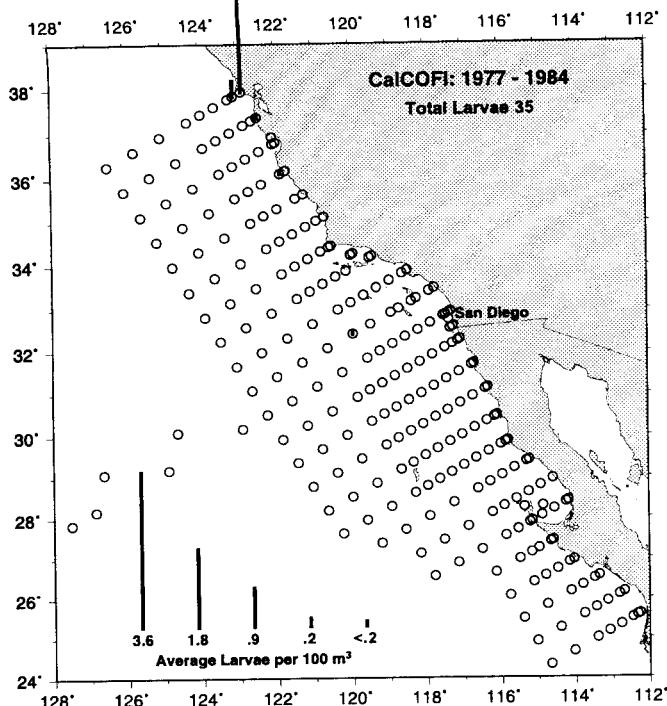
STOMIIDAE

Longfin dragonfish

Tactostoma macropus

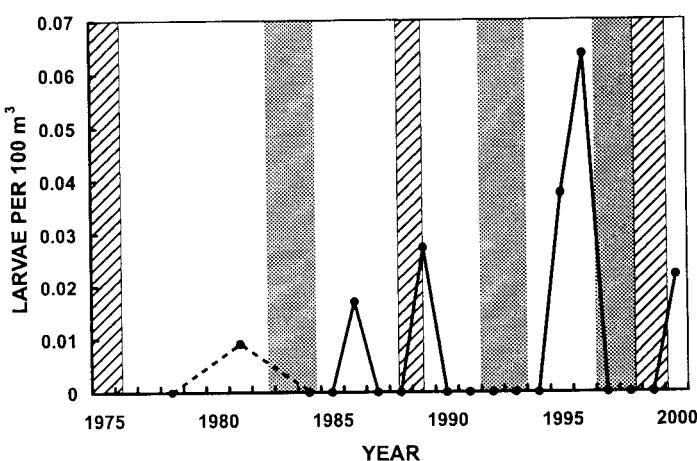
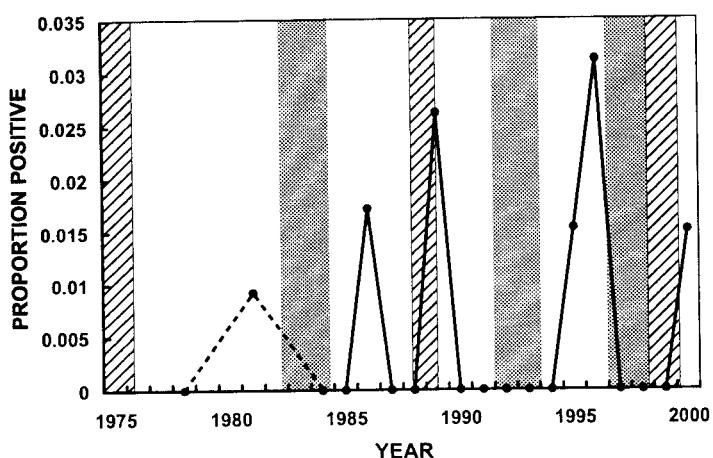
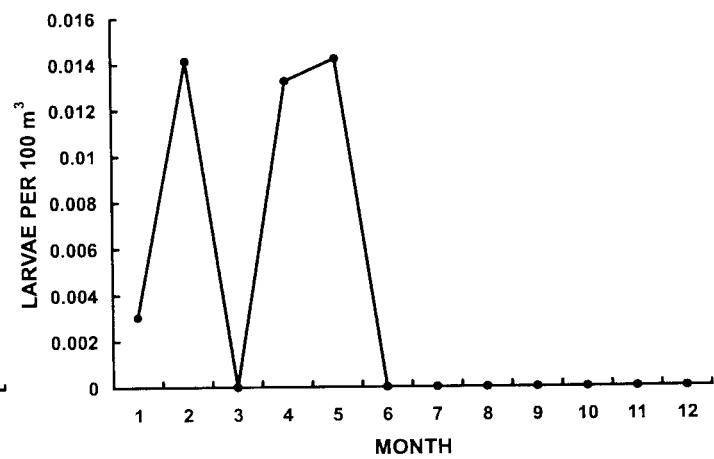
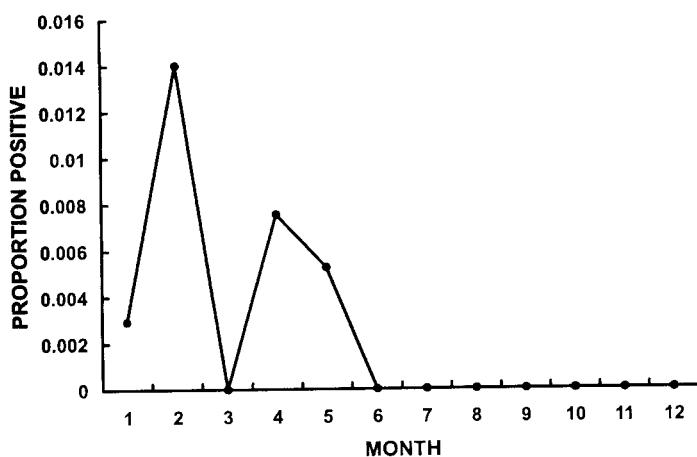
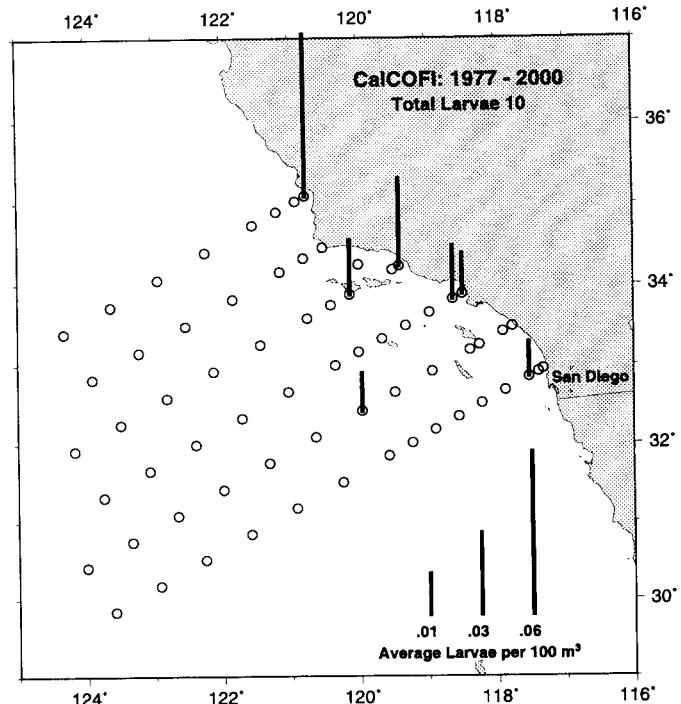


Parophrys vetulus



English sole

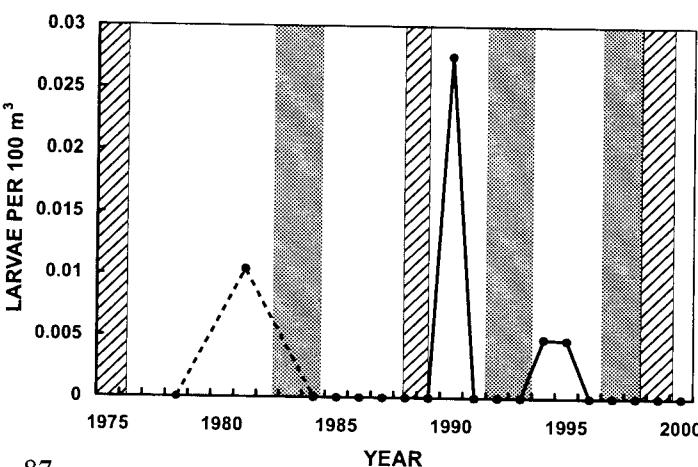
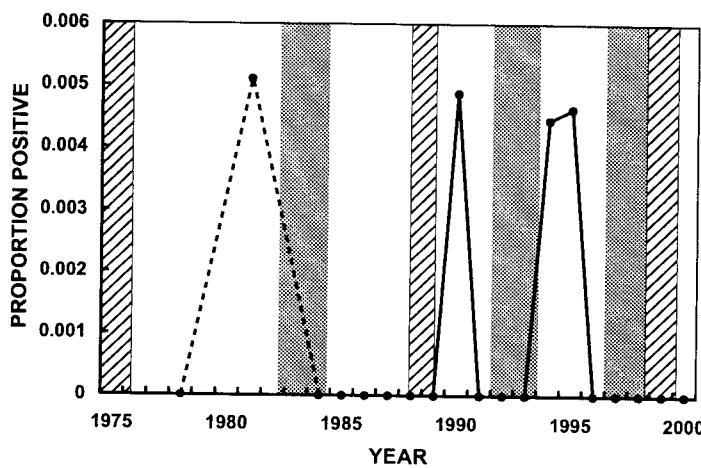
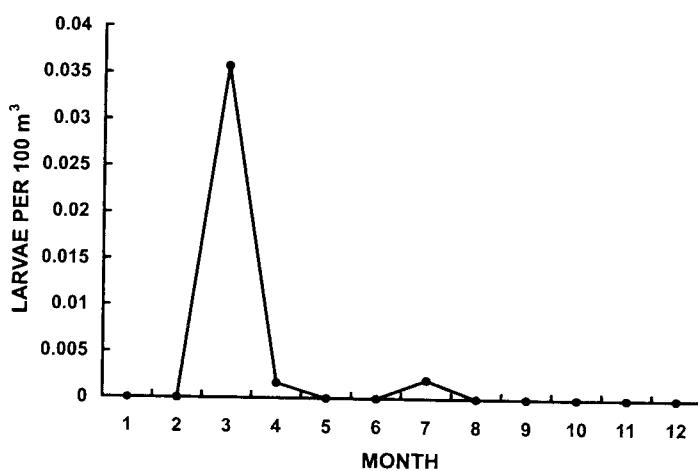
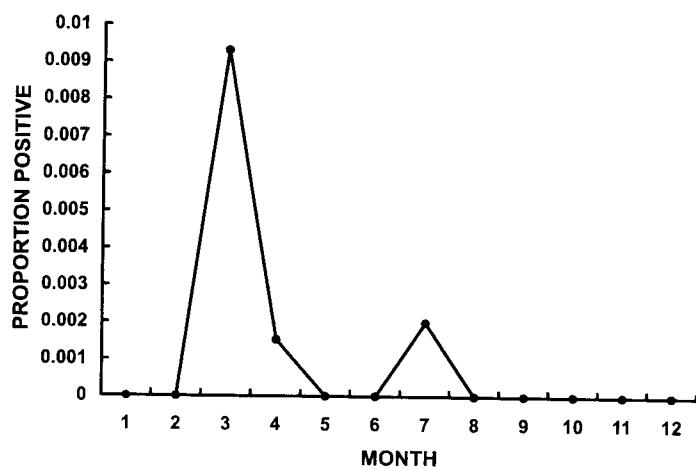
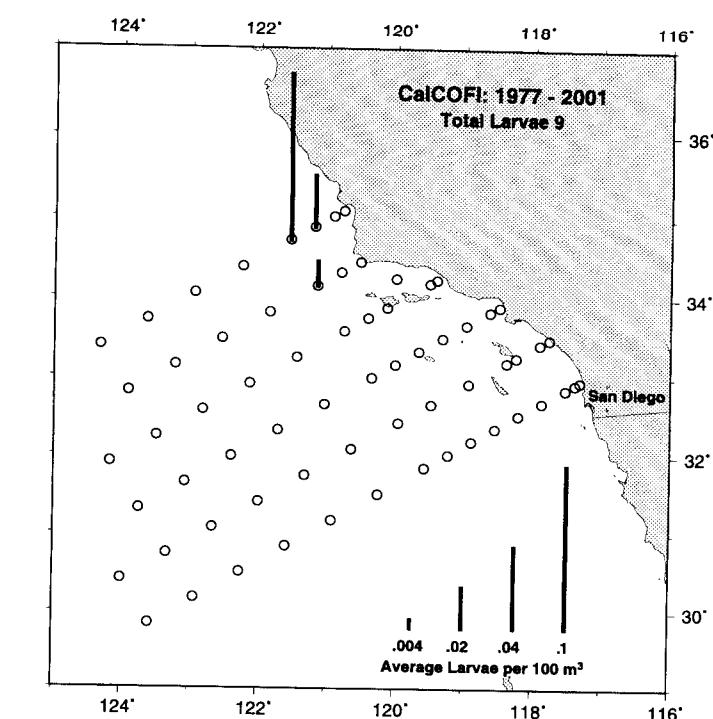
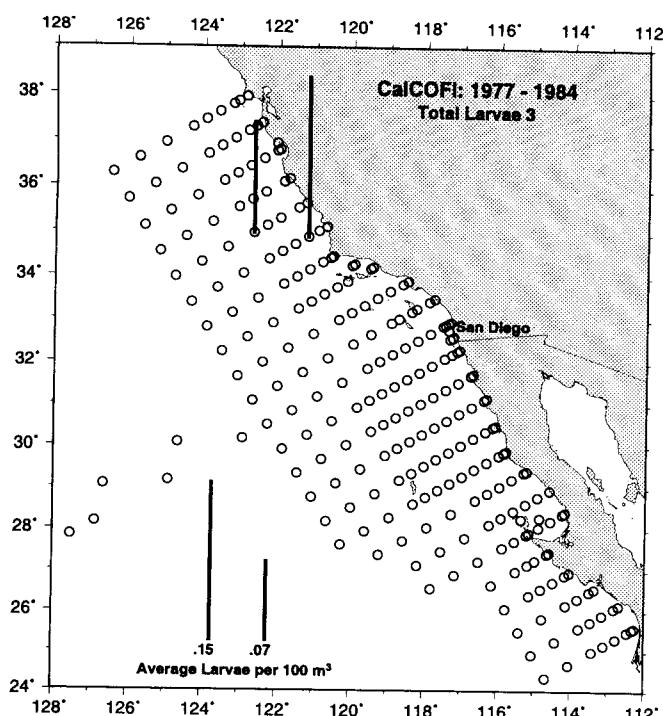
PLEURONECTIDAE



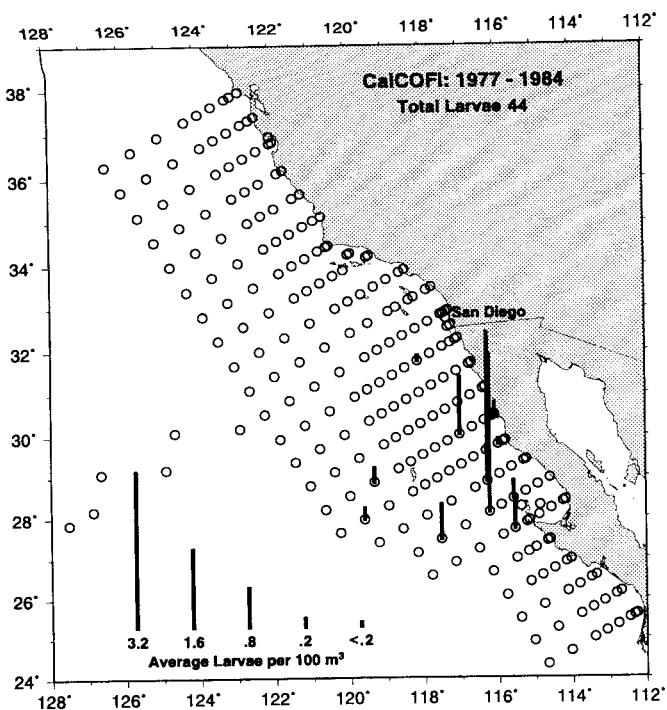
SEASTIDAE

Thornyheads

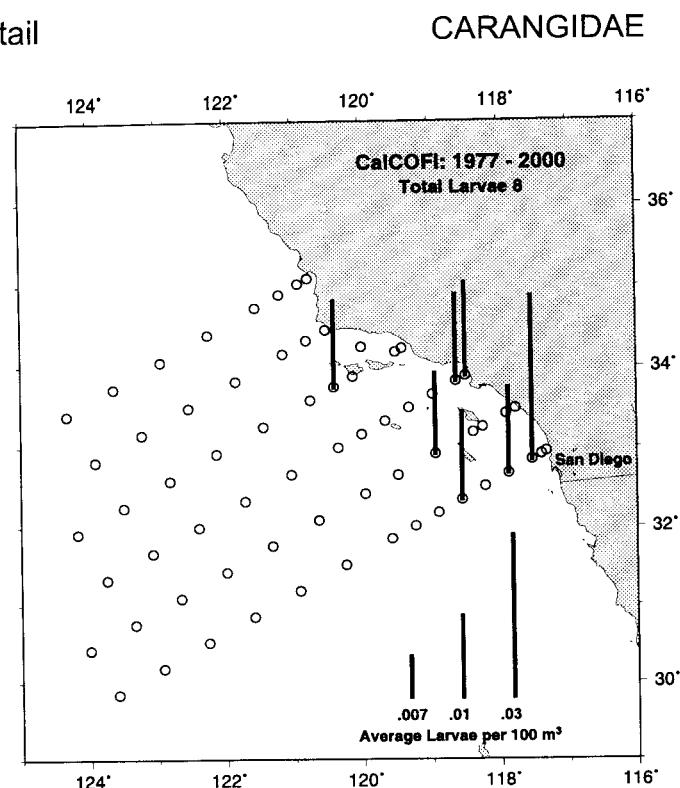
Sebastolobus spp.



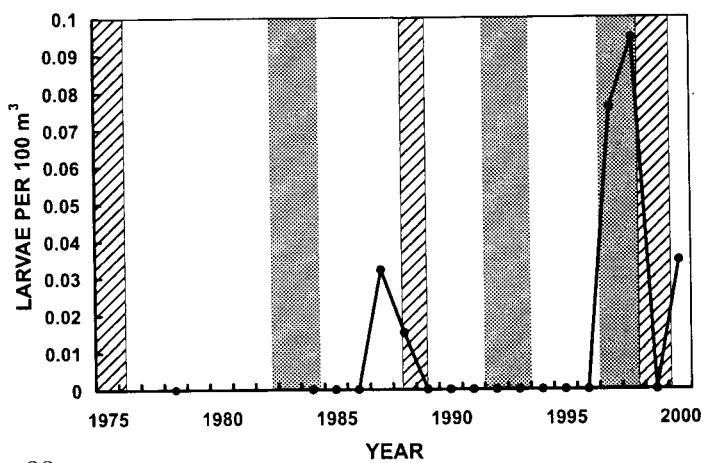
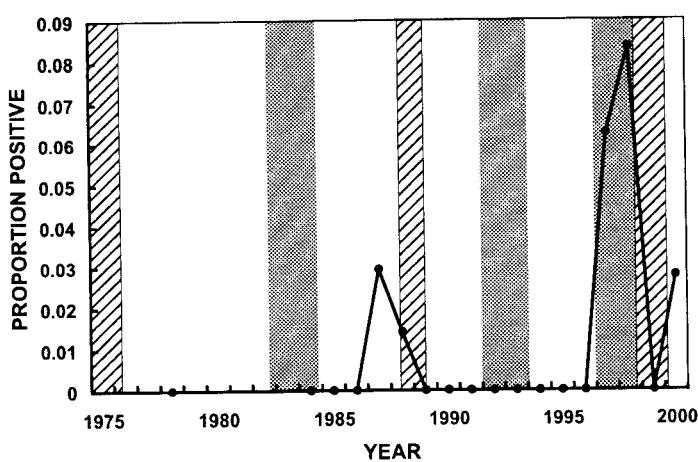
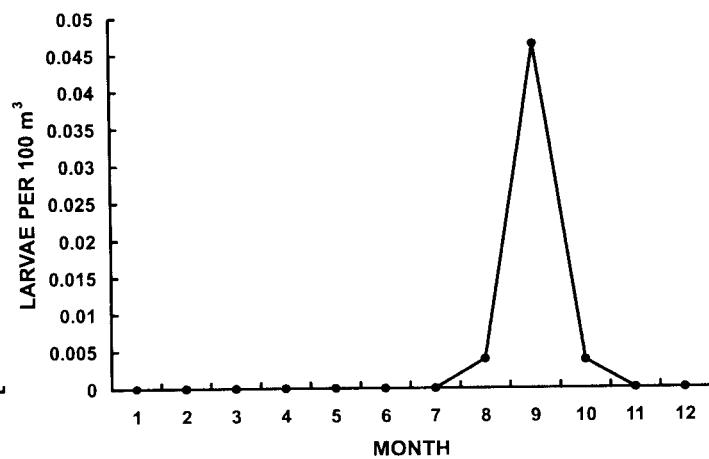
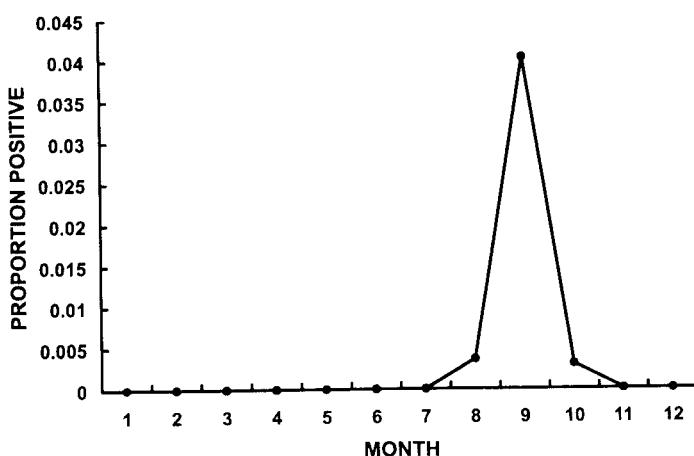
Seriola lalandi



Yellowtail



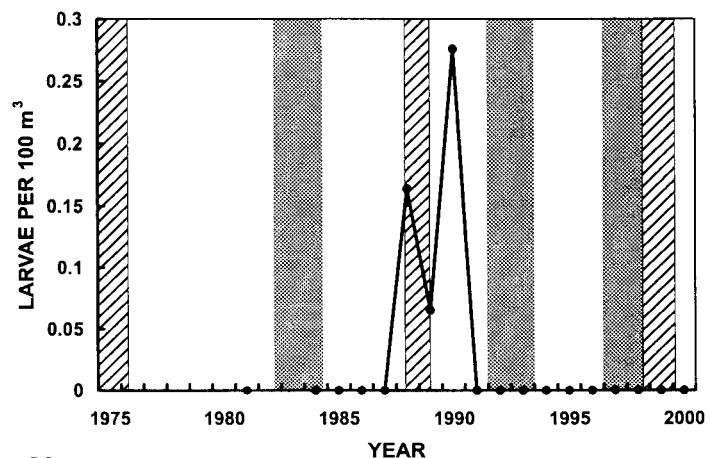
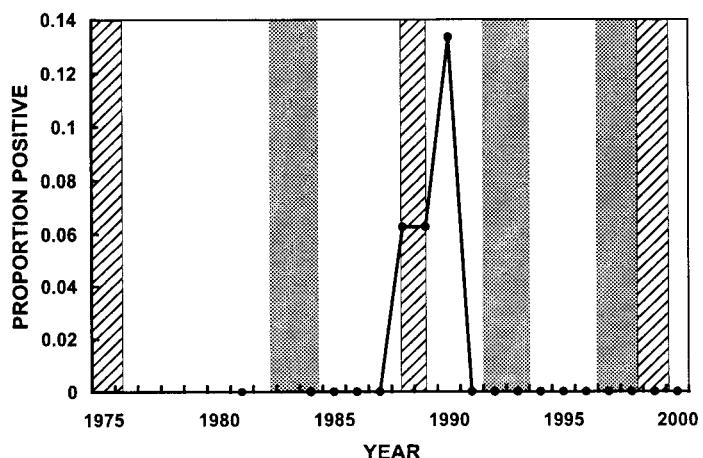
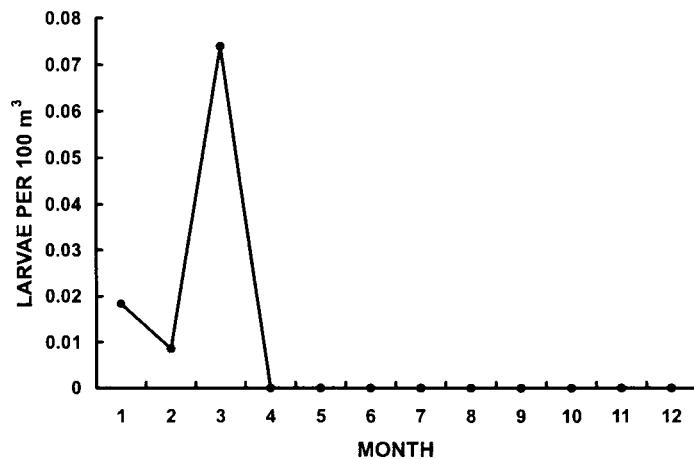
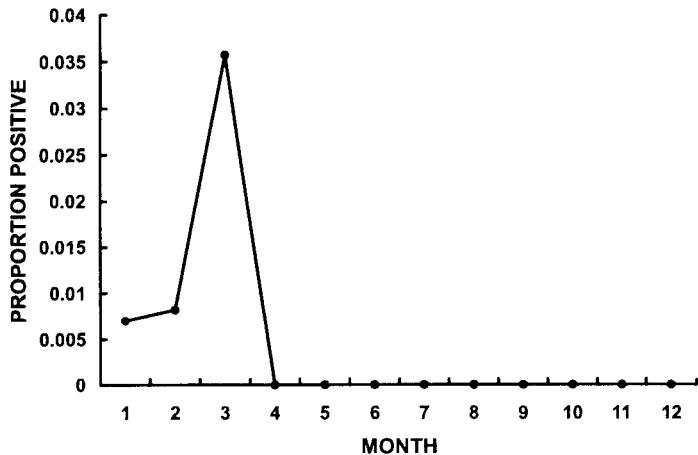
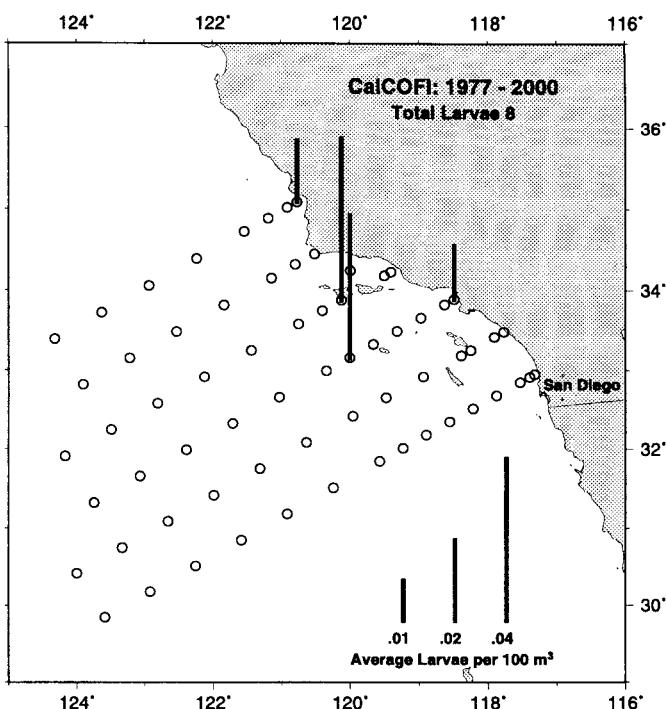
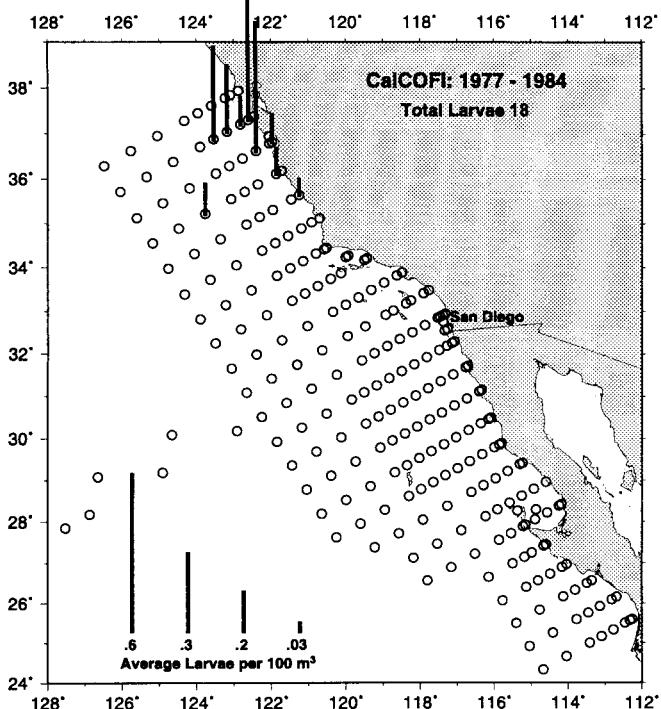
CARANGIDAE



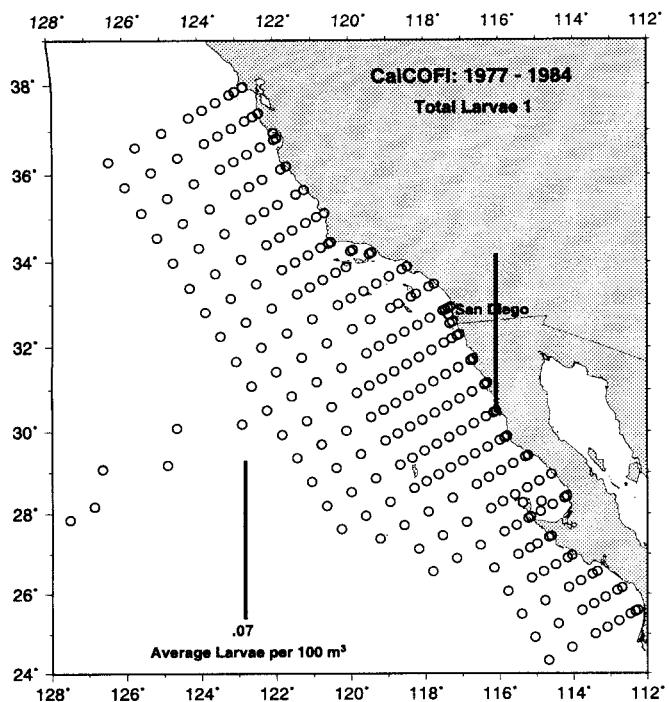
HEXAGRAMMIDAE

Rock greenling

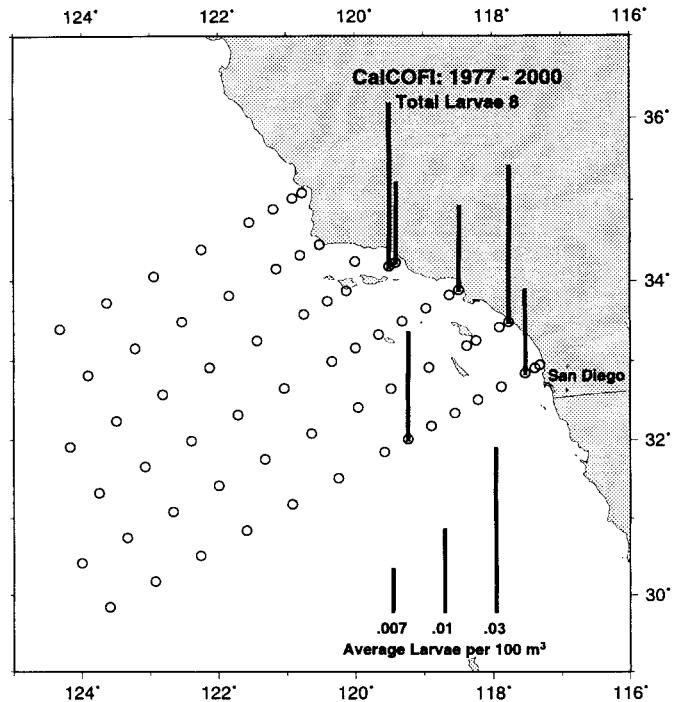
Hexagrammos lagocephalus



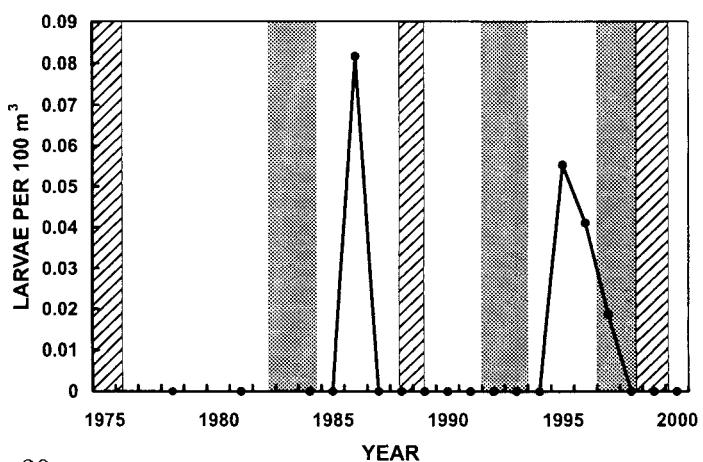
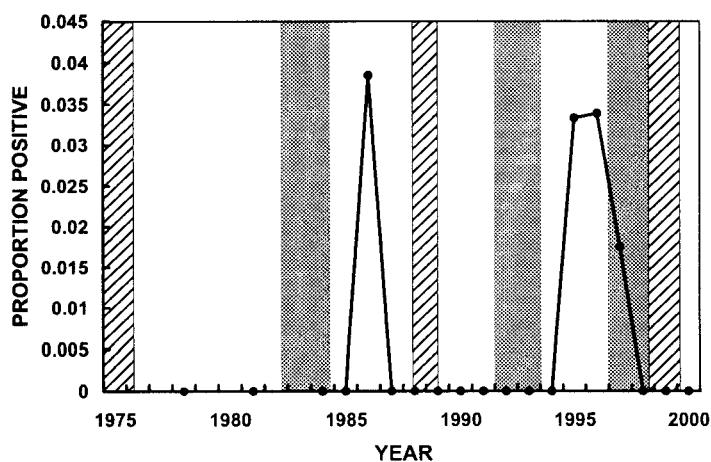
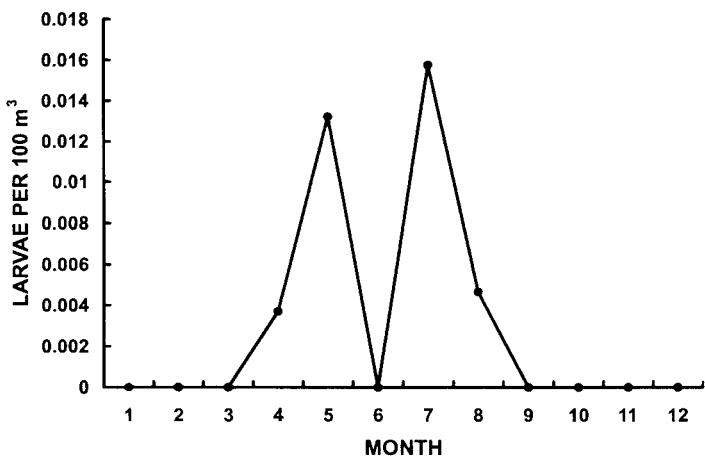
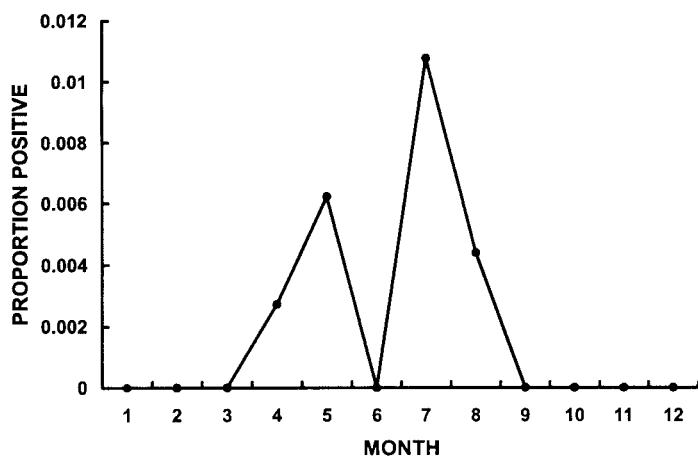
Atractoscion nobilis



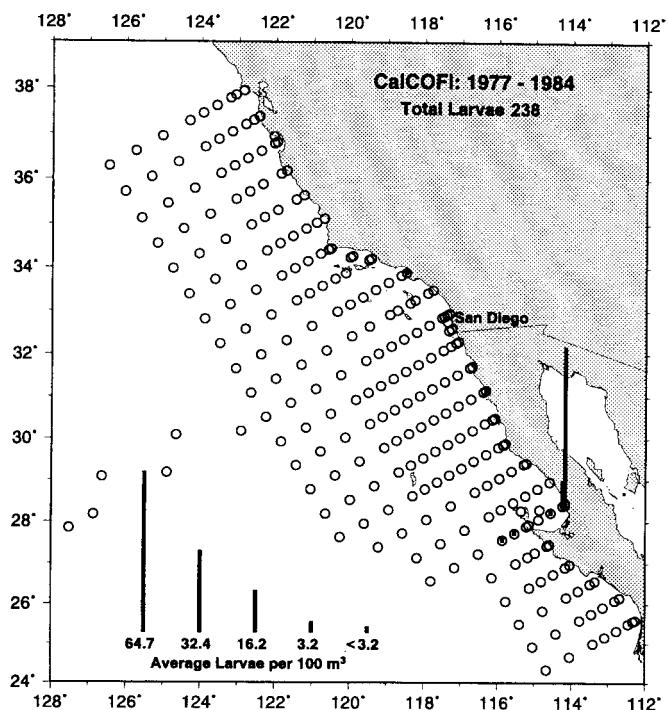
White seabass



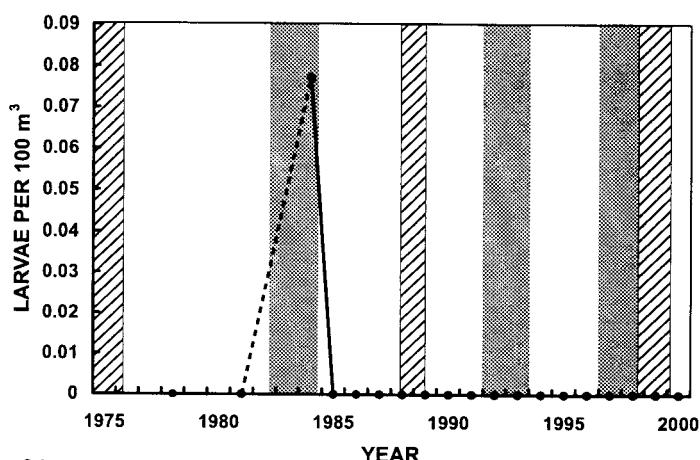
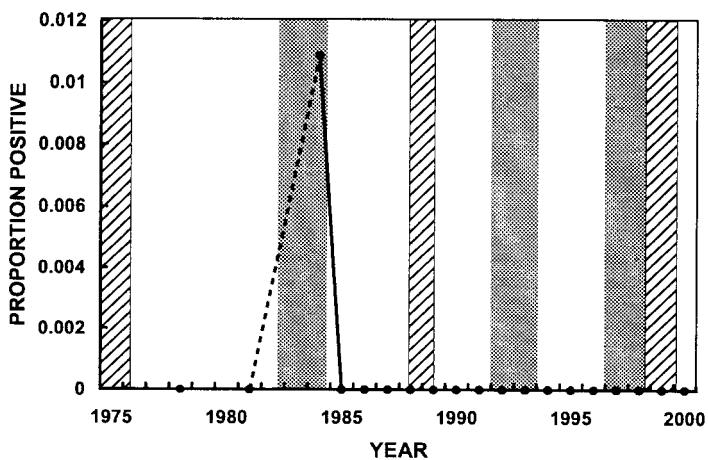
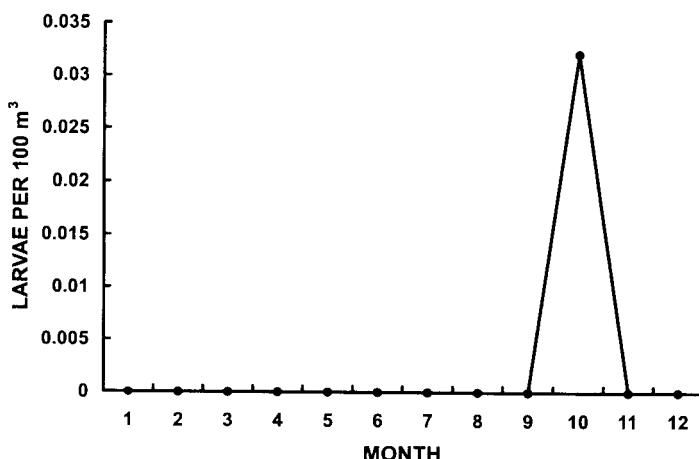
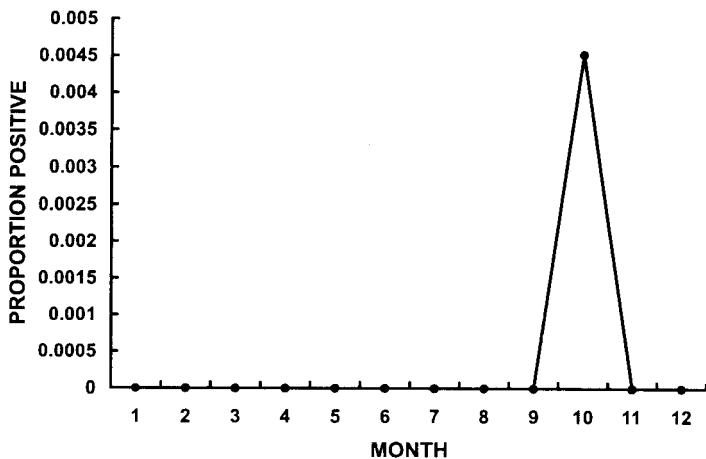
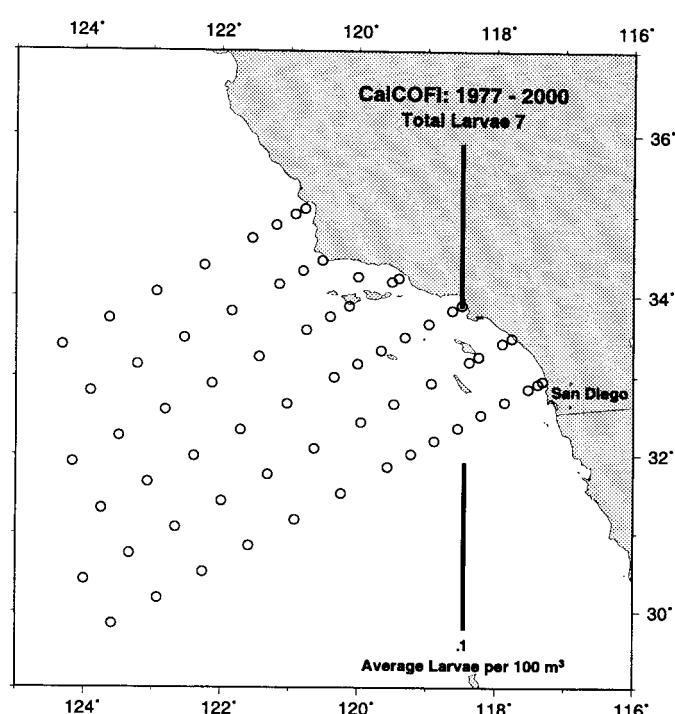
SCIAENIDAE



CLUPEIDAE



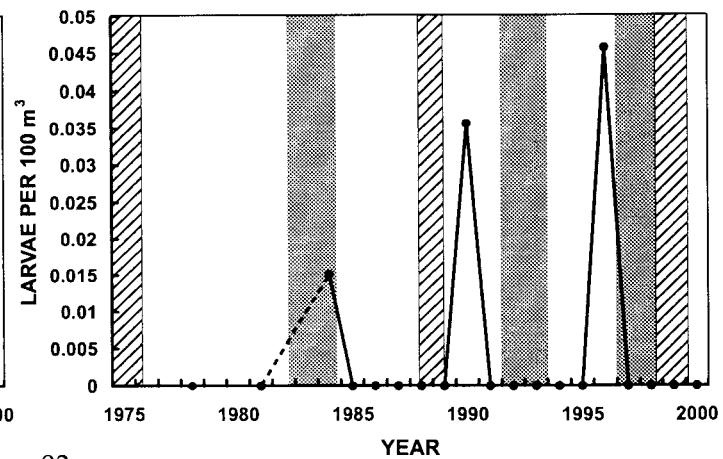
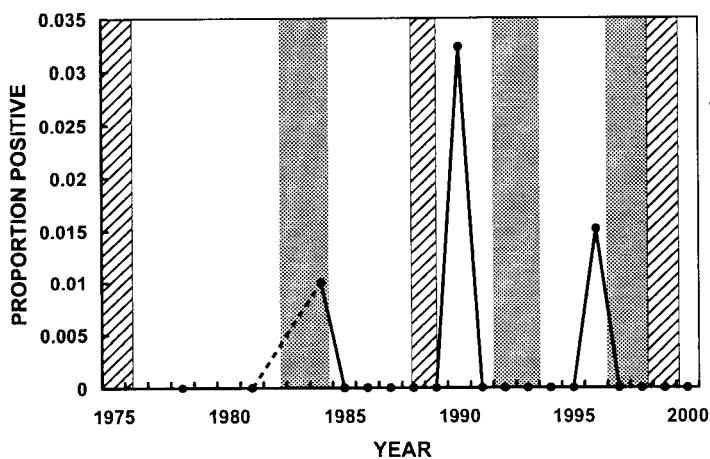
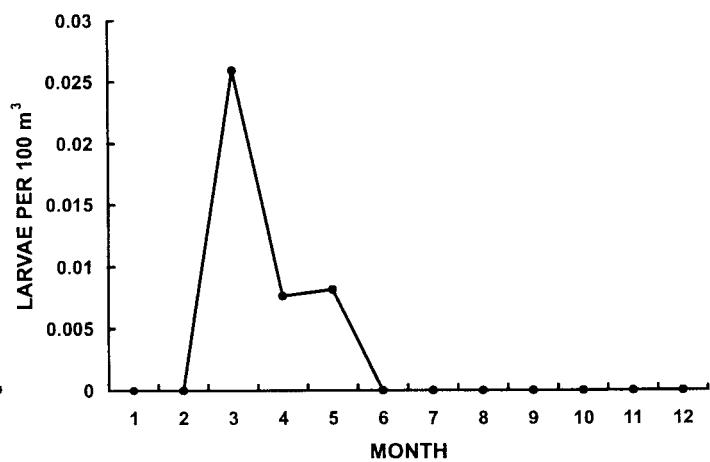
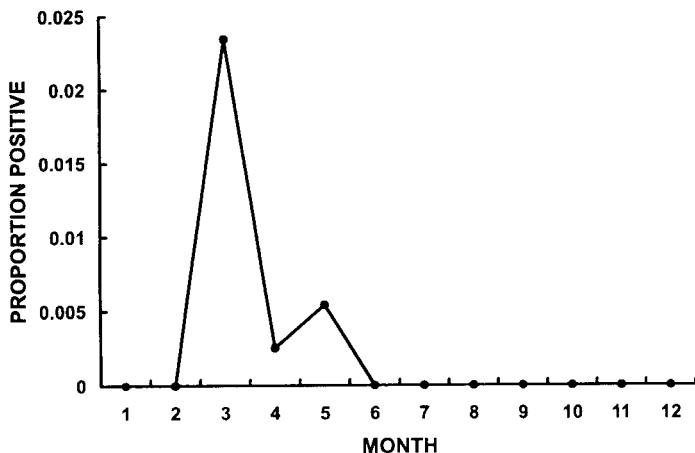
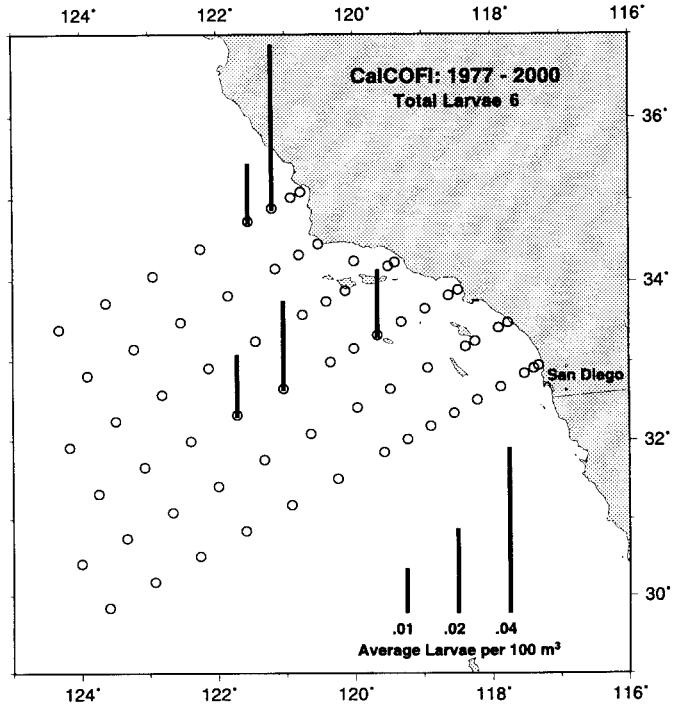
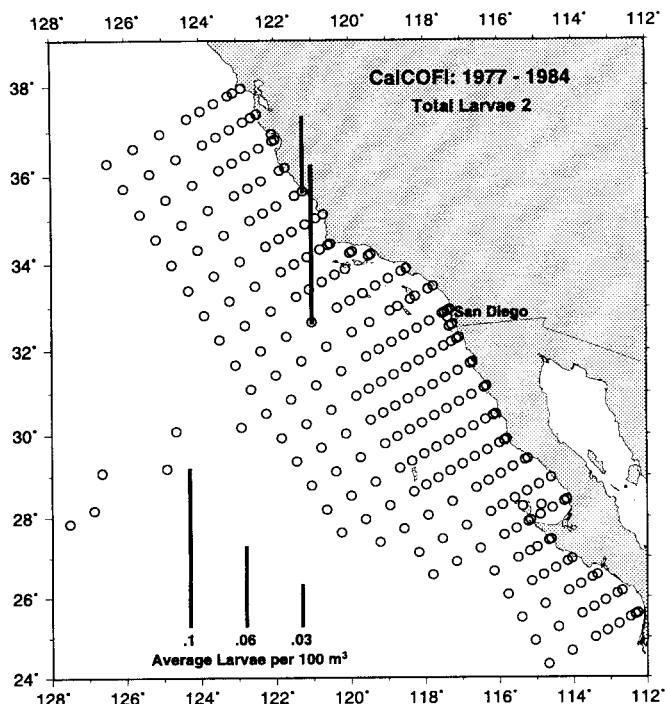
Round herring



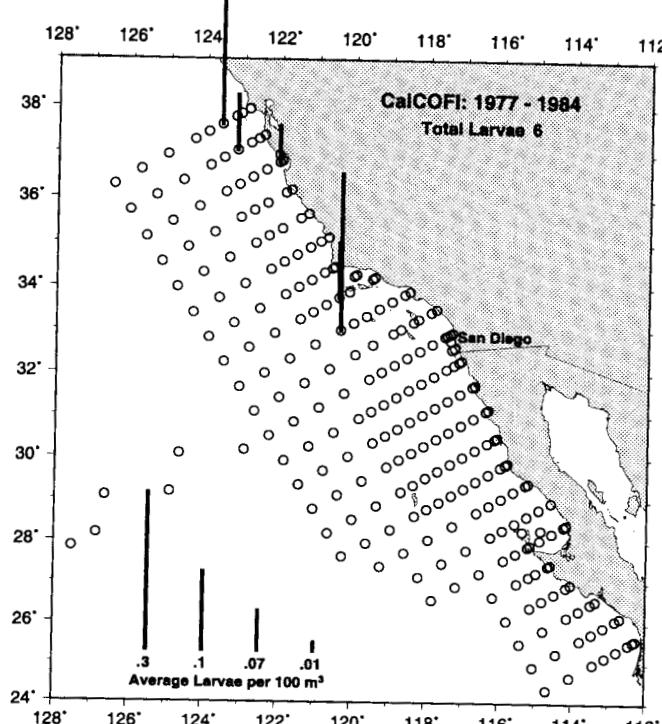
Microstomus pacificus

Dover sole

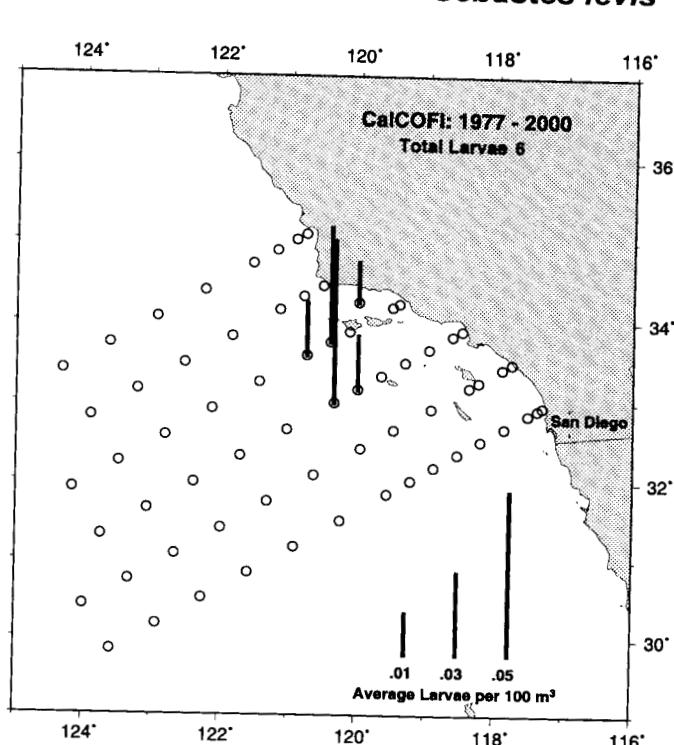
PLEURONECTIDAE



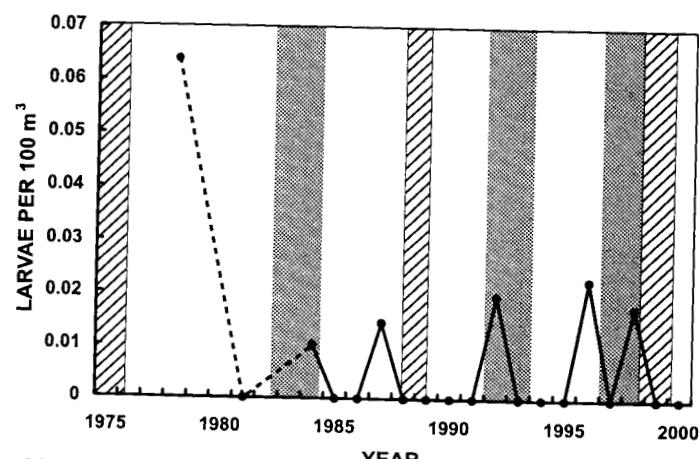
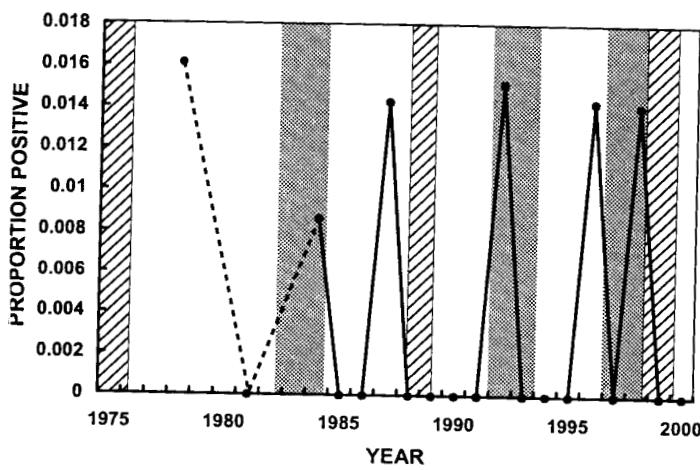
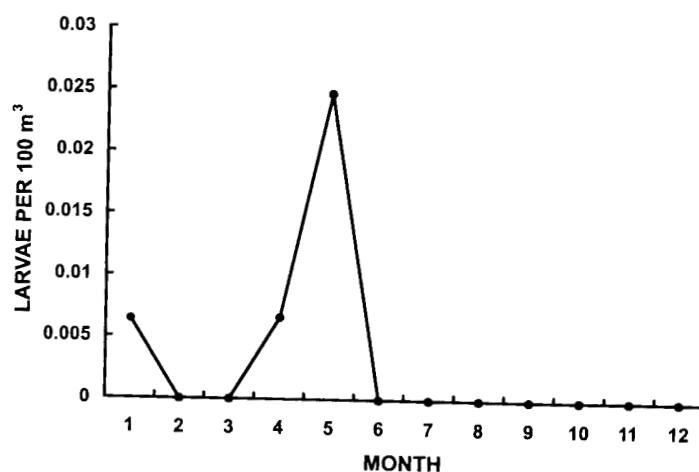
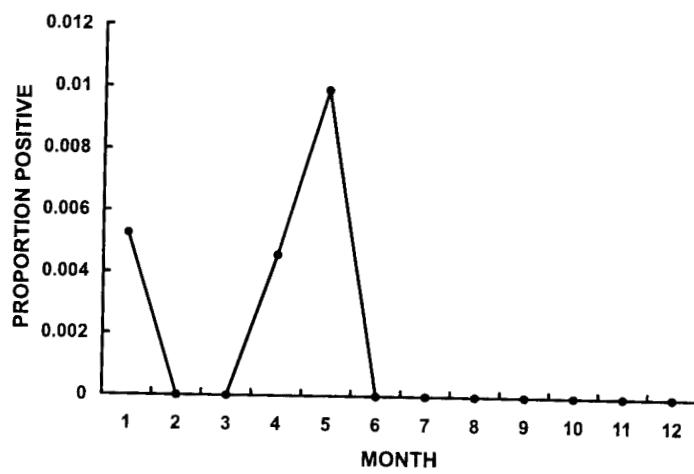
SEBASTIDAE



Cowcod



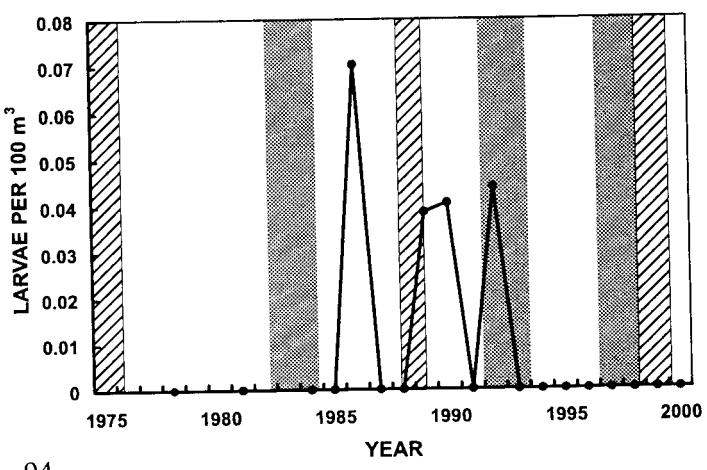
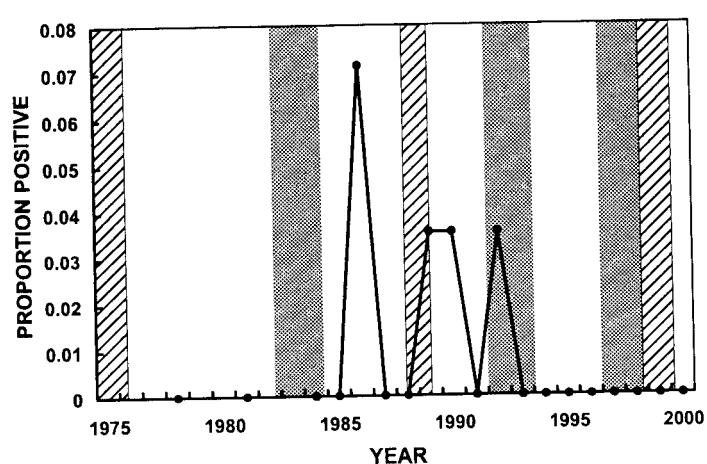
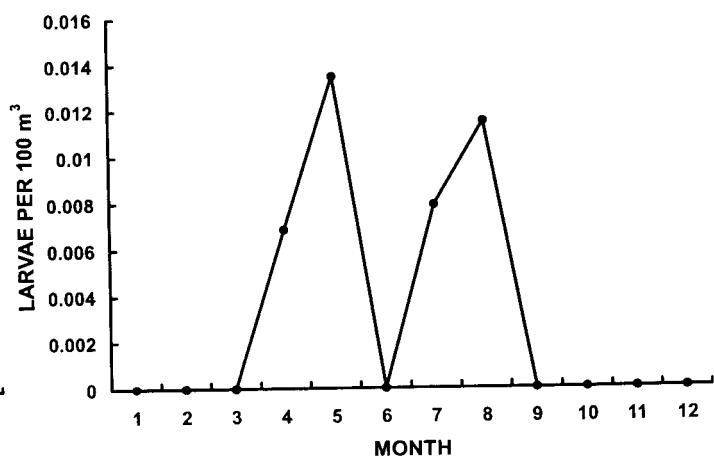
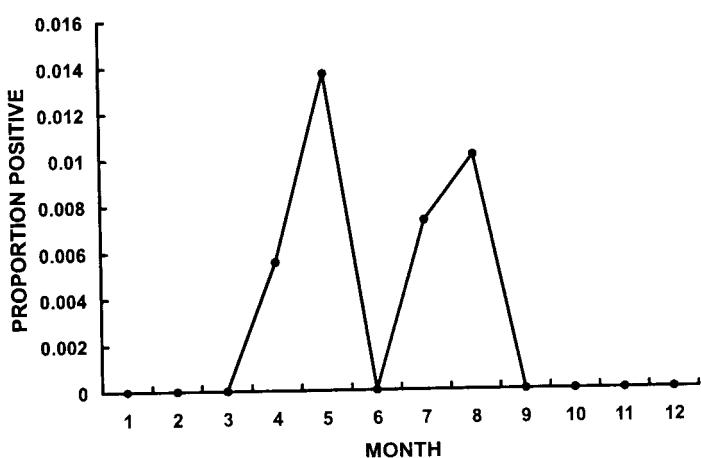
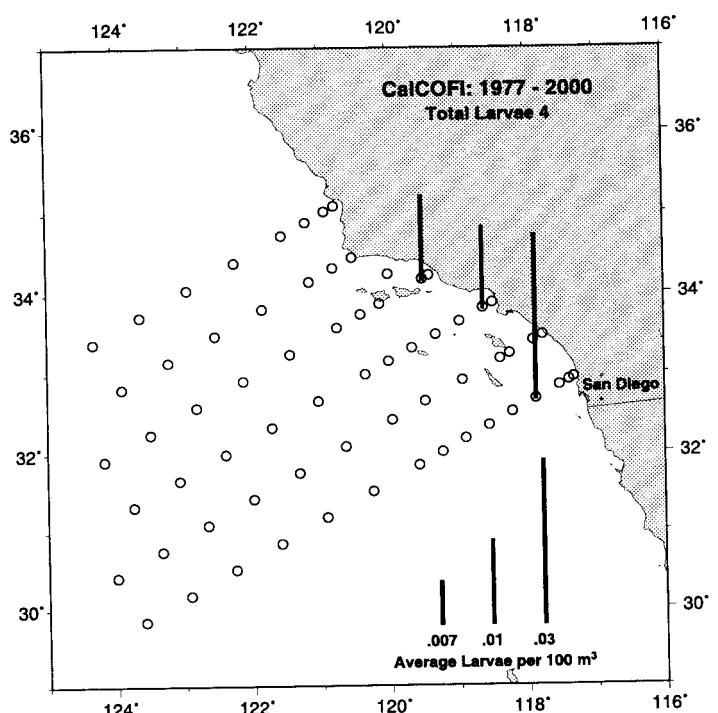
Sebastodes levis



Sarda chiliensis

Pacific bonito

SCOMBRIDAE



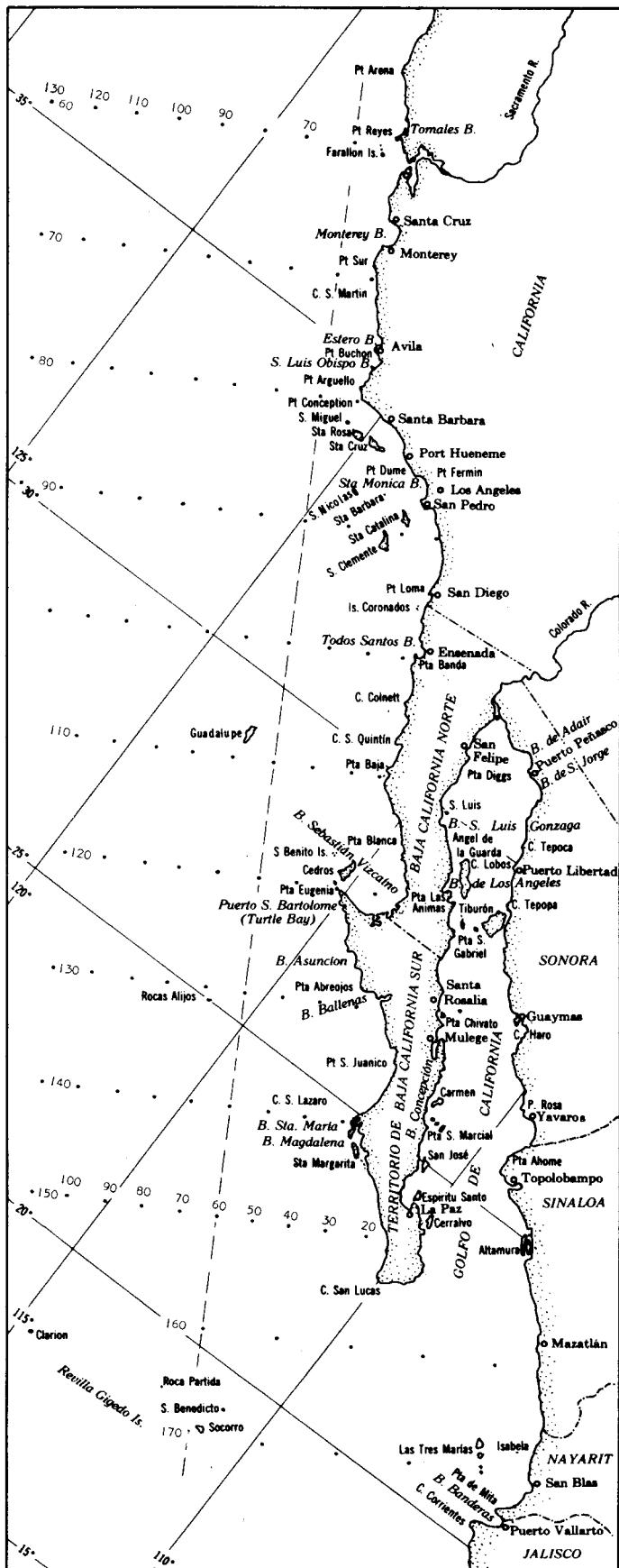
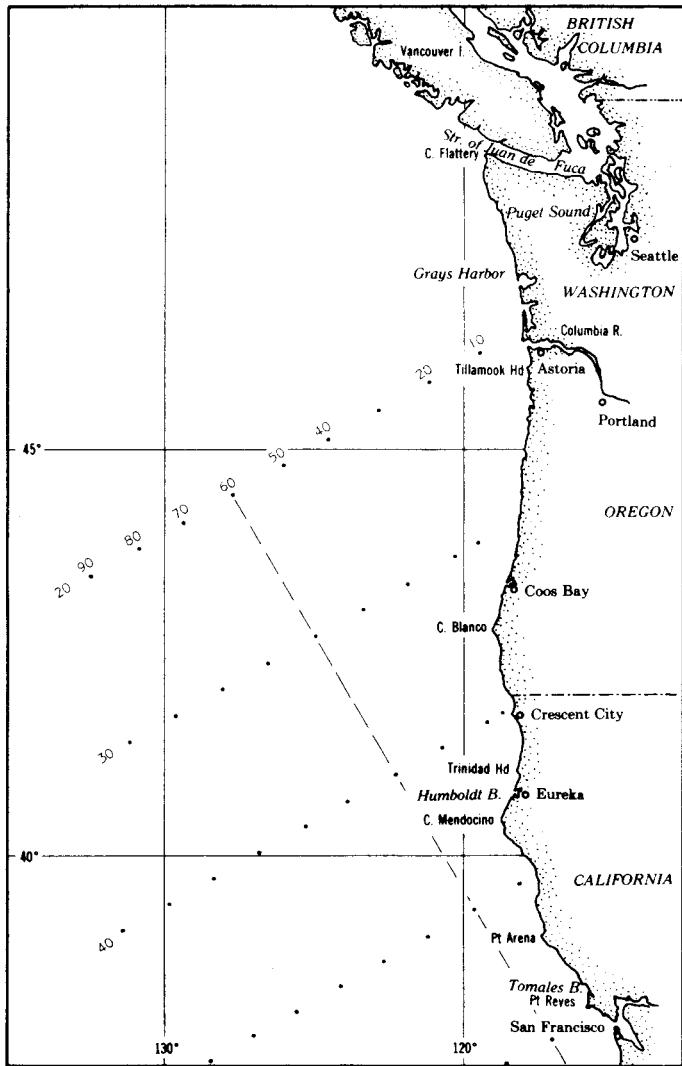
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<i>Pleuronichthys verticalis</i>	48
Total fish larvae	2
Total fish eggs	3



These maps are designed to show essential details of the area most intensively studied by the California Cooperative Oceanic Fisheries Investigations. This is approximately the same area as is shown in color on the front cover. Geographical place names are those most commonly used in the various publications emerging from the research. The cardinal station lines extending southwestward from the coast are shown. They are 120 miles apart. Additional lines are utilized as needed and can be as closely spaced as 12 miles apart and still have individual numbers. The stations along the lines are numbered with respect to the station 60 line, the numbers increasing to the west and decreasing to the east. Most of them are 40 miles apart, and are numbered in groups of 10. This permits adding stations as close as 4 miles apart as needed. An example of the usual identification is 120.65. This station is on line 120, 20 nautical miles southwest of station 60.

The projection of the front cover is Lambert's Azimuthal Equal Area Projection. The detail maps are a Mercator projection.

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